

Unclassified

ENV/JM/MONO(2012)41

Organisation de Coopération et de Développement Économiques
Organisation for Economic Co-operation and Development

18-Dec-2012

English - Or. English

ENVIRONMENT DIRECTORATE
JOINT MEETING OF THE CHEMICALS COMMITTEE AND
THE WORKING PARTY ON CHEMICALS, PESTICIDES AND BIOTECHNOLOGY

CONSENSUS DOCUMENT ON THE BIOLOGY OF THE BRASSICA CROPS (Brassica spp.)

Series on Harmonisation of Regulatory Oversight in Biotechnology
No. 54

JT03332813

Complete document available on OLIS in its original format

This document and any map included herein are without prejudice to the status of or sovereignty over any territory, to the delimitation of international frontiers and boundaries and to the name of any territory, city or area.



ENV/JM/MONO(2012)41
Unclassified

English - Or. English

Also published in the Series on Harmonisation of Regulatory Oversight in Biotechnology:

- No. 1, Commercialisation of Agricultural Products Derived through Modern Biotechnology: Survey Results (1995)
- No. 2, Analysis of Information Elements Used in the Assessment of Certain Products of Modern Biotechnology (1995)
- No. 3, Report of the OECD Workshop on the Commercialisation of Agricultural Products Derived through Modern Biotechnology (1995)
- No. 4, Industrial Products of Modern Biotechnology Intended for Release to the Environment: The Proceedings of the Fribourg Workshop (1996)
- No. 5, Consensus Document on General Information concerning the Biosafety of Crop Plants Made Virus Resistant through Coat Protein Gene-Mediated Protection (1996)
- No. 6, Consensus Document on Information Used in the Assessment of Environmental Applications Involving *Pseudomonas* (1997)
- No. 7, Consensus Document on the Biology of *Brassica napus* L. (Oilseed Rape) (1997)
- No. 8, Consensus Document on the Biology of *Solanum tuberosum* subsp. *tuberosum* (Potato) (1997)
- No. 9, Consensus Document on the Biology of *Triticum aestivum* (Bread Wheat) (1999)
- No. 10, Consensus Document on General Information Concerning the Genes and Their Enzymes that Confer Tolerance to Glyphosate Herbicide (1999)
- No. 11, Consensus Document on General Information Concerning the Genes and Their Enzymes that Confer Tolerance to Phosphinothricin Herbicide (1999)
- No. 12, Consensus Document on the Biology of *Picea abies* (L.) Karst (Norway Spruce) (1999)
- No. 13, Consensus Document on the Biology of *Picea glauca* (Moench) Voss (White Spruce) (1999)
- No. 14, Consensus Document on the Biology of *Oryza sativa* (Rice) (1999)
- No. 15, Consensus Document on the Biology of *Glycine max* (L.) Merr. (Soybean) (2000)
- No. 16, Consensus Document on the Biology of *Populus* L. (Poplars) (2000)
- No. 17, Report of the OECD Workshop on Unique Identification Systems for Transgenic Plants, Charmey, Switzerland, 2-4 October 2000 (2001)
- No. 18, Consensus Document on the Biology of *Beta vulgaris* L. (Sugar Beet) (2001)
- No. 19, Report of the Workshop on the Environmental Considerations of Genetically Modified Trees, Norway, September 1999 (2001)
- No. 20, Consensus Document on Information Used in the Assessment of Environmental Applications Involving Baculoviruses (2002)
- No. 21, Consensus Document on the Biology of *Picea sitchensis* (Bong.) Carr. (Sitka Spruce) (2002)
- No. 22, Consensus Document on the Biology of *Pinus strobus* L. (Eastern White Pine) (2002)
- No. 23, Revised 2006: OECD Guidance for the Designation of a Unique Identifier for Transgenic Plants (2006)
- No. 24, Consensus Document on the Biology of *Prunus* spp. (Stone Fruits) (2002)
- No. 25, Module II: Herbicide Biochemistry, Herbicide Metabolism and the Residues in Glufosinate-Ammonium (Phosphinothricin)-Tolerant Transgenic Plants (2002)
- No. 26, Output on the Questionnaire on National Approaches to Monitoring/Detection/Identification of Transgenic Products (2003)
- No. 27, Consensus Document on the Biology of *Zea mays* subsp. *mays* (Maize) (2003)
- No. 28, Consensus Document on the Biology of European White Birch (*Betula pendula* Roth) (2003)
- No. 29, Guidance Document on the Use of Taxonomy in Risk Assessment of Micro-organisms: Bacteria (2003)

- No. 30, Guidance Document on Methods for Detection of Micro-organisms Introduced into the Environment: Bacteria (2004)
- No. 31, Consensus Document on the Biology of *Helianthus annuus* L. (Sunflower) (2004)
- No. 32, An Introduction to the Biosafety Consensus Documents of OECD's Working Group for Harmonisation in Biotechnology (2005)
- No. 33, Consensus Document on the Biology of Papaya (*Carica papaya*) (2005)
- No. 34, Consensus Document on the Biology of *Pleurotus* spp. (Oyster Mushroom) (2005)
- No. 35, Points to Consider for Consensus Documents on the Biology of Cultivated Plants (2006)
- No. 36, Consensus Document on the Biology of *Capsicum annum* Complex (Chili peppers, Hot peppers and Sweet peppers) (2006)
- No. 37, Consensus Document on Information Used in the Assessment of Environmental Application involving *Acidithiobacillus* (2006)
- No. 38, Consensus Document on the Biology of Western White Pine (*Pinus monticola* Dougl. ex D. Don) (2008)
- No. 39, Abstracts of the OECD Expert Workshop on the Biology of Atlantic Salmon (2006)
- No. 40, Consensus Document on the Biology of *Pinus banksiana* (Jack Pine) (2006)
- No. 41, Consensus Document on the Biology of the Native North American Larches: Subalpine Larch (*Larix lyallii*), Western Larch (*Larix occidentalis*), and Tamarack (*Larix laricina*) (2007)
- No. 42, Consensus Document on the Safety Information on Transgenic Plants Expressing *Bacillus thuringiensis* – Derived Insect Control Protein (2007)
- No. 43, Consensus Document on the Biology of Douglas-Fir (*Pseudotsuga Menziesii* (Mirb.) Franco) (2008)
- No. 44, Consensus Document on the Biology of Lodgepole Pine (*Pinus contorta* Dougl. ex Loud.) (2008)
- No. 45, Consensus Document on the Biology of Cotton (*Gossypium* spp.) (2008)
- No. 46, Consensus Document on Information Used in the Assessment of Environmental Applications Involving *Acinetobacter* (2008)
- No. 47, Guide for Preparation of Biology Consensus Documents (2008)
- No. 48, Consensus Document on the Biology of Bananas and Plantains (*Musa* spp.) (2009)
- No. 49, Consensus Document on the Biology of *Picea mariana* [Mill.] B.S.P. (Black spruce) (2010)
- No. 50, Guidance Document on Horizontal Gene Transfer between Bacteria (2010)
- No. 51, Consensus Document on Molecular Characterisation of Plants Derived from Modern Biotechnology (2010)
- No. 52, Guidance Document on the Use of Information on Pathogenicity Factors in Assessing the Potential Adverse Health Effects of Micro Organisms: Bacteria (2011)
- No. 53, Consensus Document on the Biology of *Cucurbita* L. (Squashes, Pumpkins, Zucchini and Gourds) (2012)

© OECD 2012

Applications for permission to reproduce or translate all or part of this material should be made to: RIGHTS@oecd.org, Head of Publications Service, OECD, 2 rue André-Pascal, 75775 Paris Cedex 16, France.

OECD Environment, Health and Safety Publications

Series on Harmonisation of Regulatory Oversight in Biotechnology

No. 54

**Consensus Document on the Biology of the *Brassica* Crops
(*Brassica* spp.)**

Environment Directorate

ORGANISATION FOR ECONOMIC CO-OPERATION AND DEVELOPMENT

Paris 2012

ABOUT THE OECD

The Organisation for Economic Co-operation and Development (OECD) is an intergovernmental organisation in which representatives of 34 industrialised countries in North and South America, Europe and the Asia and Pacific region, as well as the European Commission, meet to co-ordinate and harmonise policies, discuss issues of mutual concern, and work together to respond to international problems. Most of the OECD's work is carried out by more than 200 specialised committees and working groups composed of member country delegates. Observers from several countries with special status at the OECD, and from interested international organisations, attend many of the OECD's workshops and other meetings. Committees and working groups are served by the OECD Secretariat, located in Paris, France, which is organised into directorates and divisions.

The Environment, Health and Safety Division publishes free-of-charge documents in eleven different series: **Testing and Assessment; Good Laboratory Practice and Compliance Monitoring; Pesticides; Biocides; Risk Management; Harmonisation of Regulatory Oversight in Biotechnology; Safety of Novel Foods and Feeds; Chemical Accidents; Pollutant Release and Transfer Registers; Emission Scenario Documents;** and **Safety of Manufactured Nanomaterials**. More information about the Environment, Health and Safety Programme and EHS publications is available on the OECD's World Wide Web site (<http://www.oecd.org/ehs/>).

This publication is available electronically, at no charge.

For the complete text of this and many other Biosafety publications, consult the OECD's World Wide Web site (www.oecd.org/biotrack/)

or contact:

**OECD Environment Directorate,
Environment, Health and Safety Division
2 rue André-Pascal
75775 Paris Cedex 16
France**

**Fax: (33-1) 44 30 61 80
E-mail: ehscont@oecd.org**

FOREWORD

Consensus Documents contain information for use during the regulatory assessment of a particular product. In the area of plant biosafety, these are being published on information on the biology of certain plant species, selected traits that may be introduced into plant species, and biosafety issues arising from certain general types of modifications made to plants.

This document addresses the biology of the *Brassica* crops (*Brassica* spp.). It updates and completes the previous document on *Brassica napus* (Series on Harmonisation of Regulatory Oversight in Biotechnology No. 7, 1997) which is therefore cancelled.

Canada served as the lead country in the preparation for the document, and the draft has been revised on a number of occasions based on the input from other member countries and stakeholders.

This document is published under the responsibility of the Joint Meeting of the Chemicals Committee and the Working Party on Chemicals, Pesticides and Biotechnology.

Acknowledgements:

The OECD gratefully acknowledges the contribution of Dr. R.K. Downey (Canada), the primary author, without whom this document could not have been written.

Note: The OECD also deals with the safety of foods and feeds derived from modern biotechnology ("novel" foods and feeds). Regarding some of the *Brassica* crops, the Revised Consensus Document on Compositional Considerations for New Varieties of Low Erucic Acid Rapeseed (Canola) was published in 2011 and is available at www.oecd.org/biotrack/.

TABLE OF CONTENTS

PREAMBLE	12
INTRODUCTION	13
SECTION I – SPECIES OR TAXONOMIC GROUP	14
1.1 Classification and Nomenclature	14
1.2 Description	19
1.2.1 <i>Brassica nigra</i>	19
1.2.2 <i>Brassica rapa</i>	20
1.2.3 <i>Brassica oleracea</i>	24
1.2.4 <i>Brassica napus</i>	28
1.2.5 <i>Brassica juncea</i>	29
1.3 Geographic Distribution, Ecosystems and Habitats, Cultivation and Management Practices, Centres of Origin and Diversity	31
1.3.1 Introduction	31
1.3.2 Geographic distribution	32
1.3.3 Ecosystems and habitats where the species occurs natively and has naturalised	34
1.3.4 Production and agronomy of <i>Brassica</i> oilseed crops	37
1.3.5 <i>Brassica</i> vegetable seed production locations and management	48
1.3.6 Centres of origin and ancestors	50
SECTION II – REPRODUCTIVE BIOLOGY	58
2.1 Generation Time and Duration under Natural and Managed Conditions	58
2.2 Reproduction	58
2.2.1 Floral biology	58
2.2.2 Pollination, pollen dispersal and viability	59
2.2.3 Outcrossing in the field	61
2.2.4 Seed development, production, and natural dispersal	65
2.2.5 Seed viability, longevity and dormancy, germination, seedling establishment	70
SECTION III – GENETICS	73
3.1 Relevant Detailed Genetic Information	73
3.1.1 Cytology	73
3.1.2 Nuclear genome size	75
3.1.3 Possible extent of repetitive or non-coding DNA sequences	75
3.1.4 Main genetic diversity or variability	75
3.1.5 Maternal and/or paternal inheritance of organelle genomes	76
3.1.6 Self incompatibility, ‘S’ alleles	76
SECTION IV – INTERSPECIFIC HYBRIDISATION AND INTROGRESSION	79

4.1	Introduction.....	79
4.1.1	<i>B. napus</i> – <i>Raphanus raphanistrum</i>	83
4.1.2	<i>B. napus</i> – <i>B. rapa</i>	84
4.1.3	<i>B. napus</i> – <i>Hirschfeldia incana</i>	85
4.1.4	<i>B. napus</i> – <i>B. juncea</i>	85
4.1.5	<i>B. napus</i> – <i>Sinapis arvensis</i>	86
4.1.6	<i>B. napus</i> – <i>Raphanus sativus</i>	86
4.1.7	<i>B. napus</i> – <i>Erucastrum gallicum</i>	87
4.1.8	<i>B. napus</i> – <i>B. nigra</i>	87
4.1.9	<i>B. napus</i> – <i>B. oleracea</i> and <i>Brassica</i> vegetables.....	87
4.1.10	<i>B. napus</i> – <i>Sinapis alba</i>	88
4.1.11	<i>B. napus</i> – Other weedy species.....	88
SECTION V – ECOLOGY.....		89
5.1	Interactions in Natural and Agricultural Ecosystems.....	89
5.1.1	Glucosinolates and their ecological interaction.....	89
5.1.2	Damaging insects.....	90
5.2	Beneficial Insects.....	92
5.3	Animal Interaction.....	92
5.4	Soil Microbial Interaction.....	92
5.5	Allelopathy.....	92
5.6	Pathogens.....	93
SECTION VI – BREEDING IMPROVED VARIETIES.....		94
6.1	Introduction.....	94
6.1.1	Breeding methods.....	96
6.2	Improvement through "Interspecific Hybrids" and "Cybrids".....	101
6.3	Biotechnology in <i>Brassica</i> Breeding.....	105
6.3.1	Introduction.....	105
6.3.2	Doubled Haploid breeding.....	105
6.3.3	Molecular markers and their application.....	106
6.3.4	Comparative genomic gene identification.....	106
6.3.5	TILLING technique.....	106
6.3.6	Gene transfer.....	107
REFERENCES.....		108
APPENDIX – COMMON PATHOGENS AND PESTS.....		137

Tables

Table 1.	World production of edible vegetable oils, averages 1996-2000 to 2006-2010.....	13
Table 2.	Nomenclature and genome relationships of cultivated <i>Brassica</i> species and related genera.....	15
Table 3.	List of 43 diploid cytodesmes and 6 amphidiploid taxa in the <i>Brassica</i> coenospecies	18
Table 4.	Area harvested, production and yield by major <i>Brassica</i> oilseed producing countries, averages 1995-1999 to 2005-2009.....	37
Table 5.	Number of herbicide products available for control of volunteer <i>B. napus</i> with nil, single, or multiple herbicide tolerances in western Canada	46
Table 6.	Short distance pollen mediated gene flow from <i>B. napus</i> pollen donor to recipient field/plots in the UK, USA and Canada.....	62
Table 7.	<i>B. napus</i> to <i>B. napus</i> outcrossing rates, by isolation distances, reported from small plot trials and/or large fields	63
Table 8.	Mean outcross percentages of pollen donor to <i>B. napus</i> recipient populations for various isolation distances and two design classes.....	64
Table 9.	Predicted outcrossing rates for spring and winter oilseed rape at three isolation distances (with 95% confidence limits), based on 2000-2003 multilocation UK field trials.....	65
Table 10.	Typical seed weight ranges (or averages) of <i>Brassica</i> crop plants by species and form	66
Table 11.	Estimated dispersal distances of spring <i>B. napus</i> seed released from transport vehicles at various heights above adjacent fields.....	69
Table 12.	Ploidy level, chromosome number, genome size and map length of <i>A. thaliana</i> and <i>Brassica</i> species of "Triangle of U"	75
Table 13.	Interspecific and intergeneric sexual crossing attempts, degree of success and potential for gene introgression	81
Table 14.	Fatty acid composition of rapeseed, canola, soybean, sunflower and linseed oils.....	95
Table 15.	Fatty acid composition of canola and specialty <i>B. napus</i> varieties grown in Canada.....	96
Table 16.	Introgression of nuclear genes conferring desirable traits to <i>Brassica</i> crops.....	103
Table 17.	Intertribal somatic hybrids in Brassiceae for the integration and incorporation of desirable traits into <i>Brassica</i> crops	104
Table 18.	Minimum population size required to select least frequent homozygote at 95% probability	105
Table A.1.	Insect, mite and other Brassicaceous crop pests and their regional distribution.....	137
Table A.2.	Diseases of Rapeseed = Canola [<i>B. napus</i> L. and <i>Brassica rapa</i> L. (= <i>B. campestris</i> L.)].....	140

Figures

Figure 1.	Genome relationships of <i>Brassica</i> species and allied genera.....	17
Figure 2.	Illustration of a <i>Brassica nigra</i> plant and its parts	20
Figure 3.	a) <i>B. rapa</i> subsp. <i>pekinensis</i> var. <i>cylindrica</i>	21
	b) <i>B. rapa</i> subsp. <i>pekinensis</i> var. <i>cephalata</i>	21
Figure 4.	<i>B. rapa</i> subsp. <i>chinensis</i> , Bok choy	22
Figure 5.	<i>B. rapa</i> subsp. <i>chinensis</i> , Baby or Shanghai bok choy	22
Figure 6.	<i>B. rapa</i> subsp. <i>parachinensis</i> , Gai Lan or Chinese broccoli	22
Figure 7.	<i>B. rapa</i> subsp. <i>nipposinica</i>	22
Figure 8.	<i>B. rapa</i> subsp. <i>rapa</i> the common turnip.....	22
Figure 9.	Growth stages in turnip rape (<i>B. rapa</i>)	23
Figure 10.	Typical intact and opened siliques of <i>B. napus</i> and <i>B. rapa</i>	24
Figure 11.	Wild <i>B. oleracea</i> plants in their first year of growth	25
Figure 12.	Heads of <i>B. oleracea</i> var. <i>capitata</i> and Savoy cabbages	25
Figure 13.	Head of cauliflower and broccoli <i>B. oleracea</i> var. <i>botrytis</i>	26
Figure 14.	Sprouting purple broccoli	26
Figure 15.	Romanesco broccoli.....	26
Figure 16.	<i>B. oleracea</i> var. <i>viridis</i> , collard plant and row of kale	27
Figure 17.	<i>B. oleracea</i> var. <i>gemmifera</i> , Brussels sprouts.....	27
Figure 18.	<i>B. napus</i> var. <i>napobrassica</i> , Rutabaga or Swede.....	28
Figure 19.	<i>B. napus</i> var. <i>pabularia</i> , Siberian or rape kale.....	28
Figure 20.	Growth stages of <i>B. napus</i> var. <i>napus</i> f. <i>annua</i>	29
Figure 21.	Upper leaves of <i>B. rapa</i> , <i>B. napus</i> and <i>B. juncea</i>	29

Figure 22.	Three forms of <i>B. juncea</i> with a) bulbous root, b) normal or ‘Ordinal’ root, and c) tuberous basal stem	30
Figure 23.	<i>B. juncea</i> subsp. <i>integrifolia</i> , Heading mustard, BauSin	30
Figure 24.	The world distribution of <i>B. rapa</i> as a reported weed.....	33
Figure 25.	Approximate areas of the phytogeographic regions containing the world’s greatest representation of Brassicaceae genera	35
Figure 26.	Areas of oilseed rape/canola production in North America	38
Figure 27.	Percentage of the total Canadian <i>B. napus</i> production area sown to herbicide-resistance varieties	39
Figure 28.	Oilseed rape (<i>B. napus</i>) production regions in Europe showing millions of hectares of winter rape per country	41
Figure 29.	Areas and concentration of <i>B. napus</i> production by Australian government districts	42
Figure 30.	Major production region of oilseed mustard (<i>B. juncea</i>) and toria (<i>B. rapa</i>) on the Indian sub-continent.....	43
Figure 31.	<i>Brassica</i> oilseeds average yield (kg/ha) and total production (metric tons) by province in China	44
Figure 32.	Regions sown to the spring and winter forms of <i>B. napus</i> in China	44
Figure 33.	Phylogenetic tree for the subtribe Brassicinae, based on PAUP analyses of the chloroplast DNA restriction site/length mutations shared by two or more taxa/accessions.....	51
Figure 34.	Evolutionary geography of <i>B. juncea</i> , <i>B. carinata</i> and <i>Sinapis alba</i>	52
Figure 35.	Distribution of wild ‘species’ of <i>B. oleracea</i>	54
Figure 36.	Dispersal of the <i>B. napus</i> species from a proposed centre of origin	56
Figure 37.	Typical flower of <i>B. napus</i>	59
Figure 38.	Outcrossing percentages as affected by distance from the pollen source	61
Figure 39.	Development stages of the <i>Brassica napus</i> zygotic embryo.....	65
Figure 40.	Distinguishing <i>Brassica</i> species by their seed coat characteristics	67
Figure 41.	Illustration of major events in the evolution of selected <i>Brassica</i> species and <i>Arabidopsis thaliana</i>	74
Figure 42.	Glucosinolate chemical structure and enzymatic breakdown products formed in broken <i>Brassica</i> plant cells with moisture	90
Figure 43.	Canola oil compared to other edible vegetable oils as to total saturated fat content and other fatty acids	95
Figure 44.	Breeding scheme combining the backcross and pedigree selection systems to develop a low erucic, low glucosinolate variety with high seed and oil yield	98
Figure 45.	Breeding scheme for development of commercial synthetic varieties of oilseed <i>Brassica</i> crops.....	99
Figure 46.	Self-incompatibility (SI) scheme for breeding cabbage hybrid seed production	100
Figure 47.	Production system for Cytoplasmic Male Sterile (CMS) hybrid seed production in oilseed rape.....	101

PREAMBLE

Transgenic crops such as maize, soybean, rapeseed, and cotton have been assessed and approved for commercial use in an increasing number of countries. Crop species that have not yet been commercialized are also in development, and either will be or are being assessed for environmental risk/safety. Recognizing the possibility of a wide application of modern biotechnology to a variety of crops and the international nature of trade in crop agriculture, the OECD's Working Group on Harmonization of Regulatory Oversight in Biotechnology decided at its first session, in June 1995, to focus its work on identifying the information which could be commonly used in countries for environmental risk/safety assessment of transgenic crops. It was hoped this effort would encourage information sharing and avoid duplication of effort among countries.

Since 1995, the Working Group has developed documents addressing key considerations in environmental risk/safety assessment of transgenic organisms. Such documents are called *Biosafety Consensus Documents*. The environmental risk/safety assessment of transgenic organisms are normally based on information on the characteristics of the organism to be modified, the introduced trait(s), the environment into which the engineered organism is introduced, the interaction between these, and the intended application. A *Biosafety Consensus Document* generally addresses one of these considerations.

Biosafety Consensus Documents are intended to be a "snapshot" of current information on a specific host organism or trait, for use during regulatory assessments. They are not intended to be a comprehensive source of information on everything that is known about a specific organism or trait; but they do address the key or core set of issues that member countries believe are relevant to risk/safety assessment. This core set of information is contained in the *Biosafety Consensus Document* and is said to be mutually acceptable among member countries. To date, 45 *Biosafety Consensus Documents* have been developed by the Working Group and published. This number includes documents which address the biology of crops, trees and microorganisms as well as those which address specific traits which are used in transgenic crops.

This *Biosafety Consensus Document* on the biology of the cultivated species of the *Cucurbita* genus is intended to supply basic information, useful in environmental risk/safety assessment. In using the *Biosafety Consensus Documents*, it is useful to consult two additional texts. The first, entitled *An Introduction to the Biosafety Consensus Documents of OECD's Working Group for Harmonization in Biotechnology* explains the purpose of the *Biosafety Consensus Documents* and how they are relevant to risk/safety assessment. The second text is *Points to Consider for Consensus Documents on the Biology of Cultivated Plants*. This document is a structured checklist of "points to consider" for authors when drafting, or for those reviewing, a Consensus Document on the biology of a cultivated plant. Amongst other things, this text describes how each point is relevant to risk/safety assessment.

The *Biosafety Consensus Documents* are of value to applicants for commercial uses of transgenic organisms, regulators in national authorities and to the wider scientific community. As each of the documents may be updated in the future as new knowledge becomes available, users of Consensus Documents are encouraged to provide any information or opinions regarding the contents of this document or indeed, OECD's other harmonisation activities.

INTRODUCTION

1. The plants within the family Brassicaceae constitute one of the world's most economically important plant groups. They range from noxious weeds to leaf and root vegetables to oilseed and condiment crops. The cole vegetables are perhaps the best known group. Indeed the *Brassica* vegetables are a dietary staple in every part of the world with the possible exception of the tropics. FAO estimates that world commercial production of cabbages, cauliflowers, broccoli and other *Brassica* vegetables in 2009 was over 84 million tonnes from some 3.5 million hectares, with a 2009 farm gate value of some 25 billion US dollars (FAOSTAT, 2009). These figures do not include the root vegetables or the production from kitchen gardens.

2. Less well known are the *Brassica* oilseed crops that annually occupy over 26 million hectares of the world's agricultural lands. Because of their ability to survive and grow at relatively low temperatures, they are one of the few edible oil sources that can be successfully produced in cool temperate regions. This characteristic makes them well adapted to cultivation at high elevations and as winter crops in the subtropics. In temperate regions, oilseed rape (*Brassica napus*¹) and turnip rape (*Brassica rapa*) predominate, while in the subtropics of Asia, Indian mustard or rai (*Brassica juncea*) is the major oil source. Among all the commodities moving in world trade, only petroleum has a greater value than vegetable oils (U.S. Census Bureau, Foreign Trade Statistics; UN Statistics Division). In total, the *Brassica* oilseeds provide 14% of the world's edible vegetable oil and are the third most important source of edible oil after soybean and palm (Table 1).

Table 1. World production of edible vegetable oils, averages 1996-2000 to 2006-2010
(in millions of tonnes)

Crop	1996-2000 (MMt)	2001-05 (MMt)	2006-10 (MMt)
Soybean	22.4	29.8	36.8
Palm	18.3	28.2	40.8
Rape/mustard	11.8	13.7	19.1
Sunflower	8.9	8.4	11.0
Groundnut	4.3	4.9	4.8
Cottonseed	3.7	4.1	4.9
Others ¹	7.9	9.5	11.2
Total	77.3	98.6	128.6

¹ Others include olive, coconut and palm kernel

Source: after USDA-Foreign Agricultural Service

¹ The authority for the scientific names used in this manuscript are given in Tables 2 and 3. The nomenclatural authority for genus and species names not listed in the tables will be included in the text where they first appear.

SECTION I – SPECIES OR TAXONOMIC GROUP

1.1 Classification and Nomenclature

3. The family Brassicaceae (= Cruciferae) contains over 338 genera and 3709 species (Al-Shehbaz *et al.*, 2006; Warwick *et al.*, 2006). The species of greatest interest to those concerned with genetically modified crops are given in Table 2 with their chromosome number, genome identification and common English name(s). Early humans recognised the edible value of many of these species and through selection modified nearly every plant part to suit their needs. Such modifications include the compacting of the leaves to form a head, the root or stem to form a bulb, the inflorescence to form a curd or bunch and the seed to provide both oil and condiment. Species grown as oilseeds include *B. napus*, *B. juncea*, *B. rapa* and *B. carinata*. The vegetable Brassicaceae include *B. napus* (rutabaga, Siberian kale), *B. rapa* (Chinese cabbage, bok choy, pai-tsai, mizuna, Chinese mustard, broccoli raab and turnip), *B. oleracea* (cabbage, broccoli, cauliflower, Brussels sprouts, kohlrabi, collards and kale), *Raphanus sativus* (radish), *Lepidium sativum* (garden cress) and *Nasturtium officinale* (watercress). The condiment crops include *B. juncea* (brown and oriental mustard), *Sinapis alba* (yellow mustard), *B. nigra* (black mustard, but now little used), *Armoracia rusticana* (horseradish) and *Eutrema japonica* (wasabi). There are a number of other minor potherbs and salad vegetables. There are numerous weedy species but those of greatest interest with regard to cross-pollination with *B. napus* are *Sinapis arvensis* (wild mustard or charlock), *Raphanus raphanistrum* (wild radish), *B. rapa* (wild or bird rape) and *Hirschfeldia incana* (hoary mustard).

4. The genus *Brassica* is classified as follows:

Order Brassicales (= Cruciales)
 Family Brassicaceae (= Cruciferae)
 Tribe Brassiceae
 Subtribe Brassicinae
Genus *Brassica* L.

5. The Brassicaceae family is comprised of 25 tribes with about an additional five under study (Al-Shehbaz *et al.*, 2006). The tribe Brassiceae, which contains the genus *Brassica* and its wild relatives, is made up of 48 genera and approximately 240 species (Warwick and Hall, 2009). Warwick *et al.* (2006) have prepared a check list and a current taxonomic database for the family on CD-ROM. Also on CD-ROM are chromosome numbers from the literature for 68.6% of the genera and 42.0% of the Brassicaceae species (Warwick and Al Shehbaz, 2006; Warwick *et al.*, 2009b). The morphological traits that characterise the tribe are conduplicate cotyledons (the radical enclosed by longitudinally folded cotyledons) and/or transversely segmented fruits, that have seeds or rudimentary ovules in both segments and, if present, only simple trichomes or hairs (Warwick and Hall, 2009). Modern molecular studies have reinforced the monophyletic origin of the tribe.

Table 2. Nomenclature and genome relationships of cultivated *Brassica* species and related genera

Species name	Common synonym	Haploid chromosome number & genome	Common name
<i>Brassica rapa</i> L.	<i>B. campestris</i> L.	10 AA	
subsp. <i>campestris</i> (L.) A. R. Clapham			Summer turnip rape, Canola
subsp. <i>oleifera</i> (DC.) Metzg.			Winter turnip rape
subsp. <i>campestris</i> (L.) A. R. Clapham	subsp. <i>eu-campestris</i> (L.) Olsson		Bird or wild turnip rape
subsp. <i>trilocularis</i> (Roxb.) Hanelt	subsp. <i>sarson</i> (Prain) Denford		Yellow & Brown Sarson
subsp. <i>dichotoma</i> (Roxb.) Hanelt			Toria
subsp. <i>chinensis</i> (L.) Hanelt	<i>B. chinensis</i> L. <i>B. chiensis</i> var. <i>parachinsis</i> (L. H. Bailey)		Pak-choi or bok choy, Chinese mustard Chinese broccoli, Gai Lan
subsp. <i>pekinensis</i> (Lour.) Hanelt	<i>B. pekinensis</i> (Lour) Rupr.		Pe-tsai, Chinese cabbage
subsp. <i>nipposinica</i> (L. H. Bailey) Hanelt			Curled mustard
subsp. <i>rapa</i>	<i>B. rapa</i> L.		Turnip
<i>Brassica tournefortii</i> Gouan		10 TT	Wild turnip
<i>Brassica nigra</i> (L.) W.D.J. Koch		8 BB	Black mustard
<i>Brassica oleracea</i> L.		9 CC	
var. <i>viridis</i> L.	var. <i>acephala</i> DC.		Kale, Collard
var. <i>botrytis</i> L.			Cauliflower & Broccoli
var. <i>capitata</i> L.			Cabbage
var. <i>gongylodes</i> L.	var. <i>caulorapa</i> Pasq.		Kohlrabi
var. <i>gemmifera</i> (DC.) Zenker			Brussels sprouts
var. <i>italica</i> Plenck.			Broccoli
var. <i>oleracea</i>	subsp. <i>sylvestris</i> (L.) Miller		Wild cabbage
subsp. <i>alboglabra</i> L. H. Bailey	<i>B. alboglabra</i> L. H. Bailey		Chinese kale, kailan
<i>Brassica juncea</i> (L.) Czern.		18 AABB	Brown and Oriental mustard, Rai
<i>Brassica napus</i> L.		19 AACC	
var. <i>napus</i>	subsp. <i>oleifera</i> (Delile) Sinskaya		Summer oilseed rape, Canola
var. <i>napus</i>	<i>B. napus</i> f. <i>biennis</i> (Schübl. & G. Martens) Thell.		Winter oilseed rape, Winter canola
var. <i>pabularia</i> (DC.) Rchb.			Rape-kale
var. <i>napobrassica</i> (L.) Rchb.	subsp. <i>rapifera</i> (Metzg.) Sinskaya		Rutabaga, swede
<i>Brassica carinata</i> A. Braun.		17 BBCC	Abyssinian mustard
<i>Hirschfeldia incana</i> (L.) Lagr.-Foss.	<i>Brassica adpressa</i> Boiss.	7 HH	Hoary mustard
<i>Sinapis arvensis</i> L.	<i>B. kaber</i> (DC.) L. C. Wheeler	9 SarSar	Wild mustard, charlock
<i>Sinapis alba</i> L.	<i>B. hirta</i> Moench	12 SalSal	Yellow or White mustard
<i>Raphanus sativus</i> L.		9 RR	Radish
<i>Raphanus raphanistrum</i> L.		9 RR	Wild radish
<i>Diplotaxis muralis</i> (L.) DC.		21 DD	Annual wall-rocket
<i>Erucastrum gallicum</i> (Willd.) O.E. Schulz		15	Dog mustard
<i>Eruca vesicaria</i> (L.) Cav. subsp. <i>sativa</i> (Mill.) Thell.	<i>Eruca sativa</i> Mill.	11 EE	Rocket salad

Source: Table modified from Yarnell (1956)

6. Taxonomic research on the tribe conducted by Schulz (1919, 1936) established the basic classification which we follow today, although it has been modified and criticised (Al-Shehbaz, 1984). Within the tribe, Schulz (1919, 1936) recognised 10 subtribes, with Gómez-Campo (1980) later recommending a reduction to nine. Of the nine subtribes three are of greatest relevance to those concerned with *Brassica* crops: namely; Brassicinae, Moricandiinae and Raphaninae. Within these subtribes *Brassica*, *Sinapis*, *Diploaxis*, *Erucastrum*, *Hirschfeldia*, *Eruca*, and *Raphanus* are of primary interest. The association and relationships among species within these subtribes have been studied cytogenetically, chemically and morphologically (reviewed by Prakash and Hinata, 1980; Takahata and Hinata, 1983, 1986) without providing a clear separation of the subtribes and their genera. Recent molecular, morphological and hybridisation data give strong support for a rearrangement of the three subtribes into two clades, namely, the Rapa/Oleracea and the Nigra lineages (See Section 1.3.6, Centres of origin and ancestors, as well as Warwick and Hall, 2009, and references therein). Such a division is also referred to in some publications as the *Brassica* and *Sinapis* lineages. It is expected that the realignment of the species from the three subtribes into the two clades will eventually require renaming of many of the species involved.

7. The difficulty in clearly separating the genera and species among the *Brassica* and their close relatives has arisen because similar plant forms and morphological traits occur in more than one genus or species. The difficulties encountered by early taxonomists in separating and classifying the various species and forms within the Brassicaceae family are well documented by Hedge (1976) and Prakash and Hinata (1980). As a result there have been numerous changes and modifications to Schulz's (1919, 1936) original species names and arrangement. The cytological studies by Morinaga (1928, 1929, 1931, 1933, 1934a, b) and his student, U (1935) clarified the broad relationships among the economically important *Brassica* species in which chromosome pairing clearly showed the three species with the higher chromosome number, *B. napus*, *B. juncea* and *B. carinata* are amphidiploids derived from the monogenomic or basic species, *B. nigra*, *B. rapa* and *B. oleracea* (Figure 1). The genome relationships among the amphidiploids were confirmed by resynthesis of the three species from their diploid parents (Frandsen, 1943, 1947; Ramanujam and Srinivasachar, 1943). Further verification of these species relationships were obtained from studies on phenolic compounds (Dass and Nybom, 1967), protein patterns (Vaughan, 1977), isozymes (Coulthart and Denford, 1982; Chen *et al.*, 1989) and nuclear DNA, RFLP (Restriction Fragment Length Polymorphisms) (Song *et al.*, 1988a, b). Additional verification has been achieved through molecular analysis of nuclear and chloroplast DNA and fluorescence *in situ* hybridisation (Snowdon *et al.*, 2003; Snowdon, 2007; Warwick and Sauder, 2005; Lysak *et al.*, 2005).

8. To further establish the true relationships among the genus and species of the subtribe, Harberd (1972, 1976) proposed grouping them into 'cytodemes' based on the crossability of related subspecies with the same chromosome number. Harberd (1976) defined cytodemes as follows: "If two populations have a common chromosome number and are easily crossed to form a hybrid, which is neither obviously weak in vigour nor of low fertility, then they belong in the same *cytodeme*. By contrast, different cytodemes (which sometimes have the same chromosome number) are (a) difficult to cross, or (b) give a weak hybrid, or (c) have a sterile hybrid, and frequently exhibit all three criteria." Harberd (1972, 1976) also defined the *Brassica* coenospecies as "the group of wild species sufficiently related to the six cultivated species of *Brassica* to be potentially capable of experimental hybridisation with them". On this basis and their chromosome number the coenospecies have been classified into 43 diploid and 13 tetraploid cytodemes (Table 3 from Warwick and Black, 1993). This grouping, with the inclusion of *Raphanus* and *Enarthrocarpus* in the subtribe, is supported by both chloroplast and nuclear DNA analysis (Warwick and Black, 1993; Warwick and Hall, 2009).

9. Cytological analyses of chromosome pairing in interspecific crosses among some of the more important *Brassica* cytodemes by Mizushima (1980) provided information on the maximum possible

number of autosyndetic¹ chromosome pairs (Figure 1). Harberd and McArthur (1980) extended the study of meiotic chromosome pairing to more than 50 species hybrids. These distant crosses were facilitated using embryo culture.

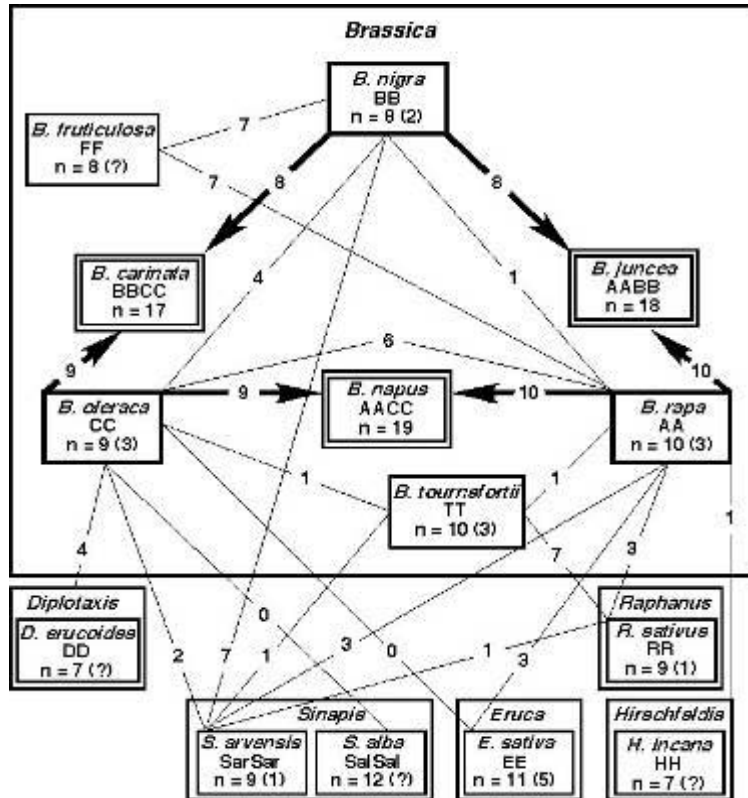


Figure 1. Genome relationships of *Brassica* species and allied genera

Source: modified from Mizushima, 1980

A, B, C... are the genome symbols. The number in brackets following the haploid chromosome number (n) indicates the maximum possible number of autosyndetic¹ chromosome pairs. The numbers within lines connecting two genomes give the maximum allosyndesis, i.e. the number of bivalents possible between the respective interspecific hybrids. (Downey and Röbbelen, 1989).

10. A chromosome analysis of the monogenomic *Brassica* species by Röbbelen (1960), established that only six chromosomes were distinctly different, the remaining being homologous with one or the other of the basic six. This evidence pointed to the presence, in the evolutionary pathway of the *Brassica* species, of a now-extinct, ancient progenitor with a basic chromosome number of $\times = 6$. The long standing hypothesis, that the cultivated diploid *Brassica* species are ancient polyploids, has been strongly supported by modern genomic investigations. The genomes of *Brassica* species are extensively triploid (Lysak *et al.*, 2005, 2007; Rana *et al.*, 2004). In *B. nigra* Lagercrantz and Lydiate (1996) reported that every chromosome region appeared to be present in triplicate and the genomes of *B. oleracea* and *B. rapa* also exhibit tripling (Rana *et al.*, 2004; Mun *et al.*, 2009; Wang, 2010). High density comparative mapping of *Arabidopsis* and *B. napus* also supported the hypothesis of a hexaploid ancestor (Parkin *et al.*, 2005). . Indeed, chromosome tripling has been documented for the entire Brassiceae tribe (Lysak *et al.*, 2005). Linkage maps and genome size data (Lysak *et al.*, 2009) indicate that the *B. oleracea* genome, and probably the other monogenomic species which exhibit a range in chromosome number from 7 to 12,

¹Autosyndesis is defined as the pairing of completely or partially homologous chromosomes during prophase of the first meiotic division.

increased or reduced their chromosome number through duplication, translocations (Quiros *et al.*, 1988; Hosaka *et al.*, 1990; McGrath *et al.*, 1990; Truco and Quiros, 1994), transposition of elements (Zhang and Wessler, 2004) as well as deletions (Hu and Quiros, 1991) and fusions (Lysak *et al.*, 2006).

11. Based on cotyledonary studies of various taxa in the tribe Brassiceae, Gómez-Campo and Tortosa (1974) proposed that *Brassica* evolved from the Macaronesian plant taxon *Sinapidendron*. This Miocene relic survived several paleo-climatic changes that destroyed most of the Mediterranean Tertiary flora and is put forward as the archetype from which *Brassica* evolved through the *Diplotaxis* and *Erucastrum* complexes. However, the use of such morphometric data to establish evolutionary relationships within the Brassiceae has not always provided results that agreed with those from cytological and molecular studies.

Table 3. List of 43 diploid cytodesmes and 6 amphidiploid taxa in the *Brassica* coenospecies

Source: Warwick and Black, 1993

n (haploid chromosome number)	Cytodeme
	<i>Diploids</i>
10	<i>Brassica barrelieri</i> (L.) Janka
7	<i>Brassica deflexa</i> Boiss.
11	<i>Brassica elongata</i> Ehrh.
8	^a <i>Brassica fruticulosa</i> Cirillo (includes <i>B. maurorum</i> Dur., <i>B. spinescens</i> Pomel, <i>Erucastrum littoreum</i> (Pau & Font Quer) Maire subsp. <i>glabrum</i> (Maire) Gómez-Campo (= <i>Erucastrum laevigatum</i> subsp. <i>glabrum</i> Maire)
10	^a <i>Brassica gravinae</i> Ten.
8	<i>Brassica nigra</i> (L.) W.D.J. Koch
9	<i>Brassica oleracea</i> L. (includes <i>B. alboglabra</i> L.H. Bailey, <i>B. bourgeauii</i> (Webb.) Kuntze, <i>B. cretica</i> Lam., <i>B. hilarionis</i> G.E. Post, <i>B. incana</i> Ten., <i>B. insularis</i> Moris, <i>B. macrocarpa</i> Guss., <i>B. montana</i> Pourr., <i>B. rupestris</i> Raf., <i>B. villosa</i> Biv.)
9	<i>Brassica oxyrrhina</i> (Coss.) Willk.
10	<i>Brassica rapa</i> L. (= <i>B. campestris</i> L.) (includes wild and cultivated varieties)
10	<i>Brassica repanda</i> (Willd.) DC. (includes <i>B. desnottesii</i> Emb. & Maire, <i>B. nudicaulis</i> (Lag.) O.E. Schulz, <i>B. saxatilis</i> DC.)
11	<i>Brassica souliei</i> (Batt.) Batt. (= <i>B. amplexicaulis</i> (Desf.) Pomel)
10	<i>Brassica tournefortii</i> Gouan
12	^a <i>Coincya</i> spp. (= <i>Hutera</i> = <i>Rhynchosinapis</i>) (includes all species in the genus)
11	<i>Diplotaxis acris</i> (Forssk.) Boiss.
9	<i>Diplotaxis assurgens</i> (Delile) Gren.
9	<i>Diplotaxis berthautii</i> Braun-Blanq. & Maire
9	<i>Diplotaxis catholica</i> (L.) DC.
7	<i>Diplotaxis cossoniana</i> (Reut.) O.E. Schulz
7	<i>Diplotaxis erucoides</i> (L.) DC.
13	<i>Diplotaxis harra</i> (Forssk.) Boiss. (includes <i>D. crassifolia</i> (Raf.) DC., <i>D. lagascana</i> DC.)
8	<i>Diplotaxis siettiana</i> Maire (includes <i>D. ibicensis</i> (Font Quer) Gómez-Campo)
10	<i>Diplotaxis stifolia</i> Kunze
11	<i>Diplotaxis tenuifolia</i> (L.) DC. (includes <i>D. cretacea</i> Kotov., <i>D. simplex</i> (Viv.) Spreng., the latter species was incorrectly listed as <i>D. pitardiana</i> Maire)
9	<i>Diplotaxis tenuisiliqua</i> Delile
10	<i>Diplotaxis viminea</i> (L.) DC.
9	<i>Diplotaxis virgata</i> (Cav.) DC.
10	<i>Enarthrocarpus</i> spp. (includes <i>E. lyratus</i> (Forssk.) DC., <i>E. pterocarpus</i> (Pers.) DC., <i>E. strangulatus</i> Boiss.)
11	<i>Eruca</i> spp. (includes <i>E. vesicaria</i> (L.) Cav., <i>E. sativa</i> Mill., <i>E. pinnatifida</i> (Desf.) Pomel)
8	^a <i>Erucastrum abyssinicum</i> R.E. Fr.
9	<i>Erucastrum canariense</i> Webb & Berthel. (includes <i>E. cardaminoides</i> (Webb) O.E. Schulz)
8	^a <i>Erucastrum nasturtiifolium</i> (Poir.) O.E. Schulz (includes <i>E. leucanthum</i> Coss. & Dur.)
8	<i>Erucastrum strigosum</i> (Thunb.) O.E. Schulz
7	<i>Erucastrum varium</i> (Durieu) Durieu
7	^a <i>Erucastrum virgatum</i> C. Presl
7	<i>Hirschfeldia incana</i> (L.) Lagr.-Foss.

9	<i>Raphanus</i> spp. (includes <i>R. raphanistrum</i> L., <i>R. sativus</i> L., <i>R. caudatus</i> L., <i>R. maritimus</i> Smith, and <i>R. landra</i> DC.)
10	<i>Sinapidendron</i> spp. (includes <i>S. angustifolium</i> (DC.) Löwe, <i>S. frutescens</i> (Ait.) Löwe, <i>S. rupestre</i> Löwe)
12	<i>Sinapis alba</i> L. (includes <i>S. dissecta</i> Lag.)
9	<i>Sinapis arvensis</i> L. (includes <i>S. allionii</i> Jacq., <i>S. turgida</i> (Pers.) Delile)
7	<i>Sinapis aucheri</i> (Boiss.) O.E. Schulz (= <i>Brassica aucheri</i> Boiss.)
12	<i>Sinapis flexuosa</i> Poir.
9	^a <i>Sinapis pubescens</i> L.
8	<i>Trachystoma</i> spp. (includes <i>T. aphanoneurum</i> Maire & Weiller, <i>T. ballii</i> O.E. Schulz, and provisionally <i>T. labasii</i> Maire)
	Amphidiploids
	(with proposed parentage in parentheses)
17	<i>Brassica carinata</i> A. Braun (<i>B. nigra</i> × <i>B. oleracea</i>)
18	<i>Brassica juncea</i> (L.) Czern. (<i>B. rapa</i> × <i>B. nigra</i>)
19	<i>Brassica napus</i> L. (<i>B. rapa</i> × <i>B. oleracea</i>)
21	<i>Diploaxis muralis</i> (L.) DC. (<i>D. tenuifolia</i> × <i>D. viminea</i>)
15	<i>Erucastrum gallicum</i> (Willd.) O.E. Schulz (<i>E. leucanthum</i> × ? unknown <i>n</i> = 7 taxon)
15	<i>Erucastrum elatum</i> (Ball.) O.E. Schulz (<i>E. littoreum</i> × ? unknown <i>n</i> = 7)

Note: Information was obtained from the following sources; Gómez-Campo (1983), Harberd (1972, 1976), Harberd and McArthur (1972, 1980), Leadlay and Heywood (1990), Snogerup *et al.* (1990), Sobrino-Vesperinas (1988), Takahata and Hinata (1983) and Warwick *et al.* (1992). Nomenclature is based on that in USDA-ARS (The Germplasm Resources Information Network) (2011).

^aAllotetraploids (4x) were also indicated for these cytodemes.

1.2 Description

12. Prakash and Hinata (1980) have summarized the early taxonomic difficulties when only morphological characteristics were used to categorize the numerous and varied forms of the commercially important *Brassica* species. The early proliferation of species names and misclassifications resulted from the wide array of plant forms that occur among plants within the same genome, plus the mimicking of the same morphological features in plants with a different genetic makeup. Although the application of advanced genetic techniques and chemical investigations has clarified relationships, there is still some disagreement among authorities as to whether a particular form should be considered a species, or a subspecies or variety within a species.

1.2.1 *Brassica nigra*

13. Sinskaia (1928) identified two major geographic forms of *B. nigra*, a western form grown in Europe, Africa, Asia Minor and Afghanistan and an eastern form grown in India and as far west as Palestine and Syria. The early forms were of short season, spreading, with semi-erect growth up to a meter tall but taller, more erect material was selected for commercial production (Hemingway, 1995). The prevalent annual weedy form of today varies in height from 0.6 m up to 2.4 m, depending on the competing vegetation and growing conditions. The plant is lightly covered with soft hairs; the lower leaves are large with upper leaves reduced in size. *B. nigra* can be easily distinguished from the commercial *Brassica* crops in that *B. nigra* does not produce a rosette of basal leaves. A typical plant image, including the tap root, is shown in Figure 2. The siliques are short (2-5 cm), hirsute and appressed to the stem of the flowering raceme, with a beak about 0.6 cm long. The small, brown to black seeds exhibit primary dormancy and tend to germinate throughout the growing season.



Figure 2. Illustration of a *Brassica nigra* plant and its parts

Source: Koehler's Medical-Plants (1887) provided by Wikipedia, the free encyclopedia

1.2.2 *Brassica rapa*

14. Plants of *B. rapa* species are widely cultivated as leaf and root vegetables, fodder, and oilseed crops. In addition, they can be a weed of cultivated land and disturbed sites. The widest array of vegetable forms evolved in China with many of the selected forms corresponding to or mimicking those found in the *B. oleracea* complex. Because the selected forms exhibited significantly different morphological traits, early botanists classified them as separate species. Today they are more correctly classified as subspecies or varieties of *B. rapa*.

1.2.2.1 *Brassica rapa* vegetables

15. The plants in the *B. rapa* subsp. *pekinensis* group of vegetables are biennials that have been classified into three variant forms. The var. *cylindrica* has broad but thin, crinkled and conspicuously veined green leaves with white petioles (Figure 3). The leaves are usually tightly wrapped in a cylindrical formation to form a head with a length of 30 to 60 cm and a diameter about 10 to 17 cm. The var. *cephalata* forms a flat head similar to a drum-head cabbage (Figure 3) while the var. *laxa* forms a loose heart. In the second year of growth bolting occurs and the flowering stem is quickly thrust upwards reaching a height of 1.5 m and bearing the characteristic raceme with typical *Brassica* yellow flowers. Common names for this group include pe-tsai, celery cabbage or Chinese cabbage.

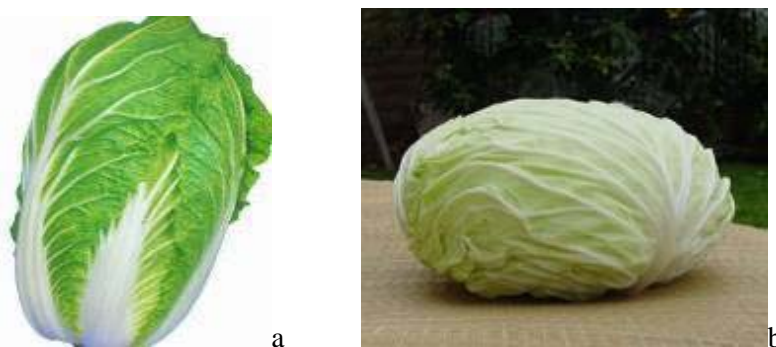


Figure 3.
a) *B. rapa* subsp. *pekinensis* var. *cylindrica*;
b) *B. rapa* subsp. *pekinensis* var. *cephalata*
 Source: courtesy Evergreen Seeds

16. The *B. rapa* subsp. *chinensis* group includes both annual and biennial forms. Bailey (1930) described the subspecies as “a very smooth biennial with large ladle-shaped upstanding radial leaves with thick ivory-white but not wing-margined or toothed petioles.” The clasping, entire leaves have prominent veins and resemble leaves of Swiss chard (Figure 4). The common name for this plant group is pak choi or bok choy. If the plants are harvested in the early stages of growth they may be called ‘baby bok choy’ or ‘Shanghai bok choy’ (Figure 5). The subsp. *parachinensis* is usually included within this group (Figure 6). It is grown for its thick stemmed flowering shoots that are cut for market as the first flowers open, allowing for several harvests. The common names for this variant include Gai Lan and Chinese broccoli. Tsen and Lee (1942) include the subsp. *rosularis* and subsp. *narinosa* in this *chinensis* group. The USDA Germplasm Resources Information Network (GRIN) database includes *rosularis* in the *chinensis* group, but keeps *narinosa* as a separate subspecies (USDA-ARS, 2011). The plants of the subsp. *narinosa* are stout, low growing, glabrous biennials. The lower leaves are small, puckered and orbicular-ovate with broad white petioles, arranged in short clusters. The upper stem leaves are very broad, entire and clasping. Siliques are about 2 cm long or less with a very short, stout beak about $\frac{1}{2}$ - $\frac{1}{3}$ as long as the pod. Tsen and Lee (1942) also place subsp. *japonica* and subsp. *nipposinica* within the *chinensis* group, however, the USDA keeps both of these subspecies separate (USDA-ARS, 2011). These two subspecies are considered synonyms for this form, exhibiting pencil-thin leaf stems supporting deeply indented feathery leaves (Figure 7). The flowering stalks produce siliques about 6 cm long.

17. *B. rapa* subsp. *rapa*, the common turnip, develops a bulbous storage organ in the first year of growth. The top 1-6 cm above ground is an expansion of the hypocotyl that is fused with the expanded root below ground. A narrow tap root extends below the storage organ (Figure 8). Most cultivars are white fleshed except for the exposed above ground portion which, when exposed to sunlight, may turn purple, red or green. Yellow, orange and red fleshed cultivars are also grown. Leaves grow directly from the above-ground shoulder of the expanded hypocotyl and not from a visible crown or neck as occurs in rutabagas (*Brassica napus* var. *napobrassica*). The leaves may be harvested and eaten as ‘turnip greens’. Turnip roots for edible purposes will each weigh about 1 kg but weight will vary with the variety and growing conditions. Cultivars grown for cattle and sheep feed produce much larger roots. The flowering stalk bolts from the over-wintered root the following spring, producing a terminal raceme with siliques about 6 cm long.



**Figure 4. *B. rapa* subsp. *chinensis*,
Bok choy**
Source: courtesy Tainong Seeds



**Figure 5. *B. rapa* subsp. *chinensis*,
Baby or Shanghai bok choy**
Source: courtesy Tainong Seeds



**Figure 6. *B. rapa* subsp. *parachinensis*,
Gai Lan or Chinese broccoli**
Source: courtesy Evergreen Seeds



Figure 7. *B. rapa* subsp. *nipposinica*
Source: courtesy North Carolina State Univ.



Figure 8. *B. rapa* subsp. *rapa* the common turnip
Source: courtesy Wikipedia, the Free Encyclopedia

1.2.2.2 *Brassica rapa* oilseed and weedy forms

18. The oilseed form of *B. rapa* subsp. *oleifera* includes both annual and biennial varieties. Both the spring and winter forms of *B. rapa* mature earlier and withstand cold temperatures better than their *B. napus* counterparts. However, the seed and oil yield is normally lower than *B. napus* so production of the winter form is limited to the more rigorous climates of central Sweden, Finland, north-west China and the foothills of the Himalayan mountains. The plant and growth stages of spring *B. rapa* are illustrated in Figure 9. Following the emergence of the cotyledons the plant quickly produces a tap root and a rosette of leaves that shades the surrounding area reducing weed competition. The lower leaves are stalked, lyrate-pinnatifid with a large end lobe exhibiting sparse hairs on the under side. The upper leaves are much smaller and slightly stalked. In the winter form the plant remains in the rosette stage until exposed to a long vernalization period (40 days) at near freezing temperatures. Day length, and where required, vernalization, determine when bolting of the flower stem will occur. Figure 9 shows only a single raceme but under field conditions the plant produces many flowering branches and with *B. rapa*, as opposed to *B. napus*, it can be difficult to identify the primary raceme. The plant grows to a height of a meter or less. The position of the flower buds on a raceme, relative to the just opened, self-incompatible flowers, can be used to distinguish plants of *B. rapa* from *B. napus*. In *B. rapa* the flowers over top the buds while the reverse is true for *B. napus*. Siliques, some 6 cm long, contain up to 30 brown to yellow seeds in two locules (Figure 10).

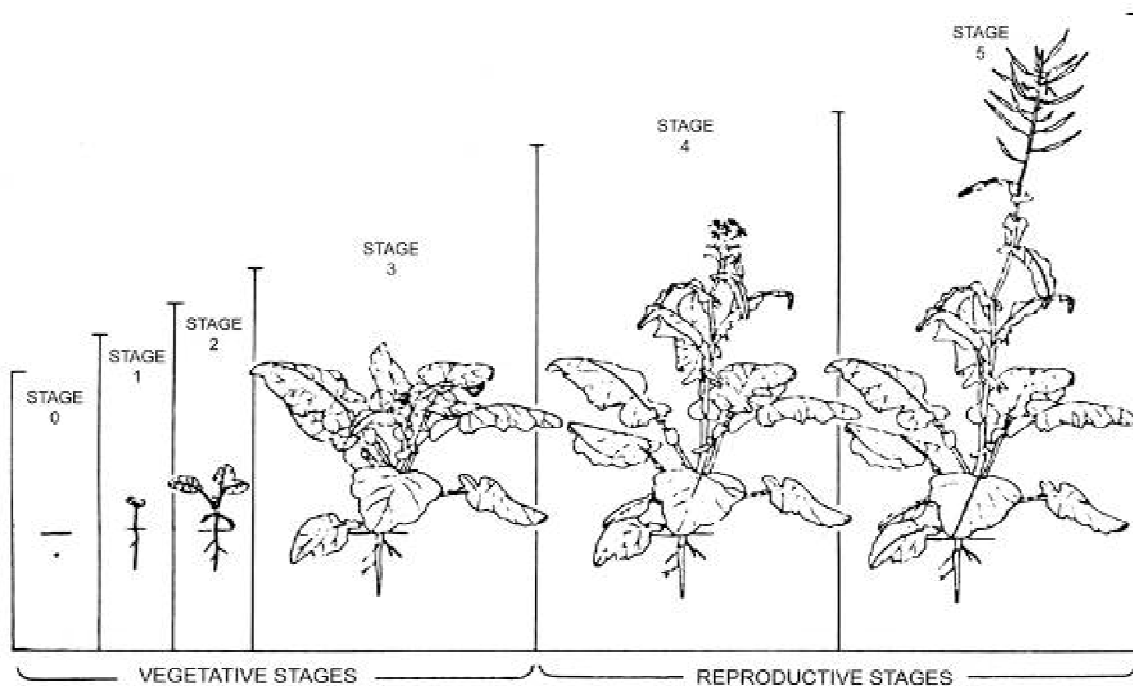


Figure 9. Growth stages in turnip rape (*B. rapa*)

Stages; 0, pre-emergence; 1, seedling; 2, rosette; 3, bud; 4, flower; 5 ripening.

Source: Harper and Berkenkamp, 1975

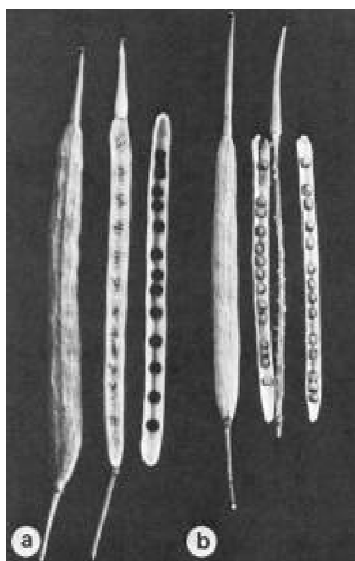


Figure 10. Typical intact and opened siliques of *B. napus* and *B. rapa*

Source: Downey, 1983

- a) *B. napus* showing intact and opened siliques with seeds of the upper locule exposed, while those of the lower locule are partially obscured by the lamella;
 b) intact and open silique of *B. rapa*.

19. *B. rapa* subsp. *campestris* (formerly subsp. *sylvestris*), the weedy form of subsp. *oleifera*, is morphologically indistinguishable from the cultivated spring oilseed *B. rapa*, except that the seed of subsp. *campestris* exhibits primary dormancy, a recessively inherited characteristic.

20. *B. rapa* subsp. *dichotoma*, commonly referred to as toria, is an oilseed form grown on the Indian sub-continent. Morphologically it is indistinguishable from the spring form of *B. rapa* subsp. *oleifera*. Other forms grown on the sub-continent are termed Yellow and Brown Sarson (*B. rapa* subsp. *trilocularis*). These forms have broad siliques containing larger seeds than toria. However, Yellow Sarson is distinguished by its introse anthers, self-compatibility and pure yellow seeds.

21. Autotetraploid *B. rapa* varieties have been developed for use as leafy vegetables, fodder (turnips) and green manure. Tetraploid plants have larger leaves, thicker stems, greater height and larger seeds than their corresponding diploids (Abel and Becker, 2007). However, tetraploids are not used as oilseed crops as their seed and oil yields are significantly lower than their diploid progenitors (Downey and Armstrong, 1962). The much larger tetraploid pollen also takes significantly longer to germinate than *B. rapa* diploid pollen. Thus pollen from *B. rapa* diploid plants has a selective advantage resulting in triploid embryos, which abort (Downey and Armstrong, 1962; Håkansson, 1956) providing strong selection pressure against *B. rapa* tetraploid plants growing in *B. rapa* diploid populations. The slower pollen germination of tetraploid plants could predispose them to out-crossing with related species. On the other hand, since tetraploid *B. rapa* crops are normally consumed or ploughed down before flowering they are unlikely to be a significant factor in gene flow.

1.2.3 *Brassica oleracea*

22. The *B. oleracea* vegetables are often referred to as the 'cole crops' and comprise cabbage, cauliflowers (including broccoli), kales and kohlrabi but not the *B. rapa* vegetables.

1.2.3.1 Wild *B. oleracea*

23. Wild *B. oleracea* var. *oleracea* or wild cabbage is native to the western and southern seaboard of Europe where its tolerance of salt and lime, but its intolerance to competing vegetation, tends to restrict its presence to limestone sea cliffs (Heywood, 1964; Rich, 1991). The plants of this subspecies are biennial or perennial and in the first year produce a rosette of thick, fleshy leaves (Figure 11). Following vernalization a flowering stalk 1 to 2 m tall arises from the centre of the rosette bearing a raceme of self-compatible, yellow flowers.



Figure 11. Wild *B. oleracea* plants in their first year of growth
Source: Wikipedia, the Free Encyclopedia

1.2.3.2 *B. oleracea* var. *capitata*, cabbage

24. The cabbage is a biennial plant that in the first year of growth produces a dense, terminal head of tightly wrapped leaves on a short stout stem. The head is surrounded by a rosette of large fleshy leaves (Figure 12a). Three main types of heads, smooth green, red and Savoy are commercially produced (Figure 12b). In the second year the head splits open and the flowering stalk bolts to 1.5 to 2.0 m tall with branches bearing flowering racemes of self-incompatible flowers.

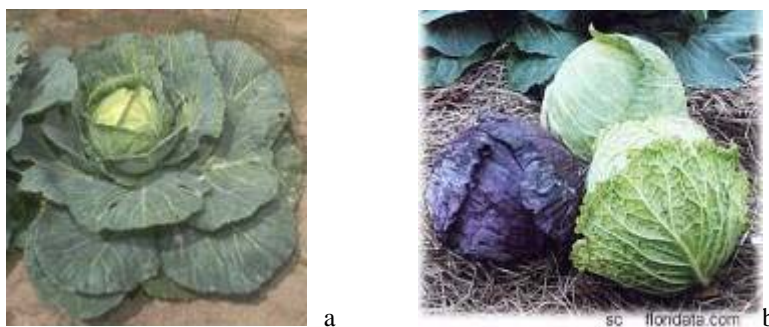


Figure 12. Heads of *B. oleracea* var. *capitata* and Savoy cabbages

Source: courtesy Floridata

- a) Head of cabbage, *B. oleracea* var. *capitata* with its rosette leaves intact
b) Heads of red, smooth green and Savoy cabbage with lower leaves removed

1.2.3.3 *B. oleracea* var. *botrytis*, broccoli and cauliflower

25. Cauliflower is derived from broccoli, being selected for short stout stems with a dense, terminal head or curd, made up of arrested inflorescence meristems, over topped by leaves (Figure 13). About 10% of the meristem mass will eventually develop into normal flowers and set seed (Sadik, 1962). Specific alleles of the *BoCAL-a* gene have been shown to be associated with discrete inflorescence

morphologies (Smith and King, 2000; Purugganan *et al.*, 2000). Smith and King (2000) present evidence suggesting that the cauliflower curd arose in southern Italy from a heading Calabrese broccoli via an intermediate Sicilian crop type. Broccoli differs from cauliflower in that broccoli flower heads tend to be smaller with more slender floret-stalks and are made up of arrested green (or purple) flower buds whereas the heads of cauliflower are formed by a condensed and thickened, malformed white (also purple or lime green) flower cluster. Both crops are biennial and, provided the plants have been vernalized, produce viable flowers and pods in the second year from the stump or parts of the head that remain. Vernalization requires a prolonged cold period of at least 10 days with temperatures between 2 to 10 °C. The larger the plant when exposed to the cold treatment the greater the incidence of bolting. Plants of both crops are more susceptible to frost and less tolerant of heat and drought than cabbage. The cultural requirements of broccoli and cauliflower are similar but broccoli generally grows more rapidly. Most varieties are now F₁ hybrids.



Figure 13. Head of cauliflower (left) and broccoli (right) *B. oleracea* var. *botrytis*
Source: courtesy Cavaganaro, David/Sunset/Invision

26. The broccoli referred to above is more correctly known as “calabrese” broccoli. It produces a single head and is the form that is of greatest commercial importance. The “sprouting” broccoli, var. *italica*, produces a succession of small flowering heads over an extended period (Figure 14) while the “Romanesco” broccoli produces a head characterised by multiple cone shaped spirals consisting of masses of small flower buds (Figure 15).



Figure 14. Sprouting purple broccoli
Source: courtesy Mr. Fothergill’s Seeds Ltd. UK



Figure 15. Romanesco broccoli
Source: courtesy Mr. Fothergill’s Seeds Ltd. UK

1.2.3.4 *B. oleracea* var. *viridis*, collards and kale

27. The kales and collards are biennials but are usually harvested in the first year for their edible leaves. They closely resemble their wild cabbage progenitors. Collards have large, smooth fleshy leaves with smooth margins (Figure 16). The leaves of kale are smaller and thinner than those of collards and many cultivars produce fringed, wavy-edged or feathery leaves (Figure 16). A thick flowering stem up to 1.5 m tall emerges in the second year. One form called ‘Walking Stick’ kale produces a tall straight stem which, when dried and polished, makes a fine walking stick.



Figure 16. *B. oleracea* var. *viridis*, collard plant (left) and row of kale (right)
Source: courtesy Flordata

1.2.3.5 *B. oleracea* var. *gemmifera*, Brussels sprouts

28. *B. oleracea* var. *gemmifera* plants are cool season biennials with simple erect stems up to 1 m tall, bearing round to heart-shaped simple leaves with lengthy petioles. The leaves are glabrous with the colour varying from light to deep greyish-green. In the first year auxiliary buds or sprouts are borne beneath the leaves on an elongated stem (Figure 17). The buds are modified leaves that form small heads up to 30 mm in diameter. Following vernalization a seed head is produced from which a flower stalk emerges bearing perfect, self-incompatible flowers on terminal racemes. The seeds, weighing about 2.8 g/1000 are borne in typical, two locule siliques.



Figure 17. *B. oleracea* var. *gemmifera*, Brussels sprouts
Source: courtesy of Limagrain

1.2.3.6 *B. oleracea* var. *alboglabra*, Chinese kale

29. The var. *alboglabra* is widely grown throughout south-east Asia as a leaf and stem vegetable. The perennial plants are grown as annuals, producing dull or glossy thick green, glaucous, elliptic leaves about 25 cm long. The plants commonly called Chinese kale and kailan attain a height of up to 40 cm in the vegetative stage and 1 to 2 m at the end of flowering. Upper stem leaves are oblong, petioled or non-clasping. The white flowered inflorescences develop siliques 5 to 8 cm long (Herklots, 1972).

1.2.4 *Brassica napus*

1.2.4.1 *B. napus* var. *napobrassica*, rutabaga or Swede

30. *B. napus* var. *napobrassica*, the common rutabaga or Swede, is a biennial with similar characteristics to the turnip. The bulbous root develops from the hypocotyl in the first year of growth (Figure 18). The surface of the root may be purple, white or yellow with the inner content solid yellow or white fleshed. The thick, smooth, dark green leaves emerge from the crown or neck of the root to form a ground covering rosette that shades out competing weeds. The presence of a root crown or neck distinguishes rutabagas from turnips. Early in the second year the flower stalk bolts from the root crown and the self-compatible flowers produce short beaked siliques on short pedicels containing two rows of round black seeds. Rutabagas are used for human consumption and for late fall cattle grazing.



Figure 18. *B. napus* var. *napobrassica*, rutabaga or Swede
Source: courtesy Floridata

1.2.4.2 *B. napus* var. *pabularia*, Siberian or rape kale

31. This sub-species has both annual and biennial forms with much branched erect stems up to 1.5 m tall. Lower leaves are glaucous and lobed. Upper stem leaves are lanceolate, sessile and clasping (Figure 19). The much branched inflorescence is an elongated raceme producing siliques 5 to 11 cm long and 2.5 to 4 mm wide with a slender 0.5 to 3 mm long beak. The tap root produces many side branches. The crop is grown as a leafy vegetable and for fodder.



Figure 19. *B. napus* var. *pabularia*, Siberian or rape kale
Source: courtesy Floridata

1.2.4.3 Oilseed rape, *B. napus* var. *napus* f. *annua* and f. *biennis*

32. Oilseed *B. napus* has both an annual (spring) and a biennial (winter) form. The biennial form is less winter hardy than winter wheat which restricts its production to areas with mild winter conditions such as northern Europe and central China. The annual form is grown as a spring crop in western Canada and northern China but also as a winter crop in Australia and other countries with very mild winters.

33. Growth stages of annual *B. napus* plant development are illustrated in Figure 20. The glaucous lower leaves form a rosette from which the flowering stalk emerges bearing a dominant, indeterminate main raceme. The upper stem leaves are small, lanceolate, sessile and clasping. Plants of the species *B. napus*, *B. rapa* and *B. juncea* can be distinguished by their upper leaf attachment to the stem as illustrated in Figure 21. Flowering begins with the lowest bud on the main raceme and continues upward with three to five or more flowers opening per day. The buds, unlike those of *B. rapa* are held above the uppermost open flowers. Flowers on the secondary branches begin to open about three days after the opening of the first flowers on the main raceme. The siliques are ascending on slender pedicles and about 7 to 10 cm long with a beak about 1.3 cm long. Seeds are dark brown to black, and weigh 2.5 to 5.5g per 1000 seeds.

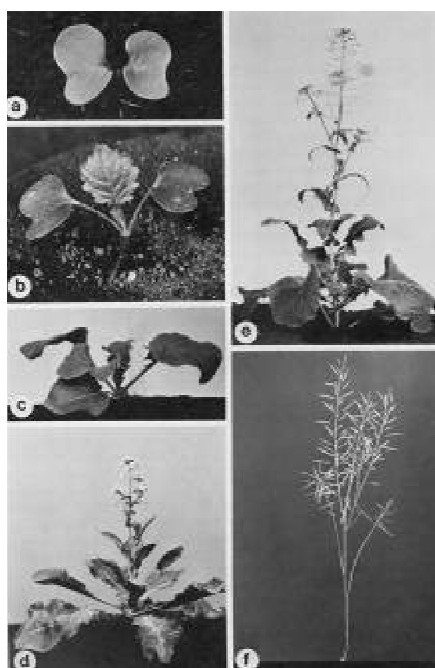


Figure 20. Growth stages of *B. napus* var. *napus* f. *annua*

Source: Downey, 1983

- (a) seedling cotyledons;
 (b) cotyledons and first true leaf; (c) rosette;
 (d) flowering; (e) pod set and (f) mature plant

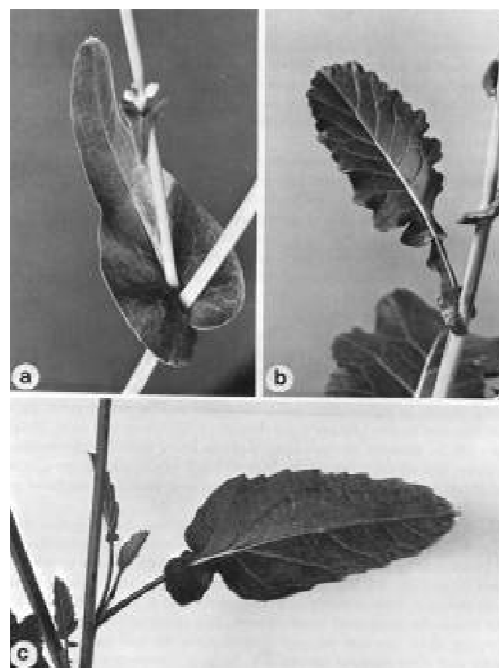


Figure 21. Upper leaves of *B. rapa*, *B. napus* and *B. juncea*

Source: Downey, 1983

- (a) *B. rapa*, fully clasping stem;
 (b) *B. napus* partially clasping and
 (c) *B. juncea*, non-clasping

1.2.5 *Brassica juncea*

1.2.5.1 *B. juncea* vegetables

34. In China and south-east Asia many vegetable forms of *B. juncea* have been developed and classified as species or subspecies under numerous names, depending on the morphological features given

the greatest importance. Kumazawa and Able (1955) examined some 200 East Asian cultivars of *B. juncea* vegetables grown in China, Japan, Chinese Taipei and Nepal on the basis of their plant size, root form, tillering and leaf characteristics. All accessions of *B. juncea* and its subspecies were described as annuals and placed in 25 different groups within eight classes. These classes were further condensed into four subspecies. The authors state that the subspecies evolved from the leafy and oilseed forms of Brown mustard, *B. juncea* (L.) Cross. From the collection, the authors illustrated a normal or “Ordinal” root form, a turnip-like rooted form, [also described by Dixon (2007)] and a little known form with a tuberous basal stem (Figure 22). The four subspecies were grouped and characterized as follows.

1. subsp. *napiformis* (Pailleux & Bois) Gladis, grown for its tuberous turnip-like root. This subspecies bolts late and has a high tolerance to cold;
2. var. *japonica* (Thunb.) L.H. Bailey, characterised by curled, narrow or dissected leaves;
3. subsp. *integrifolia* (H. West) Thell., characterised by entire or little lobed basal leaves. Herkots (1972) notes that some cultivars may form a tight head (Figure 23); and
4. var. *rugosa* (Roxb.) N. Tsen & S. H. Lee, includes cabbage leafed forms with large entire or serrated radical leaves. The tuberous basal stem form (Figure 22) is included in this subspecies.

35. Herkots (1972) places var. *rugosa* within subsp. *integrifolia* but also puts forward the var. *sareptana* as characterised by lyrate-lobed basal leaves and var. *crispifolia* as having dissected, crisped lower leaves. More recently, a *B. juncea* Biology Document (CFIA, 2007) quoted the grouping by Spect and Diederichsen (2001) into the following four sub-species.

1. subsp. *integrifolia*, used as a leaf vegetable in Asia;
2. subsp. *juncea*, cultivated mainly for its seeds, occasionally as fodder;
3. subsp. *napiformis*, used as a root-tuber vegetable. Dixon (2007) describes this subspecies as having high cold tolerance and an enlarged conical root; and
4. subsp. *tsatsai* from which stalks and leaves are used as vegetables in China.

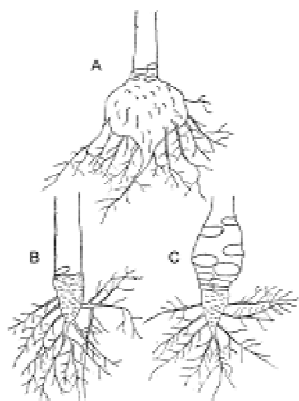


Figure 22. Three forms of *B. juncea* with a) bulbous root, b) normal or ‘Ordinal’ root, and c) tuberous basal stem
Source: Kumazawa and Able (1955)



Figure 23. *B. juncea* subsp. *integrifolia*, Heading mustard, BauSin
Source: courtesy AgroHaitai Ltd.

1.2.5.2. *B. juncea*, oilseed and condiment mustards

36. Plants of this species, grown for their seed oil or condiment production, are normally referred to simply as *B. juncea* without the attachment of a subspecies name. However, Spect and Diederichsen (2001) classify this plant group as *B. juncea* subsp. *juncea*. Plants for both oil and condiment are similar in their morphology but differ in seed oil percentage and the type and amount of glucosinolates present in the seed.

These forms are annuals that grow to about 1.2 m as spring sown crops in western Canada and Europe. On the Indian sub-continent they are grown as a winter crop where, under short days, plants grow up to 2.1 m tall. The plants are green and sometimes slightly glaucous. The lower leaves of the rosette are rather thin, elliptic to obovate and lyrate-lobed or divided. The upper stem leaves are small, narrow and not clasping (Figure 21). Depending on the day length and temperature the flowering stalk bolts and produces a raceme with no terminal flower. As with *B. napus* the buds are borne above the open flowers. Apical dominance is present with the secondary racemes initiated about three days after flowers open on the main raceme. The silique is about 7 cm long containing seed weighing 2.5 to 3.0 g/1000 seeds.

1.3 Geographic Distribution, Ecosystems and Habitats, Cultivation and Management Practices, Centres of Origin and Diversity

1.3.1 Introduction

37. From an ecological and agronomic point of view, both the spring and winter forms of oilseed rape exhibit two undesirable characteristics. First, mature pods tend to shatter, leaving large but variable amounts of seed on the ground at harvest (See Section 2.2.5.1 below, Contribution of *B. napus* harvest losses to persistence). Pod shatter not only results in lost yield but also sets the stage for large numbers of volunteer plants in subsequent crops. Fortunately *B. napus* seeds have no primary dormancy so if moisture and temperature are adequate the vast majority of these seeds germinate and are killed by frost, herbicides, cultivation or predators (See below). The opportunity for *B. napus* to acquire primary dormancy is limited due to the vast majority of fields being sown each year with high germination certified seed.

38. The second undesirable characteristic is the tendency for a proportion of the shattered seed to acquire secondary dormancy. Such dormancy is induced by abiotic stresses (See Section 2.2.5.3, Persistence). Although most of the shattered seed will quickly be reduced by fatal germination, predation, disease and abiotic stress, a small percentage can remain dormant and viable for 10 years or more (Schlink, 1998; Lutman *et al.*, 2003). Thus *B. napus* is able to establish seed banks within cultivated fields (See Lutman *et al.*, 2005 and below). As a result traits or genes that have been genetically silenced or augmented within improved varieties may be reintroduced. Examples would be the genetic blocking of the biosynthesis of erucic acid in rapeseed oil, the reduction in linolenic acid content and the augmentation of oleic fatty acids in the oil, or reduction of glucosinolates in the oilseed meal.

39. It should be noted however, that there is considerable genetic variability within the species and its close relatives in both the degree of pod shatter and the percentage of induced dormancy. Until recently these characteristics have not been a priority for oilseed rape breeders but progress is possible. Wang *et al.* (2007b) have clearly demonstrated that selection for reduced pod shatter in *B. napus* can be achieved. In addition, Østergaard *et al.* (2006) has shown that expressing the *Arabidopsis* FRUITFULL gene in *B. juncea*, using a CaMV 35S promoter, produces shatter-resistant plants. Although the shatter-resistant pods held their seed too tightly for combine harvesting, a weakened form of the FRUITFULL gene could result in an economically and environmentally valuable advance. It is unlikely that conventional breeding will lead to complete elimination of the shattering characteristic but there appears to be considerable room for improvement. Further, Pekrun *et al.* (1997b), Gruber *et al.* (2004, 2009) and Gulden *et al.* (2004a) have all shown that among *B. napus* varieties, of both spring and winter forms, there is a wide range in the percentage of seed susceptible to induced dormancy. Thus the application of conventional breeding techniques to select varieties producing seed resistant to secondary dormancy should greatly reduce the presence of volunteers in subsequent crops.

40. Ecologically *B. napus* is described as a cultivated crop where escaped plants become colonisers of waste places. However they are not invasive of natural habitats. Colonisers are defined as species that occupy disturbed sites or habitats but with populations that keep moving, founding new populations while

losing old ones (Williamson, 1996). Feral populations of *B. napus* are most frequently found along road and rail verges, field margins and in disturbed soils. The reports on the abundance and persistence of such feral populations vary considerably from country to country and between the spring and winter forms. Williamson (1996) noted that colonising species are not the same as invaders, even if they have high intrinsic rates of increase, as exhibited by *B. napus*. He classifies *B. napus* in Britain as intermediate between naturalised and casual. On the other hand, recent intensive surveys of feral sites in mainland Europe have identified feral populations in higher frequencies than anticipated, with some sites able to sustain themselves in a semi-permanent state (Pivard *et al.*, 2008). Such reports have given rise to concerns by some that a proportion of feral populations could become permanent and in time result in the invasion of natural habitats.

41. Although the species does have the weedy characteristics noted above, producing many propagules (seeds), plus the ability to cross with some weedy relatives, it is not competitive with perennial grasses that dominate the natural habitat. It should also be noted that oilseed rape has been part of the European landscape for a very long time as have the truly weedy, related species, *Sinapis arvensis* and *B. rapa*. However, none have become invasive of natural habitats. In recent years the area of oilseed rape cultivation and intensity of production has increased worldwide. For example since 1970 oilseed rape production in France and Germany has increase 4.5 and 8 fold, respectively. In the same period the Canadian oilseed rape acreage has quadrupled, thus a wider and more frequent occurrence of feral populations is to be expected. The spring form of oilseed rape is much less likely to form feral populations or to be self-sustaining since fall germination is normally fatal while frosts will kill many seedlings that germinate in the spring. Although Knispel *et al.* (2008) has reported some transient feral population in the province of Manitoba, Canada, such roadside populations are rare over most of the Prairie Provinces, except near collection points and to a limited extent along railroad verges. This is because in western Canada most road verges are mowed in late August before feral populations set viable seed. Such roadside mowing is essential to prevent snowdrifts across roads that tall vegetation can cause. In contrast, in Europe the winter form can avoid being killed by the fall road maintenance since mowing does not usually affect the established first year rosettes, leaving some plants to flower and set seed before the next fall mowing.

42. Different agronomic practices also influence the size and persistence of volunteer populations. In Europe, the large amount of straw remaining after harvest plus the short time between the July harvest and August sowing dates encourages ploughing down of residue, resulting in seed burial. In Canada on the other hand, ploughing is not practiced and most fields are spring sown into undisturbed stubble (minimum or zero tillage) from the previous year's September harvest (Hall *et al.*, 2005). Thus, seed burial is minimized and harvest seed losses are exposed to environmental hazards. The result is that in Europe old or discontinued cultivars or genotypes will persist in the seed bank for a much longer time than in Canada. This is clearly illustrated in the changeover from high to low erucic acid *B. napus* varieties. In the German oilseed rape growing province of Schleswig-Holstein it required 10 years to reduce the commercial crop from the traditional high erucic varieties (50% erucic) to the desired level of 2% (Sauermann, 1987). In Canada the same results were obtained in 3 years (Daun, 1983).

1.3.2 Geographic distribution

43. The genus *Brassica* and its wild relatives are part of the tribe Brassiceae that has its origin in the Mediterranean basin and in south-western Asia. However, the geographic centre is thought to be in the south-western Mediterranean region (Algeria, Morocco and Spain) where some 40 genera have been shown to be endemic or exhibit maximum diversity (Hedge, 1976; Gómez-Campo, 1980, 1999; Al-Shehbaz, 1984; Al-Shehbaz *et al.*, 2006; Warwick and Hall, 2009). For the subtribe Brassicinae, Hedge (1976) leaves little doubt that it originated in the Mediterranean basin. The species distribution of the Brassicaceae family is concentrated in the northern temperate zone and south-western and central Asia (Holm *et al.*, 1997). Few species are found in hot, humid tropics.

1.3.2.1 *B. nigra*

44. *B. nigra* or black mustard was widely grown for the sharp pungency of its seeds and as a leaf vegetable. Prakash and Hinata (1980) placed the species origin in central and south Europe. It is one of the oldest recorded spice crops which undoubtedly resulted in its early and widespread distribution across Europe, Africa, Asia and the Indian sub-continent, and its dehiscent siliques with primary dormancy of the seed ensured its persistence. The GRIN describes the species distribution as widely naturalised in the following regions and countries. **Africa;** countries along the south shore of the Mediterranean as well as Eritrea and Ethiopia. **Temperate Asia;** Afghanistan, Iran, Iraq, Israel, Lebanon, Syria, Turkey, Armenia, Kazakhstan and northwest China. **Indian sub-continent;** India, Pakistan, Nepal. **Europe;** All countries in western and eastern Europe as well as the Balkans and Greece. The crop was introduced to the Americas and Australia as a spice. However, in the 1950s the crop was displaced by the higher yielding, pungent *B. juncea* that was better suited to mechanical harvesting. Although in many regions Black mustard is now a weed of waste places, it has never become established on the Canadian prairies, although it is present throughout much of the United States.

1.3.2.2 *B. rapa*

45. *B. rapa* is thought to have originated in the mountainous areas near the Mediterranean sea (Tsunoda, 1980). The time of domestication is unknown. Sinskaia (1928) proposed two main centres of origin, one being the Mediterranean and the other the Afghanistan-Pakistan region. The species appears to have attained a wide distribution throughout Europe, parts of Africa, Asia and the Indian sub-continent before recorded history. Excavations in China reported the presence of *B. rapa* seed at a 6,000 to 7,000 years old archaeological site (Liu, 1985). Indian Sanskrit literature mentions the plant about 1599 B.C. (Prakash, 1961), and Renfrew (1973) indicated that *B. rapa* seed was consumed in Scandinavia as early as 350 B.C. *B. rapa* is grown as an oilseed crop in northern Europe, north west China, the foothills of the Himalayas, and northern India while the vegetable forms were selected and modified in Asia, primarily in China. The oilseed form was introduced to Canada by a Polish immigrant about 1936 (Boulter, 1983) and Australia began its first investigations on the *B. rapa* crop in the early 1960s (Salisbury, 2002) but it has now been superseded by *B. napus* varieties. *B. rapa* also has a weedy form that differs from the cultivated plant in exhibiting primary dormancy and has a worldwide distribution (Figure 24).



Figure 24. The world distribution of *B. rapa* as a reported weed.

Source: modified from Holm *et al.*, 1997

1.3.2.3 *B. oleracea*

46. The centre of origin for the *B. oleracea* species is along the European Atlantic coast while the wild related forms still grow on the islands and along the northern coast of the Mediterranean. The various forms of this species were developed in Europe and did not reach Asia until about the 16th century (Liu, 1985). The many cultivated forms of this species have been introduced and grown worldwide, with the exception of some tropical areas.

1.3.2.4 *B. napus*

47. *B. napus* is of relatively recent origin (<10,000 years; see Section III and Figure 41) resulting from the interspecific cross between plants of *B. oleracea* and *B. rapa*. The cross must have occurred where the two species were growing in close proximity along the European Atlantic or Mediterranean coasts. Dispersal of the species is thought to have occurred throughout Europe in the 16th century with the introduction to the Americas in the 17th and 18th centuries and the Far East in the 19th century (Liu, 1985).

1.3.2.5 *B. juncea*

48. *B. juncea* is believed to have arisen about 10,000 years ago as the result of an interspecific cross or crosses between plants of *B. rapa* by *B. nigra*. Evidence suggests that one primary centre of origin is China, where the greatest divergence of forms evolved (Prain, 1898; Sinskaia, 1928; Vavilov, 1949). A second centre of origin is thought to be Afghanistan and adjoining regions (Olsson, 1960; Mizushima and Tsunoda, 1967; Tsunoda and Nishi, 1968) from where it spread to a secondary centre on the Indian subcontinent and became a major oilseed crop (Hemingway, 1995; Prakash and Hinata, 1980). GRIN (USDA-ARS, 2011) lists *B. juncea* as native to temperate Asia including China, Mongolia, eastern and western Siberia, Kazakhstan, Kyrgyzstan, Tajikistan and Turkmenistan. It has been introduced as a condiment crop to Europe, the Americas, Australia and New Zealand. It has been designated a weed of southern European Russia, the Caucasus, central Asia and southern Siberia and a casual or feral plant in southern and southeast Asia, Africa and America (CFIA, 2007).

1.3.2.6 *B. carinata*

49. *B. carinata*, like *B. juncea*, is believed to have arisen about 10 thousand years ago as a result of an interspecific cross between plants of *B. nigra* and *B. oleracea*. The cross is thought to have occurred in the Mediterranean region where both species were present. As the climate in North Africa became dryer, *B. carinata*, along with the flora of the moist Mediterranean region, moved south to the highlands (1300-1500 m) of Ethiopia. The species distribution from its Ethiopian centre of origin has been limited to neighbouring east African countries. Recently it has been introduced as an oil crop to India and as a species of commercial interest in Spain and Canada.

1.3.3 *Ecosystems and habitats where the species occurs natively and has naturalised*

50. There are few areas of the world where members of the family Brassicaceae are totally absent. The exceptions are the Antarctic and some parts of the tropics. However, even in the tropics the family is thinly represented by some introduced cosmopolitan weeds that have become established. The genera and species of the family occur in greatest number and diversity in the temperate zone of the northern hemisphere and in particular, the areas surrounding the Mediterranean basin and throughout the southwest and central regions of Asia (Figure 25) (Hedge, 1976). Although the generic and specific endemism in the family is highest in the Irano-Turanian region, the centre of the present day subtribe, Brassicinae, lies in the Mediterranean basin (Hedge, 1976).

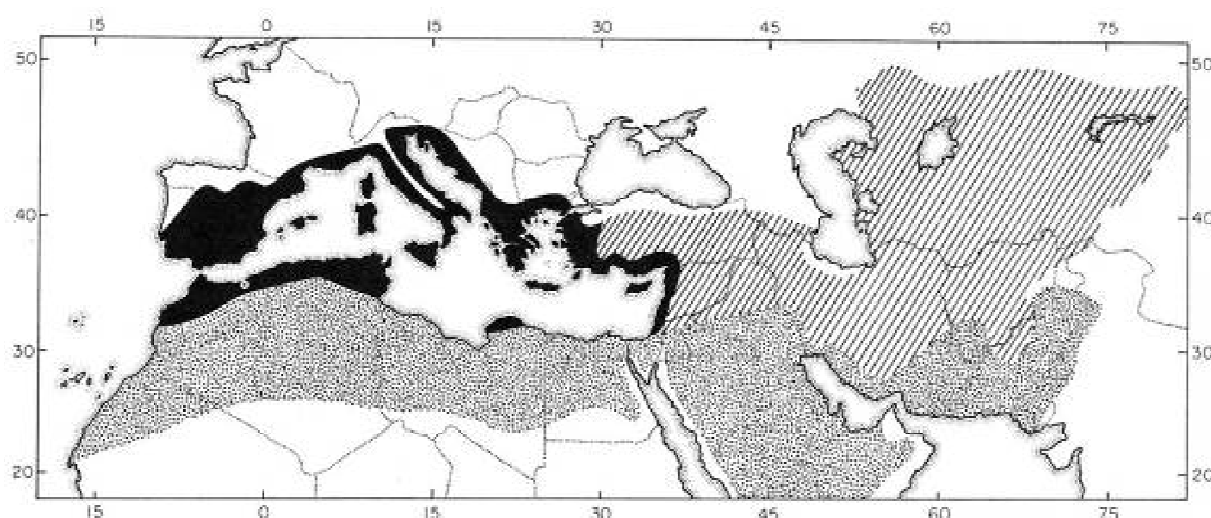


Figure 25. Approximate areas of the phytogeographic regions containing the world's greatest representation of Brassicaceae genera.

Source: after Hedge, 1976. They encompass the Mediterranean (black); the Irano-Turanian (striped) and the Saharo-Sindian (dotted) regions.

1.3.3.1 Feral populations in disturbed soils

51. Due to the large seed losses in commercial *B. napus* fields and the potential loss during transport and handling, the surviving seeds give rise to volunteers in subsequent crops and feral populations in non cultivated areas (CETIOM 2000; MacDonald and Kuntz, 2000; Orson, 2002; Pessel *et al.*, 2001; Price *et al.*, 1996). Volunteers are controlled by cultivation and herbicide application. In both Canadian and United-Kingdom trials, the numbers of genetically modified (GM), herbicide resistant (HR) *B. napus* volunteers in the year following GM trials were comparable to, or less than, conventional *B. napus* (Crawley *et al.*, 1993; Booth *et al.*, 1996; Hails *et al.*, 1997; Rasche and Gadsby, 1997; Sweet *et al.*, 1997, 1999a, b; Sweet and Shepperson, 1998; Norris *et al.*, 1999). In their survey of Canadian commercial fields, MacDonald and Kuntz (2000) found the same trend, with similar numbers of volunteers in the year following cultivation of GM HR canola compared to conventional varieties. Furthermore, prior to any field operations, they found an average over all fields of 200 volunteers /m². Initial soil disturbance was effective in controlling these emerged *B. napus* volunteers, but shallow cultivation resulted in the emergence of an even greater number of volunteers. Post-emergent weed control programme employed by the producer for the non-GM volunteers was also effective in controlling the GM volunteers (MacDonald and Kuntz, 2000). Downey and Buth (2003) reported that GM HR volunteers with single or stacked traits were readily controlled in western Canada by the same agronomic practices that are standard for controlling conventional canola volunteers. In Australia, post harvest monitoring of GM HR (glufosinate or glyphosate) trial locations for six years indicated volunteer populations were adequately controlled by herbicide application or broadacre cultivation (either in-crop or by conservation tillage) (Salisbury, 2002).

52. Feral populations of *B. napus* can be found at various densities on road verges, along field margins and railway lines in all countries where it is grown (*e.g.* Crawley and Brown, 1995; Wilkinson *et al.*, 1995; Squire *et al.*, 1999; MacDonald and Kuntz, 2000; Agrisearch, 2001; Pessel *et al.*, 2001; Orson, 2002; Salisbury, 2002). Populations may also become established in port areas where *B. napus* cargos are handled (Ramsay *et al.*, 2003; Saji *et al.*, 2005; Aono *et al.*, 2006). Annual recruitment to such sites is likely to be more from passing transport vehicles than from an established seed bank. *B. napus*, as with other *Brassica* species, is a coloniser of disturbed soils where it competes with other primary colonisers.

However, *B. napus* is a poor competitor and is not regarded as an environmentally hazardous colonising species (European Commission, 1998a, b, 1999, 2000; Beckie *et al.*, 2001; Dignam, 2001). Unless the habitats are disturbed on a regular basis, *B. napus* will be displaced (OECD, 1997).

53. In western Canada, roadside verges, field margins and railway lines were surveyed for canola plants (MacDonald and Kuntz, 2000). Only 13 and 27 volunteer *B. napus* plants were found in the mowed roadside over the respective 7 and 27 km surveyed, and no plants were found in tall, un-mowed grass. Surveys of rail beds leading from local grain elevators, approximately 3 and 5 km long, identified 287 and 29 plants, respectively, growing at the interface of the rail bed gravel and the tall grass of the right of way. No plants were located on the rail tracks or in the tall grass of the right of way. Similarly in Australia, a survey, making 400 observations in 5 × 20 m areas along 4,000 km of roads in oilseed rape growing areas, found *B. napus* plants in only 31, 20, 13 and 9% of the observation points in southern New South Wales, Western Australia, Victoria and South Australia, respectively. Nearly all the plants were growing within 5 m of the roadside with the vast majority close to or alongside the road edge, suggesting they originated from seed dropped from passing vehicles (Agrisearch, 2001).

54. In the United Kingdom, Crawley and Brown (1995) found that along undisturbed roadways the persistence of *B. napus* is about 3 to 4 years and that the density of such feral populations is correlated with human activities, such as vehicle transport. In a three year assessment of feral populations in Scotland, Wilkinson *et al.* (1995) found that the turnover of populations was high with only 19% of the 1993 populations persisting into 1994 and 12% of the 1994 population persisting into 1995. Crawley and Brown (1995) obtained similar results in southern England. In a study conducted in Germany from 2001 to 2004 Dietz-Pfeilstetter *et al.* (2006) found persistence rates for feral populations of 29% between 2001 and 2002, of 12% between 2002 and 2003 and 80% between 2002 and 2004. However, molecular profiling using ISSR-PCR (Inter-Simple Sequence Repeats) revealed that plants appearing in successive years largely belonged to different genotypes, suggesting new seed input and an even higher turnover of populations.

55. Reuter *et al.* (2008) investigated a 500 km² area in the region of Bremen, Germany and reported average densities of 1.19 and 1.68/km² of feral and volunteer oilseed rape populations in rural and urban areas, respectively. The investigation showed that population density varies between years and feral plants tend to be smaller in stature (by at least 40%) than plants growing on cultivated land.

56. Surveys by Agrisearch (2001) and MacDonald and Kuntz (2000) suggest that, to survive, spring *B. napus* roadside populations need to be regularly replenished. However, in France, Pessel *et al.* (2001) found roadside feral populations contained plants of old varieties that had not been grown for 8 to 9 years, indicating that the seed source was not entirely from recent vehicle spillage. These results are in keeping with previous reports that seed of old rapeseed varieties can persist for at least 5 to 10 years after they were last reported grown (Squire *et al.*, 1999; Orson, 2002). Pessel *et al.*, (2001) suggested that the analysed roadside feral populations arose from multiple spillages from different fields or germination of seed from a mixed seed bank or most likely, both.

57. In Austria, Pascher *et al.* (2006) genetically analysed plants from nine selected feral populations consisting of 50 to 150 individuals. They found the feral populations were genetically more diverse than could be explained by the dominant varieties grown in the area in the previous five years. They concluded that even though the feral populations largely reflected the genetic makeup of the dominate varieties being grown, a significant portion of plants had originated from seed banks older than five years. They also found that feral populations disappeared more quickly under dense grass cover than at sites with little vegetation, but genetic diversity remained unchanged. Their results indicated that genetic migration from commercial varieties to feral populations was five times greater than the reverse.

1.3.3.2 Feral populations in natural habitats

58. In natural (undisturbed) ecosystems *B. napus* is not considered to be invasive or even a significant component of any natural plant community (AAFC, 1994; Warwick *et al.*, 1999; Beckie *et al.*, 2001; Dignam, 2001).

1.3.4 Production and agronomy of Brassica oilseed crops

59. The world demand for edible oils and more recently for biodiesel, has led to a rapid growth in the production of most oilseeds with total seed oil produced increasing by about 4% each year. The percentage growth in the world *Brassica* seed oil production has increased some 60% between 1995-1999 and 2005-2009 (Table 1). The locations of the major rapeseed/mustard producing regions are shown in Table 4. The expanded *Brassica* oilseed production has resulted from both an increase in the area sown, primarily in China, as well as the yield per unit area that has increased in China and to a lesser extent in the EU (Table 4).

Table 4. Area harvested, production and yield by major *Brassica* oilseed producing countries, averages 1995-1999 to 2005-2009

Producing country	Area harvested '000 ha			Production '000 tonnes			Seed yield kg/ha		
	1995-99	2000-04	2005-09	1995-99	2000-04	2005-09	1995-99	2000-04	2005-09
EU 27¹	3,944	4,255	5,407	11,038	12,221	18,108	2,786	2,862	3,082
China	6,708	7,244	6,740	9,391	11,573	12,070	1,399	1,597	1,842
Canada	4,917	4,366	5,322	6,866	6,237	10,510	1,398	1,415	1,790
India	6,541	5,110	7,280	5,756	5,045	7,239	884	982	1,079
Australia²	929	1,335	913	1,230	1,529	1,395	1,370	1,146	1,084
U.S.A.	292	497	452	443	762	665	1,505	1,555	1,635

¹EU 27 = Total production of the 27 member states of the European Union

²Extreme drought greatly reduced Australian production and seed yield in the 2005-09 period

Source: FAOSTAT

1.3.4.1 Cultivation and management of oilseed crops

60. The small seeds of the *Brassica* oilseed crops require that the seed be sown at shallow depths, 2-3 cm below the soil surface, into a firm, moist seedbed. Under favourable growing conditions the seedlings emerge within 4 to 5 days of sowing. Cotyledon expansion is quickly followed by the formation of a rosette of 7 to 8 true leaves from which the flowering stalk bolts. The length of time the crop remains in the rosette stage can vary from less than 30 days to more than 210 days depending on climatic conditions and the species and form grown. The complete growth cycle may be as short as 70 days (*B. rapa*) or as long as 380 days for winter *B. napus* varieties in China (Sun *et al.*, 1991).

61. Although the *Brassica* oilseed crops prefer a deep loam soil it does well when sown in a wide range of soil types and conditions and can tolerate a pH range from 5.5 to 8. Compared to most other grain crops, *Brassica* oilseed crops require greater nutrient inputs to achieve high yields. Generally speaking, they need about 25% more nitrogen (N), phosphorus (P) and potassium (K) and up to 5 times more sulphur (S) than a wheat crop. Harvested seed should be stored at no more than 9% moisture when cooled to 10°C to prevent deterioration due to fungal and/or insect activity. The usual rotation is as a break crop with cereals. Wheat yields following a *B. napus* crop invariably improve in Europe and Australia due the reduced level of cereal pathogens present and the control of grassy weeds (Almond *et al.*, 1986).

1.3.4.2 North and South America

62. The oilseed rape/canola grown in North America is concentrated in the northern part of the Western Great Plains (Figure 26). The species and form grown is almost exclusively the spring or annual *B. napus*. In western Canada, less than 1% of the 5 million ha is sown to spring *B. rapa*. Production of the winter or biennial form of *B. napus* in North America is confined to a few thousand hectares in the Province of Ontario, Canada and a few west and central states in the United States. In South America, both spring and winter *B. napus* is produced on some 17,000 ha in central Chile.

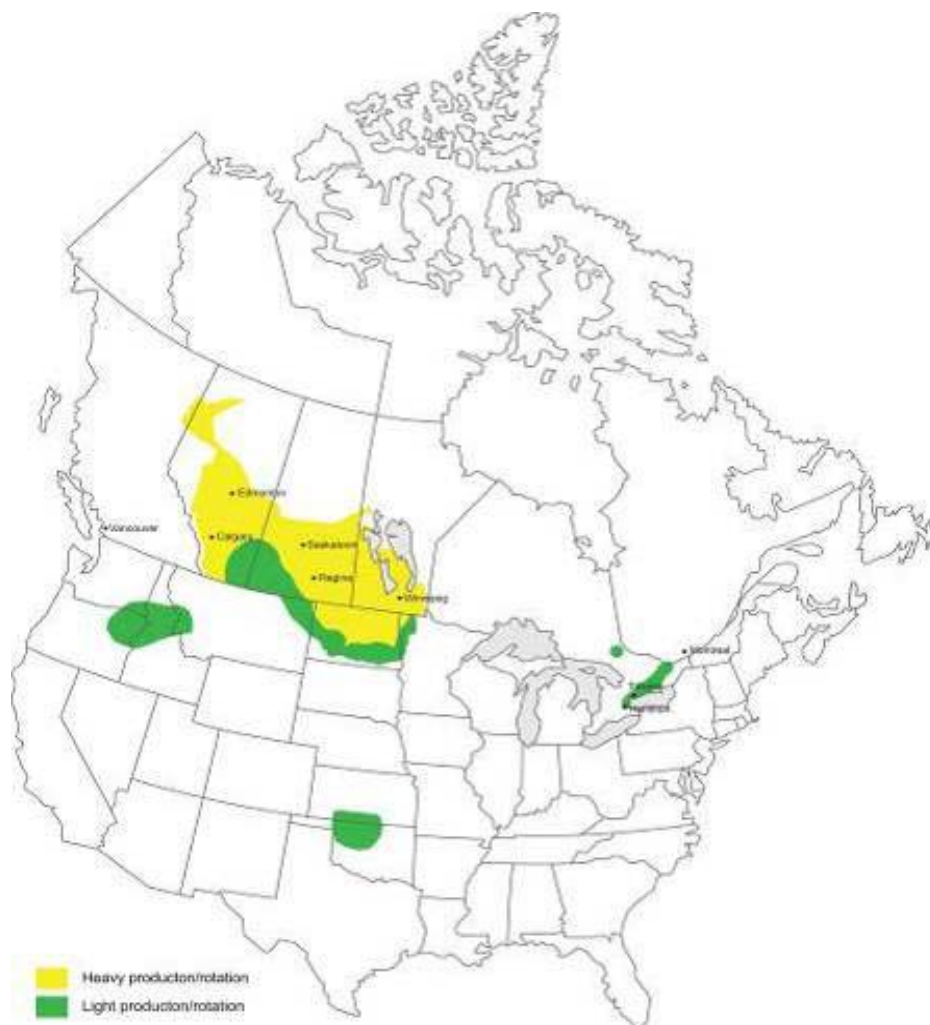


Figure 26. Areas of oilseed rape/canola production in North America
 Source: courtesy Canola Council of Canada
 Yellow (light grey) indicates heavier production concentration

63. Cultural practices in the main oilseed rape production regions of western Canada and the United States have changed in recent years. Traditionally the crop was sown into summer fallow, land laid fallow the previous year. With the shift to continuous cropping and minimum tillage, *B. napus* is now sown into the undisturbed stubble of the previous year’s cereal crop. Weed control, which would normally be a problem with this direct seeding system, can now be easily achieved with the new broad spectrum, post emergence herbicides such as glyphosate, glufosinate and the imidazolinones. The adoption of these herbicides and their associated herbicide resistant varieties has been extremely rapid (Figure 27).

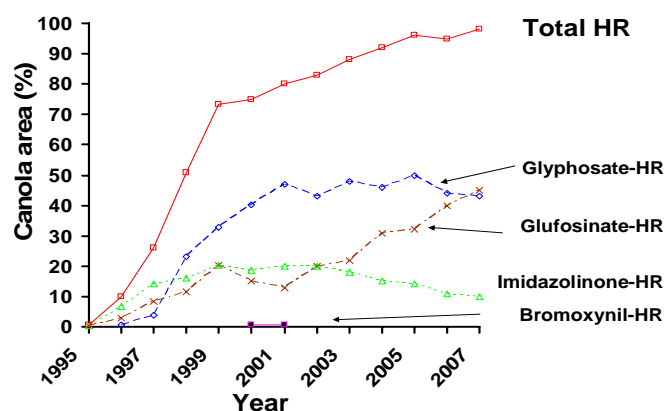


Figure 27. Percentage of the total Canadian *B. napus* production area sown to herbicide-resistant varieties (1995-2008) (HR=Herbicide Resistant)

64. The double disc grain drill has now been largely replaced by large air seeders that place the seed and fertilizer some 2 to 3 cm below the soil surface, at a seeding rate of 5 to 8 kg per hectare. Seed is treated with an insecticide-fungicide coating. The herbicide glyphosate is usually spring applied prior to seeding to control early germinated or biennial weeds. In North America, seeding generally occurs in early May. The herbicide of choice is applied at the recommended rate when the weeds are small and the leaves of the *B. napus* plants have not fully covered the ground. *B. rapa* fields begin flowering in mid June while *B. napus* fields begin to flower about two weeks later in late June or early July. Recommended fungicides and/or insecticides may be applied as a spray if the pest incidence warrants. At harvest, in late August through September, the crop is normally swathed into windrows to allow more uniform ripening and protect against seed losses due to pod shatter. Combining the swaths is done when the seed is mature and dry. However, some straight combining of the standing crop is also practiced. Usually the seed is farm stored at less than 9% moisture until marketed.

65. Chile is the only country in South America that produces a significant quantity of oilseed rape with planting of winter and spring *B. napus* on some 17,000 hectares in the southern provinces of the central part of the country. The crop is predominantly winter *B. napus*. The winter crop is sown in March and April, flowers in October and November and is harvested by straight combining in January. Winter kill may occur in May or June due to the wet soil freezing and heaving, causing broken roots. The spring crop is sown in August-September, flowers in October and is harvested in late December or early January. The crop is normally sown on land broken out of grass pasture using a disk or mould board plough, disked twice with a double disk cultivator and packed. Seed is sown with a double disk seeder or the less satisfactory one way disk at 7 to 8 kg per hectare. Fertilizer requirements vary widely due to the sharply different soil types encountered in rapeseed growing areas. Levels of macronutrients N, P and, in some soils, S are very low. Also lacking in some soils are the micronutrients manganese, copper, and boron. At harvest, a desiccant is applied and after the appropriate interval, the crop is straight combined. The seed is normally artificially dried to less than 9% moisture prior to storage or marketing.

1.3.4.3 European crop cultivation and management

66. Winter oilseed rape (*B. napus*) is the dominant species grown in both Western and Eastern Europe (Poland, Western Russia and the Ukraine); however, the area sown to spring *B. napus* is rapidly expanding (Figure 28). Some spring and winter *B. rapa* is grown in Sweden and Finland. In Germany over the past 13 years the area sown to spring oilseed rape decreased from 10% to 1% of the oilseed rape growing area. Spring *B. napus* is used primarily as a replacement crop on winter oilseed rape fields that have been winter-killed. The optimum date for sowing the winter form varies with the latitude and

the onset of winter. In Northern European countries the optimum sowing date is the last half of August while more southerly regions in France and Germany can delay seeding until early September. The objective is to produce plants that are large enough and have stored sufficient food reserves to withstand the rigors of winter. It is recommended that plants entering the winter show a vigorous growth, a well developed root system (taproot about 8 to 10 mm in diameter) and have at least six to eight true leaves. Seed is sown into well worked soil at 5.0 to 5.5 kg per hectare when drilled and 8 to 9 kg per hectare if broadcast, to obtain fall stands of 50 to 85 plants/m² to allow for some winter kill.

67. The seeding rates recommended for precision drilled hybrid varieties with a high branching density and a 1000 seed weight of 4 g/1000 is 1.2 to 1.6 kg per hectare and for seed of 7 g/1000, 2.1 to 2.8 kg per hectare. The seeding rates for drilled hybrids are lower than for open pollinated varieties since the hybrid seed is likely to produce a more vigorous plant that better withstands the winter. The optimum spring plant population is reported to be 80 to 100 plants/m². Winter varieties are heavy users of nitrogen so frequently some nitrogen is incorporated prior to planting, with the balance top-dressed in the spring. Excessive nitrogen promotes vigorous fall growth but tends to make the crop more susceptible to winter kill. Phosphorus and potassium are applied before planting at the recommended levels. Sulfur is used in early spring in combination with N-fertilization. Boron is often applied in late spring in combination with fungicides. Nearly all seed is treated with a fungicide- insecticide combination (often with more than two active ingredients) to control seedling pests.

68. Disease, insect and weed control in the emerged crop is achieved by spraying the recommended products when needed. Flowering in Northern Europe begins the last days in April, and harvest starts with some swathing at the end of July with the vast majority of the crop straight combined a week or so later. Harvest can continue through to the end of August. In southern regions, harvest commences about one to four weeks earlier.

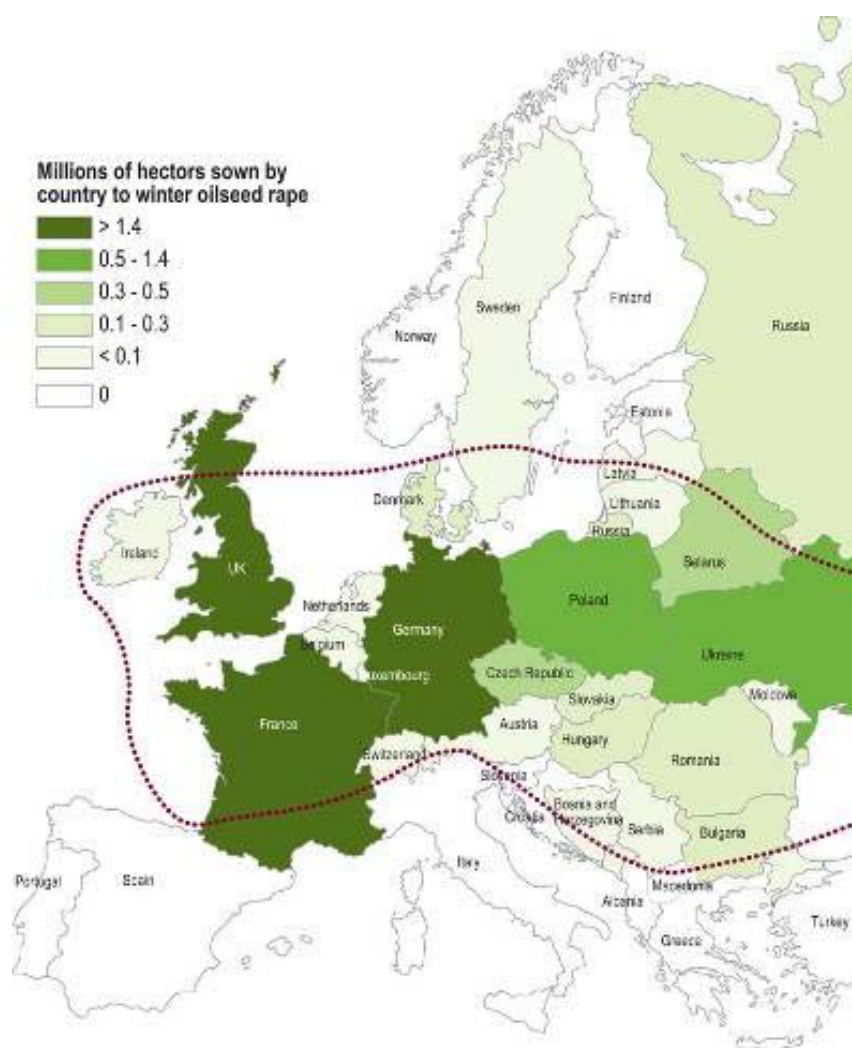


Figure 28. Oilseed rape (*B. napus*) production regions in Europe showing millions of hectares of winter rape per country

The dotted line encircles the primary growing region for winter oilseed rape.

Spring rape production is concentrated in Eastern Europe primarily Russia (>0.5 M ha) and the Ukraine (0.1-0.5 M ha). (Information supplied by Norddeutsche Pflanzenzucht)

1.3.4.4 Australia crop cultivation and management

69. Oilseed rape production in Australia is relatively recent with the first commercial production undertaken in 1969. In the early years both *B. rapa* and *B. napus* spring varieties from Canada were imported and grown in the winter season. Today production is almost exclusively from Australian bred *B. napus* varieties. Canola is grown in most cropping areas of Southern Australia, including Western Australia (Figure 29). Most of the *B. napus* crop is sown in late autumn or early winter (April to June) during the rainy period. The seed is primarily sown with air seeders at seeding rates of 4 to 6 kg per hectare with hybrid varieties being sown at about 3 kg per hectare. All seed is treated to control blackleg disease [*Leptosphaeria maculans* (Desm.) Ces. et de Not.] and some seed is treated for control of the red-legged earth mite (*Halotydeus destructor* Tucker). Flowering occurs in August and September with harvest in late spring or early summer (November and December). The growing season ranges from about 150 to 210 days, depending on latitude, rainfall, temperature and sowing date. Growth and yield of the crop is almost always limited by the amount of water available to the crop, particularly during maturation.

70. Due to the age of Australian soils, macronutrients (particularly nitrogen, phosphorous and sulphur) and micronutrients are deficient. Deficiencies in boron, manganese, molybdenum, and zinc have been reported for *B. napus* crops, as has toxicity on the more acid soils due to high levels of aluminium and manganese. Most soils are strongly acidic and liming is necessary to achieve high yields. Initially oilseed rape was sown into well worked soil but with the availability of glyphosate as a pre-planting herbicide and varieties resistant to triazine and imidazolinone herbicides, direct seeding has become standard practice. Oilseed rape is most frequently preceded by a pulse crop or pasture while fallow and wheat are other alternatives. When the canola crop precedes wheat in the rotation, substantial wheat yield benefits occur.

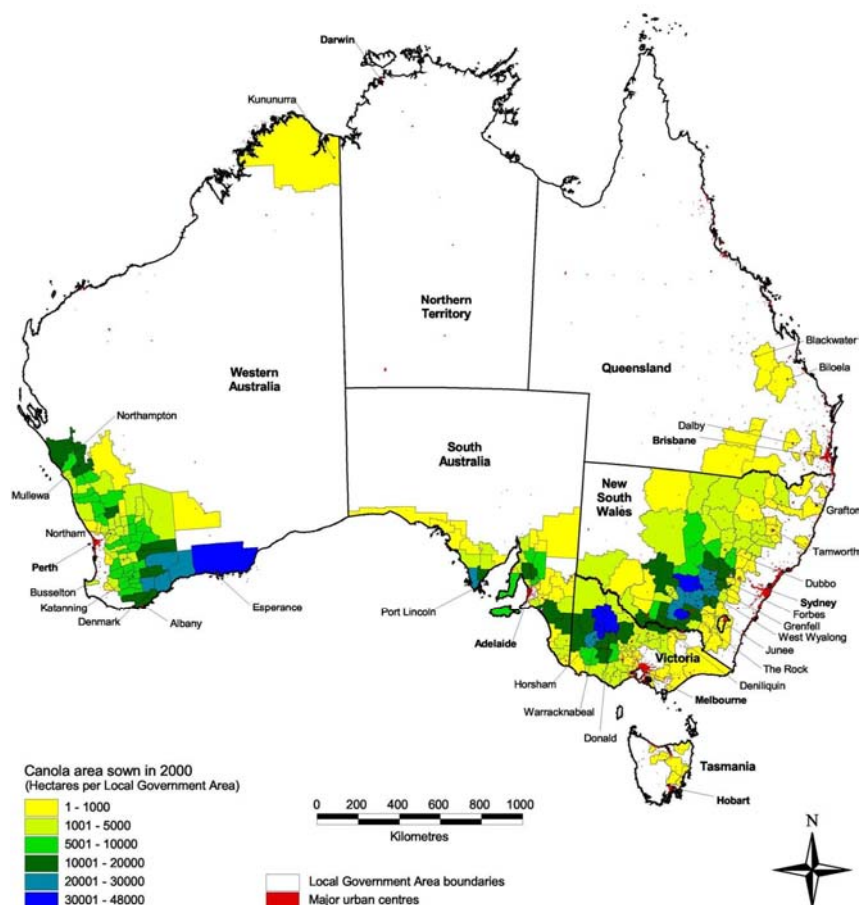


Figure 29. Areas and concentration of *B. napus* production by Australian government districts
Data source: Canola statistics, Australian Bureau of Statistics, Agricultural Census 2001 (published 2002)

1.3.4.5 Indian sub-continent cultivation and management

71. The dominant *Brassica* oilseed crop on the Indian sub-continent is *B. juncea* although a limited hectareage is sown to the *B. rapa* form, toria, which is grown in the September through December period in northern areas. *B. napus* and *B. carinata* are grown to a limited extent in some irrigated and dry land areas of northern and central Indian states, respectively. The major crop of *B. juncea* as well as small pockets of Yellow and Brown Sarson (forms of *B. rapa*) are sown in October or early November and harvested in late March or early April. Flowering occurs in early January. Production is centred in the northern half of the sub-continent, in what is called the mustard belt (Figure 30). The untreated seed is normally broadcast on the ploughed and levelled fields and the seed buried by drawing a heavy plank over the field. The traditional practice of sowing the *Brassica* species mixed with a cereal grain is no longer employed to any degree and the sowing of pure stands of each crop is now normal practice.

However, mixed cropping is still practiced in several areas by few farmers. Double cropping in the mustard belt is the standard practice with mustard sown on the same land each year following the summer crop which may be pulses (mung and urd bean) or green manure. Other alternatives are rice, cotton or millets (such as sorghum or pearl millet). The recommended seeding rate for *B. juncea* is 4 to 5 kg per hectare. Fertilization with nitrogen-phosphorous-potassium, in the ratio of 80-40-40 kg per hectare, together with 40 kg of zinc and 25 kg of sulphur, is recommended.



Figure 30. Major production region (striped area) of oilseed mustard (*B. juncea*) and toria (*B. rapa*) on the Indian sub-continent

1.3.4.6 China cultivation and management (Wang *et al.*, 2007a)

72. China is the world's largest producer of *Brassica* oilseed crops, annually producing some 11.5 million tonnes. Species contributing to this output include winter and spring *B. napus* (95%), *B. juncea* (4%) and both winter and spring forms of *B. rapa* (1%). The location and concentration, by province, where *B. juncea* and *B. rapa* are grown is illustrated in Figure 31.



Figure 31. Brassica oilseeds average yield (kg/ha) and production (metric tons) by province in China
 Production is primarily (95%) from *B. napus* but both *B. juncea* (4%) and *B. rapa* (1%) oilseed crops are also produced in the concentration and provinces indicated.

73. *B. napus* is grown throughout the country with the winter form dominating in the southern provinces and the spring form in the north (Figure 32).

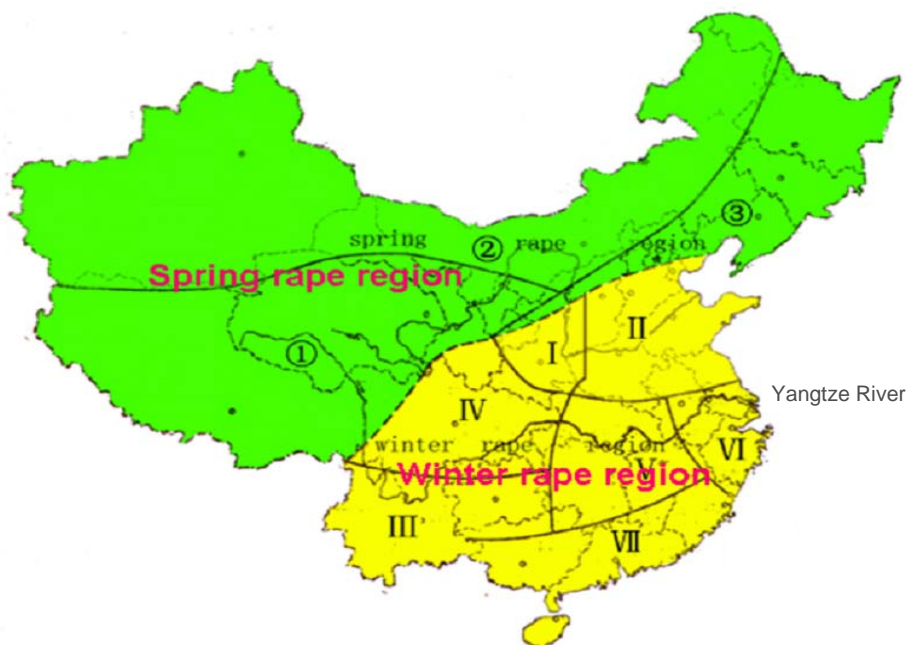


Figure 32. Regions sown to the spring and winter forms of *B. napus* in China
 Map numerals designate environmental regions where cultivars with certain agronomic conditions are best adapted

74. The provinces along the Yangtze River provide the bulk of China's production (Figures 31 and 32). The level of winter hardiness required is not great. Indeed Canadian and European varieties of the spring form have successfully survived the winters in the Chinese winter growing region.

75. The spring-sown crops (*B. napus*, *B. juncea* and *B. rapa*) are sown in May, flower in June or early July and are harvested in September. The growing cycle for *B. napus* takes about 120 days. In the southern portion of the spring growing area, half a season may be used to grow a forage or vegetable in conjunction with *B. rapa*. Because of the small field sizes, most are sown by hand or walking plough, although some large fields are mechanically sown. In the winter rape areas the seed is sown into small seedling beds in September and the seedlings later transplanted into the production fields in mid to late September. Flowering takes place in late March and harvest is in May. The total production cycle is about 220 days. The rotation in the triple cropping winter rape area is either rape-rice-rice or rape-maize-potato and in the double cropping regions rape-cotton or rape-rice.

76. Soil fertility is a limiting factor in production, with the area devoted to winter rape being particularly deficient in P. While all soils require N, P, and K, significant areas are deficient in the micronutrients Zinc (Zn) and Boron (B), while shortages of Manganese (Mn), Copper (Cu) and Iron (Fe) also occur.

1.3.4.7 Herbicide resistant *B. napus*

77. *B. napus* is not considered a significant weed in managed ecosystems (AAFC, 1994). However, due to the high level of seed lost during harvest it can be an abundant weed in subsequent crops. Légère *et al.* (2001) ranked *B. napus* as 18th in relative abundance among Canadian weed species in western Canada, and Leeson *et al.* (2005) found *B. napus* plants in 10.5% of the fields surveyed. Studies in both Canada and Europe have shown that the incorporation of genes for resistance to specific herbicides imparts no altered weediness or invasive potential for glyphosate, including different events (AAFC, 1995b, 1996a; Norris *et al.*, 1999; Crawley *et al.*, 2001); glufosinate-ammonium, including its combination with the hybrid system (AAFC, 1995a,d, 1996b; Rasche and Gadsby, 1997; Norris *et al.*, 1999; MacDonald and Kuntz, 2000); bromoxynil (PBO, 1998) and non-GM imidazolinone (AAFC, 1995c). Experience in western Canada from 1995 through to the present (2011), with all HR systems, have confirmed the validity of these earlier assessments (Beckie *et al.*, 2006; Warwick *et al.*, 2009a).

78. However, GM-HR volunteers can occur in subsequent *B. napus* crops. The level will depend on the interval between oilseed rape crops in the rotation and how well the producer has controlled volunteer *B. napus* in the intervening years. The shorter the rotation and the less volunteer control, the greater the contamination level in the second planting. The presence of one GM-HR canola plant per m² throughout a field of conventional oilseed rape calculates to a GM content of 2.5% in the harvested conventional crop (planted at 40 plants/m²). This calculation assumes that the numbers of seeds produced by a volunteer plant is the same as that produced by the conventional plants (CETIOM, 2000). However, Gruber and Claupein (2007) report that volunteer winter *B. napus* plants, growing in a sown rapeseed crop, only yield 45% of the seed produced by corresponding sown plants.

79. Off-type volunteer plants can come from multiple sources, including the seed bank from previous crops, movement of farm equipment and animals, pollen flow and contaminated seed stocks. In Australia, Stanton *et al.* (2002) found sheep can excrete viable or germinable *B. napus* seed up to 5 days after ingestion. Similarly, Martens (2001) claimed that manure from oilseed rape-fed chickens resulted in volunteer plants when the manure was spread on a field 12 months later. In Canada, Downey and Beckie (2002) and Friesen *et al.* (2003) found certified pedigreed seed lots of conventional varieties contained unacceptable levels of GM seeds, apparently resulting from pollen flow in breeding nurseries. The seed industry quickly purified their breeding stocks but absolute exclusion cannot be guaranteed.

Feral populations may disseminate genes to nearby oilseed rape crops but the incidence would be very small and far less than several of the sources noted above (CETIOM, 2000; Wilkinson *et al.*, 1995).

80. In all oilseed rape growing regions, leaving the soil untilled for a period after harvest and using non-inversion tillage is an effective strategy for minimizing the size of the seed bank (Gruber and Claupein, 2007; Gulden *et al.*, 2003a). Ploughing, as done in Europe, will bury the seeds below germination depth but when the field is again ploughed the dormant seeds will be brought to the surface. Pre-emergence and in-crop post emergence herbicide applications are effective in controlling volunteers even if they contain one, two or three different herbicide resistance genes (Table 5) (Downey and Buth, 2003). In western Canada, where herbicide tolerant oilseed rape has been grown extensively for 15 years, there is no evidence that volunteer *B. napus* has increased or is more prevalent because of the herbicide resistance traits (Hall *et al.*, 2000; Beckie *et al.*, 2004, 2006).

Table 5. Number of herbicide products available for control of volunteer *B. napus* with nil, single, or multiple herbicide tolerances in western Canada

Herbicide system	Number of products
Susceptible	27
Liberty Link (LL) ¹	26
Roundup Ready (RR) ²	25
Clearfield (CF) ³	19
RR × LL	24
RR × CF	17
LL × CF	18
RR × LL × CF	16

¹LL Glufosinate, ²RR Glyphosate, ³CF Imidazolinone

Source: Downey and Buth, 2003

1.3.4.8 Oilseed certified seed production

81. The production of oilseed *Brassica* sowing seed is normally undertaken within the areas where the *Brassica* crop is commercially grown. The rules under which pedigreed seed is produced and identified in the market place are stringent and extensive. Regulations vary from country to country but the minimum requirements for certified seed moving in international trade are governed by two international certification organisations. Both the OECD Seed Schemes and the Association of Official Seed Certifying Agencies (AOSCA) were developed to facilitate seed trade through mutual recognition of the official certification labels of member agencies. Member countries must meet OECD and AOSCA standards, but countries can and most of them do have domestic certification standards that exceed those minimums.

82. AOSCA has a United States focus but its members include also Canada, Argentina, Chile, New Zealand, Australia and South Africa. AOSCA standards cover not only varietal certification of seed but also germination, physical purity, disease and other quality traits. Their varietal certification requirements include a maximum variety impurity “seed” standard that is used for post-control verification testing.

83. The OECD Seed Schemes, which largely reflect the requirements of the European Union seed certification system, are increasingly implemented at the global level. They comprise 58 member countries including most of the countries discussed above. China and Pakistan are currently not members of the OECD Seed Schemes (situation November 2012). However, China is developing standards for *Brassica* crops, and Pakistan has regulations that are similar to those of India. OECD seed standards do not deal with germination or physical purity but focus on varietal certification, based mainly on morphological characteristics during inspections of seed production crops. In addition, minimum requirements and standards for verification, using post-control field testing, are mandatory.

84. Seed classes allowed are normally designated by the breeder or maintainer of the variety. For *Brassica* oilseed, the seed multiplication factor for each generation is typically large (> 1,000:1). Thus the seed classes designated for its species are normally limited to three and identified under the OECD Seed Schemes as Basic, Certified 1st and Certified 2nd generations, with the equivalent generations designated under AOSCA as Breeder, Foundation and Certified Seed. Normally only one generation is allowed for the foundation and certified classes. The OECD seed regulations for Brassicaceae oilseeds require a 5-year interval between crops of the same species. AOSCA standards for production of foundation seed of *B. napus*, *B. juncea* and *B. rapa* require 4 years between crops of these species and a 2-year interval when producing certified seed. Under OECD regulations basic and certified seed production fields of *B. napus* must be isolated from any possible source of cross-pollinating pollen by a minimum of 200 m and 100 m, respectively. AOSCA regulations require foundation producing fields of *B. napus*, *B. juncea* and *B. rapa* to be isolated from any other crop of the same kind by 201 m, 402 m and 402 m respectively. For certified producing fields of these three species, the respective isolation distance required is 100 m, 402 m and 100 m. Both sets of regulations require all seed production fields to be inspected by the designated authority at least three times for Basic Seed production and three times on each parental line for the production of certified seed of hybrid varieties, *i.e.* before the flowering stage, in the early flowering stage, and before the end of the flowering stage. Fields must also meet stringent standards for varietal purity (visual characteristics) as well as freedom from cross-pollinating species and other crop kinds.

85. It must be emphasized that the above are minimum standards with most countries having higher requirements as well as many seed companies exceeding the more stringent domestic regulations. Open-pollinated varieties of *B. napus* are rapidly being replaced by F₁ hybrid varieties, and a similar situation is likely to occur in *B. juncea* within the next few years. The requirement for nearly absolute purity of the female parent is mandatory if the hybrid is to produce the desired level of heterosis. The male restorer parent must also breed true for restoration of hybrid fertility. Thus the hybrid regulations for isolation distances under AOSCA are much greater at 804 m while most seed companies use 1,000 m or more. Also, foundation and certified producing crops for hybrid seed production cannot be grown on land which has grown *B. napus*, *B. rapa*, *B. juncea* or oilseed *R. sativa* in the past 5 and 3 years, respectfully.

86. The studies by Downey and Beckie (2002) and Friesen *et al.* (2003) that identified some Canadian certified *B. napus* seed lots as containing undesirable levels of foreign herbicide resistance traits are often cited as sources of contamination. Regulators and the seed industry moved quickly to correct this situation. Today the Canadian Food Inspection Agency (CFIA) carries out seed testing of *Brassica* oilseed varieties for a) adventitious presence (AP) of approved events and b) for herbicide trait purity of glyphosate and glufosinate ammonium resistant varieties. All official reference control samples for oilseed rape varieties submitted to the CFIA's Variety Registration Office at the time of registration of a new variety are subject to AP testing and if the variety is herbicide resistant, to herbicide purity trait testing. Furthermore, the CFIA also monitors AP and trait purity of foundation and certified seed. In instances where AP and/or trait purity issues are identified, the breeder of the variety is notified and appropriate action is taken (CFIA, 2009).

87. The Canadian Seed Growers Association (CSGA) have also revised their 'Regulations and Procedures for Breeder Seed Crop Production' so that seed certificates are only issued for Breeder seed crops that are produced within a third party audited Quality Management System (QMS) and verified to preserve varietal identity. Further, non-compliance with QMS requirements can lead to suspension or cancellation of the professional recognition of a Plant Breeder which is required in both CFIA variety registration and CSGA seed crop certification.

1.3.5 *Brassica vegetable seed production locations and management*

88. The market for *Brassica* vegetables has, in recent years, experienced a steady increase in demand. This expansion has been aided by widespread refrigerated transportation systems that can provide a year round supply of such vegetables to most markets. The *Brassica* vegetable crop with the greatest demand for seed is cabbage, followed by the *B. rapa* Asian vegetables and broccoli. The world requirement for cauliflower seed is less while the demand for turnip, rutabagas and kohlrabi is relatively small. Accompanying the increased commercialisation of *Brassica* vegetable production has been the need to provide large quantities of seed of high quality and varietal purity. This requirement has resulted in the majority of the seed being produced in specific locations where climate and isolation from other *Brassica* crops are favourable for consistent high yield and quality. To aid the growing international trade in vegetable seed the OECD has established a Scheme for the Certification or Control of Vegetable Seed which requires field and seed inspection by an accredited authority, within the country of origin, to ensure the seed meets varietal purity standards including freedom from cross-pollinating species. The OECD Vegetable Seed Scheme provides for the production of "Certified Seed", and the designation of "Standard Seed", corresponding to two different control requirements. Other organisations that facilitate the seed trade include the International Seed Federation (ISF) that has defined trading terms and rules dealing with sales, can arbitrate settlements and assists with import and export licenses: ISF regional seed industry organisations such as the Asia and Pacific Seed Association (APSA) seeks to improve vegetable seed production and trade in the region (George, 2009). Many companies also use a Quality Management System as described above for oilseed seed production.

1.3.5.1 **Locations of concentrated vegetable seed production**

89. In developing countries vegetable seed is primarily supplied from farm saved seed, and more rarely from the formally-organised seed sector. In countries with strong agricultural and horticultural industries, nearly all the seed is from commercial pedigreed sources. For large scale seed production of the biennial *Brassica* vegetables, seed companies have concentrated production in areas with relatively mild winters and moderate summer temperatures. In Europe such areas are found in Brittany (France), the Netherlands, Belgium and northern Italy.

90. In North America, among the *Brassica* vegetables, broccoli has the greatest seed demand followed by cabbage and cauliflower. The market for the seeds of collard, Brussels sprouts and the Asian vegetables is much smaller. Seed production of these crops is concentrated in valleys of Oregon and Washington states, USA (e.g. Oregon's Willamette Valley). Selected areas in California and Arizona are also important producers of broccoli and cauliflower seed. Essentially all broccoli and cabbage varieties produced in the USA are F₁ hybrids. In contrast, most cauliflower varieties are highly inbred and uniform, self-pollinating populations but in recent years more and more F₁ hybrids have entered the market (Farnham, 2007). F₁ and inbred varieties of collards, Brussels sprouts and kale provide seed to the commercial market. In South America, Chile is a significant supplier of vegetable *Brassica* seed.

91. In Australia, seed production of *Brassica* vegetable crops is centred in Tasmania in the regions of the Coal River Valley, Derwent Valley, central East Coast, Hagley/Westbury and Devonport. Tasmania is climatically suited for 'counter-season' seed production for the Northern Hemisphere markets of Asia and Europe. Major seed crops produced in 2001 were hybrid cabbage (150 ha) and cauliflower (97 ha) (Government of Tasmania, 2003). Cabbage and cauliflower are high value autumn planted crops while the lower value mustard and Chinese vegetable types are spring sown. Locations for hybrid seed production of cabbage and cauliflower are determined by the need for an isolation zone of 1.5 to 3 km from other crops of the same botanical family. Grower awareness and consultation between companies ensures adequate isolation distances.

92. In New Zealand the Canterbury Plains and other smaller areas of the South Island (43° south) have become a major vegetable seed production location, particularly for the Asian *Brassica* vegetables. In this region the seed merchants and growers have put in place an isolation mapping system to avoid cross pollination among different species and varieties. The system is operated by a government-owned company called AgriQuality that displays an internet map of every farm field involved in seed production. When a seed contract is arranged and a field is selected, the seed merchant logs the details into the system and can see if there are any conflicts within the isolation distance required. Normal minimum isolation distance for the *Brassica* crops is 1000 m but that can be extended, particularly with hybrid seed production.

93. Not all seed production regions are maritime based. In China cauliflower and broccoli seed production is concentrated in semi-desert regions around the cities of Jiuquan and Jiayuguan in Gansu and in Yunnan provinces. In these high elevation areas precipitation is minimal, but irrigation is available and the temperatures remain within the required range. Cabbage and Chinese cabbage seed production is located further south in Hebei, Henan, Shandong and Shanxi provinces. (X-W. Wang, Pers. Comm.). In these regions the normal isolation distances between production fields is 1000 m.

94. On the Indian sub-continent no concentrated areas for seed production were identified. However, small individual fields occur scattered in the foothill valleys of the Himalayas. For the production of certified cauliflower seed the minimum isolation distance is 1,000 m (Indian Minimum Certification Standards).

95. In Japan, no concentrated area exists for large scale seed production. However, various *Brassica* vegetables (*B. rapa* and *B. juncea*) are cultivated locally (Inomata, 2007) and seed production is practiced on a small scale. The minimum isolation distance required for seed production is 600m.

96. George (2009) notes that most authorities recommend having a greater distance (up to 1500 m) between different types of *B. oleracea* (cabbage vs. kohlrabi) than between varieties of the same type (two cabbage varieties, up to 1000 m).

1.3.5.2 Vegetable seed cultivation and management

97. The optimum pH for cole crops is reported to be 6.0 to 6.5 with the generally recommended ratio of N-P-K nutrients being 1:2:2 at soil preparation, but it varies depending on the production region (George, 2009). The lower ratio of nitrogen is to avoid 'soft plants' that are less winter hardy. Extra nitrogen is normally topdressed in the spring. It is important to ensure adequate levels of sulphur as well as the micronutrients boron, manganese and molybdenum are available. The development of hybrids in Brussels sprouts has become very important (George, 2009) with self-incompatible and CMS (cytoplasmic male sterility) hybrids becoming more frequent in cabbage, cauliflower, kale and kohlrabi. The ratio of male to female in hybrid production fields is normally 1:1 or 1:2 (Takahashi, 1987).

98. Most cabbage and cole crops and some Asian vegetables are biennials and will not bolt until they have been exposed to temperatures of 4°C to 7°C for six to eight weeks. Day length has no effect on bolting or flower initiation (Nieuwhof, 1969). At the end of the first year cabbage plants can withstand temperatures of -12°C to -14°C for extended periods, but lower temperatures can cause much damage as can alternating periods of frost and thaw (Nieuwhof, 1969). The usual practice in producing cabbage seed is to sow in the summer with the plants over wintering, bolting in the spring and harvest the seed in summer. Cultivars differ in their winter hardiness with red cabbages the least hardy and savoy the hardiest. Summer temperatures are also important in determining seed yield. Temperatures above 25°C arrest growth and cause seed abortion. Because of these environmental constraints commercial production tends to be concentrated in areas with mild winters, sufficiently cold to ensure vernalization without winter kill, combined with moderate summer temperatures. The availability of irrigation is also important to obtaining uniform high yields.

99. Seeding of the biennial crops in the northern hemisphere is normally done in mid-June to mid-August. If the seed is to be sown in beds for transplanting, rather than direct seeding into the field, seeding should be done about 10 days earlier than the field sowing to allow for the plant setback brought on by transplanting (Nieuwhof, 1969). The recommended rate for field sowing is 3 to 5 kg per hectare, unless precision sowing is practiced where only 1 to 2 kg per hectare is needed. Plants are thinned to 35 to 40 cm between plants within the row. To increase the over wintering survival rate, plants may be earthed up covering the most sensitive plant portion just below the head. Weed control is critical, as in mild winters weeds may over grow the crop. Most of the cole crops are self-incompatible and depend on insects, primarily honey bees, to effect fertilization. Harvesting is done once the pods have turned yellow and the seeds turned brown. Depending on field size and seed value, harvesting may be done by various methods from hand cutting and threshing to straight combining. Kohlrabi, although a true biennial, can be vernalized by initiating germination through pre-soaking the seed for 8 to 9 hours at 20°C followed by a cold treatment of -1°C for 35 to 50 days. The treated seed can then be sown directly into the field in the spring and the seed crop harvested in the fall. Brussels sprouts and kale are grown for seed in the same manner as cabbages.

100. For cauliflower and broccoli crops, only a mild vernalization period is required so environmental limitations are less stringent. However, as a seed crop these forms normally require an extended growing season. Selection of cauliflower varieties for a tighter curd has resulted in slow and incomplete bolting, thus further extending the required growing season. In Western Europe cauliflower is sown in September and over-wintered under glass with transplanting to the field in early spring. Transplants are spaced on a 50 × 50 cm or smaller grid. Flowering occurs in July or August and the crop is harvested in September or early October. Seed production of tropical and subtropical cauliflower is discussed by Lal (1993).

101. Drying the harvested *Brassica* vegetable seed is frequently required. To maintain germination capacity the maximum air drying temperature should not exceed 60°C. If seed is to be stored for a year, maximum moisture content should not exceed 9% with a storage temperatures of 5 to 10°C.

102. The biennial turnips and Swedes (rutabaga) regenerate from growing points at or near ground level. This means they can benefit from a large underground source of nutrients for seed production. Thus these crops are more winter hardy than cole crops and can be grown for seed over a much wider environmental range. However the market for their seed is relatively small, so seed companies tend to contract their production with growers in areas already producing seed of other *Brassica* vegetable crops.

103. The *B. rapa* vegetables prefer a soil pH between 6.0 and 7.5 with an N-P-K fertilization ratio at planting of 2:1:1. Additional nitrogen fertilizer is normally applied at anthesis (George, 2009). The seed is produced by either the head-to-seed or the seed-to-seed method described by Opeña *et al.* (1988). As with the cole crops the ratio of male to female in hybrid production fields is 1:1 or 1:2 (Takahashi, 1987).

1.3.6 Centres of origin and ancestors

1.3.6.1 Introduction

104. There are few areas of the world where members of the family Brassicaceae are totally absent. The exceptions are parts of the tropics, where the family is thinly represented, but where some introduced cosmopolitan weeds have become established. The genera and species of the family occur in greatest number and diversity in the temperate zone of the northern hemisphere and in particular, the areas surrounding the Mediterranean basin and throughout the southwest and central regions of Asia (Figure 25) (Hedge, 1976). Although the generic and specific endemism in the family is highest in the Irano-Turanian region, the centre of origin of the current subtribe Brassicinae, lies in the Mediterranean basin (Hedge, 1976).

105. Using chloroplast DNA restriction sites together with cpDNA probes, Warwick and Black (1991) surveyed 33 diploid taxa of the Brassicinae. The phylogenetic results indicated there were clearly two ancient and distinct evolutionary lineages within the subtribe. They found the “Nigra” lineage to include *B. nigra*, *B. fruticulosa*, *B. tournefortii*, *Sinapis pubescens*, *S. alba*, *S. flexuosa*, *S. arvensis*, *Coincya cheiranthos*, *Erucastrum canariense* and *Hirschfeldia incana*. The other lineage, termed “Rapa/Oleracea”, was made up of *Brassica rapa*, *B. oleracea* and subsp. *alboglabra*, the *B. rupestris-villosa* complex (*B. rupestris*, *B. drepanensis*, *B. macrocarpa*, *B. villosa*), *B. barrelieri*, *B. deflexa*, *B. oxyrrhina*, *B. gravinae*, *Diptotaxis eruroides*, *D. tenuifolia*, *Eruca sativa*, *Raphanus raphanistrum*, *R. sativus* and *Sinapis aucheri*. In the “Nigra” lineage *B. nigra* was most closely related to the annual *Sinapis* species *S. arvensis* and *S. alba* (Figure 33). Only a single mutation difference was found between the crop and weedy accessions of *B. rapa* and between crop accessions of *B. oleracea* and wild accessions of *B. oleracea* subsp. *oleracea* and subsp. *alboglabra* (Warwick and Black, 1991). The weedy species *R. raphanistrum* and the crop species *R. sativus* differed by only four mutations.

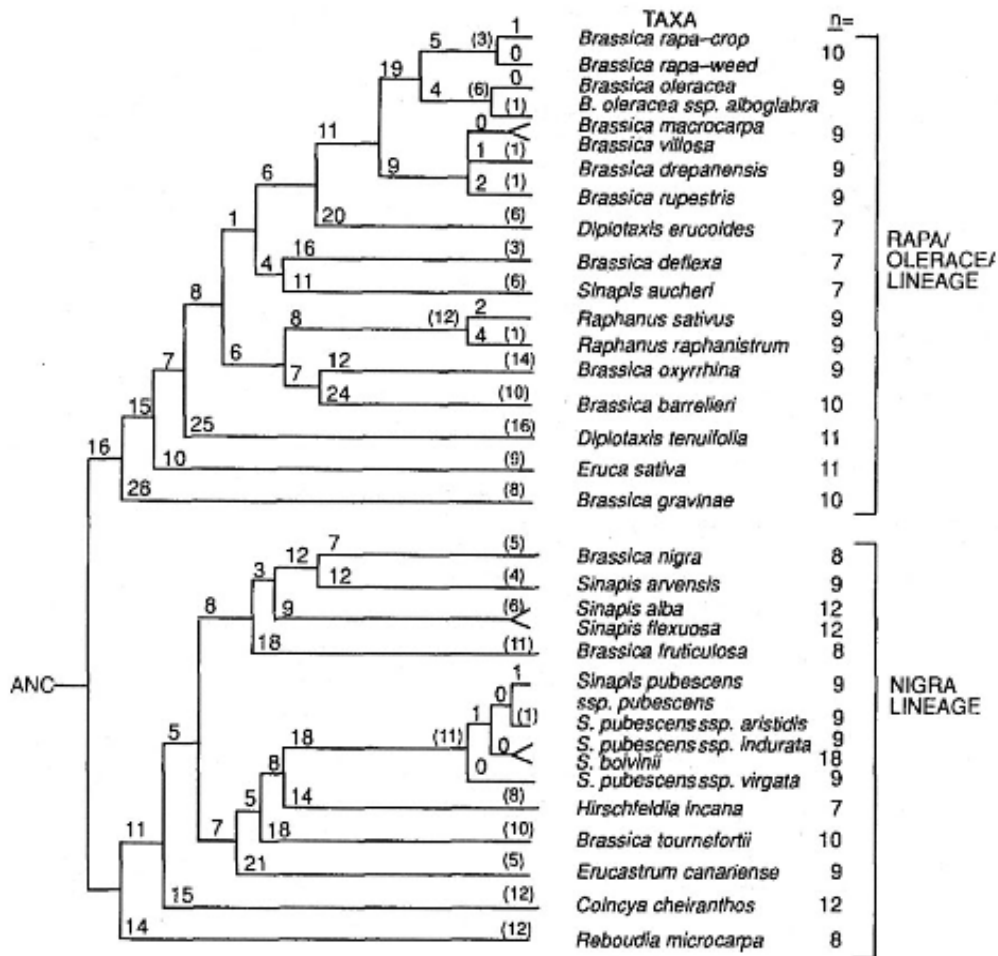


Figure 33. Phylogenetic tree for the subtribe Brassicinae, based on PAUP analyses of the chloroplast DNA restriction site/length mutations shared by two or more taxa/accessions

Source: Warwick and Black, 1991

PAUP is a computational phylogenetics programme for Phylogenetic Analysis Using Parsimony that infers evolutionary trees (phylogenies). Tree length in this tree is 489 steps, consistency index is 0.491. Tree topology indicates how accessions are related, and branch length (numbers above the branches) indicates the minimal number of mutational steps occurring during the evolution of a particular taxa. Mutations unique to a given species and to the genus *Raphanus* (number indicated in brackets at end of branch) should be added to determine terminal branch length. ANC indicates a common hypothetical common ancestor.

106. Although the economically important *Brassica* species arose from ancestors in the Mediterranean region, wars and trade ensured their wide dispersal, resulting in islands of isolated environmental and selection pressure. The earliest widely distributed species were those that exhibited seed dormancy combined with useful traits. Seed dormancy allowed the introduction to survive long after its introduction. The fast growing, weedy type of *B. rapa*, providing lamp oil and animal feed, and *B. nigra* as an oil and spice source would be prime candidates. The Mission trail in Southern California is a case in point: Priests scattered *B. nigra* seed to mark the trail between the early Missions. Parts of those trails can still be seen each year as the black mustard blooms on the California hillsides.

1.3.6.2 *B. nigra*

107. *B. nigra* is amongst the oldest recorded spices, being noted in the Sanskrit writings of about 3000 B.C. as 'Sarshap' (Prakash, 1961). However, little is known about *B. nigra*'s true centre of origin. Hemingway (1995) placed it in Irano-Turanian, Saharo-Sindian region, (Figure 34). However, Prakash and Hinata (1980) favoured an origin in Central and South Europe. Its use as a commercial spice ensured its very early, widespread distribution across Europe, Africa, Asia and India, and its dehiscent siliques ensured its persistence. The crop was grown for the sharply pungent chemical (allyl isothiocyanate) released when the crushed seed was mixed with a small amount of water, in the same way that *B. juncea* powdered mustard is used today. Until the 1950's *B. nigra* was the world's major source of pungent mustard, but because it shatters as soon as the pods are ripe it required hand harvesting. Thus it was replaced in a single decade by highly pungent *B. juncea* varieties well suited to mechanical harvesting. Today there is essentially no commercial production of *B. nigra* and it has become a weed of waste places in many regions. It is an introduced species to the Americas and Australia. It has never become established on the Canadian prairies although it is present throughout much of the United States.

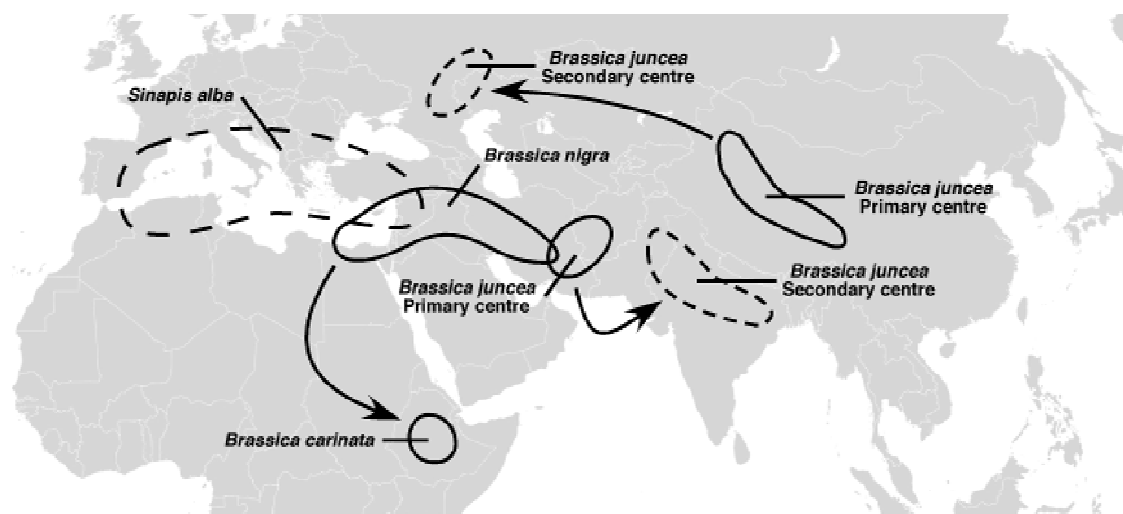


Figure 34. Evolutionary geography of *B. juncea*, *B. carinata* and *Sinapis alba*

Source: greatly modified from Hemingway, 1995

1.3.6.3 *B. rapa*

108. *B. rapa* is generally believed to have originated in the mountainous areas near the Mediterranean sea rather than the coastal areas (Tsunoda, 1980). As with *B. nigra*, *B. rapa* had a wide distribution before recorded history. Indian Sanskrit literature first mentions the plant about 1599 B.C. as 'Siddharth' (Prakash, 1961). Burkill (1930) proposed that the leafy vegetable forms were developed in China from the oilseed form about 2000 years ago. Seeds of both *B. rapa* and *B. juncea* were found in excavations of the ancient village of Banpo, Xian, Shanxi province, China that existed in Neolithic times 6,000 to 7,000

years ago (Liu, 1985). Turnip seeds were also found in pottery jars from the 5th century B.C. at the Yangshao agrological site in Shensi Province (Chang, 1970). Cultivation of *B. rapa* is also mentioned in the oldest collection of Chinese poetry, Shi Jing (The book of Odes), written during the Chunqui period about 535 A.D. (Liu, 1985; Chapman and Wang, 2002). In Scandinavia *B. rapa* seeds were being consumed as early as 350 B.C. as indicated by their presence in the stomach of the Tollund man (Renfrew, 1973).

109. Sinskaia (1928) proposed two main centres of origin, with the Mediterranean area as primary centre for the European form, and Afghanistan with the adjoining portion of Pakistan as the other primary centre. Asia Minor, the Transcaucasus and Iran were considered secondary centres. Alam (1945) concluded that the Sarson and Toria types of *B. rapa*, now grown as oil crops in India and Pakistan, evolved in the Afghan – Persian area and migrated south to India and further east. McNaughton (1995a) concluded that multiple domestication of the wild forms for oilseed occurred from the Mediterranean to India about two millennia B.C. with later selection for short stature and leafiness in the Far East (China) resulting in the numerous *B. rapa* vegetable forms. Tsunoda and Nishi (1968) proposed that, with selection for increased leaf number, subsp. *chinensis*, and *japonica* evolved and with increased leaf size and head forming, *pekinensis*, *narinosa* and *nipposinica* were selected. Cultivation of the oilseed form in Europe as a source of lamp oil is thought to have been under way by the 13th century, first as an annual form from which the biennial form was selected (Appelqvist and Ohlson, 1972). In Northern Europe, turnip evolved from the biennial oilseed form through selection for bulbous roots (McNaughton, 1995a). Cartier in 1540 is credited with the first introduction of turnips into North America and more specifically to eastern Canada. They were also being grown in the Virginia colony by 1609 (Sauer, 1993). Canadian commercial production of the oilseed form began in 1943.

1.3.6.4 *B. oleracea*

110. *B. oleracea* has its centre of origin in the Mediterranean region (Snogerup, 1980). The wild forms of the *B. oleracea* complex still grow along the coast of the Mediterranean sea and Atlantic ocean from Greece to England (Figure 35). Snogerup *et al.* (1990) concluded from morphological and crossing studies among the wild *B. oleracea* forms, including *B. oleracea*, *B. cretica* Lam., *B. bilarionis* Post., *B. insularis* Moris., *B. villora* Biv., *B. incana* Ten., *B. macrocarpa* Guss. and *B. montana* Pourr., that these species should be considered subspecies of *B. oleracea* along with the cultivated forms. These conclusions were confirmed by Bothmer *et al.* (1995) through a crossing programme involving 10 wild taxa and 6 major cultivated forms. Snogerup *et al.* (1990) reported that all wild forms of the *B. oleracea* complex were suffrutescent perennials, exhibiting no primary dormancy. They are also self-compatible and readily intercross within the group and with cultivated forms. They also identified some wild *B. oleracea* tetraploid plants and reported a higher fertility rate in F₁ hybrids between the wild *B. oleracea* and the cultivated forms than with the other wild subspecies.

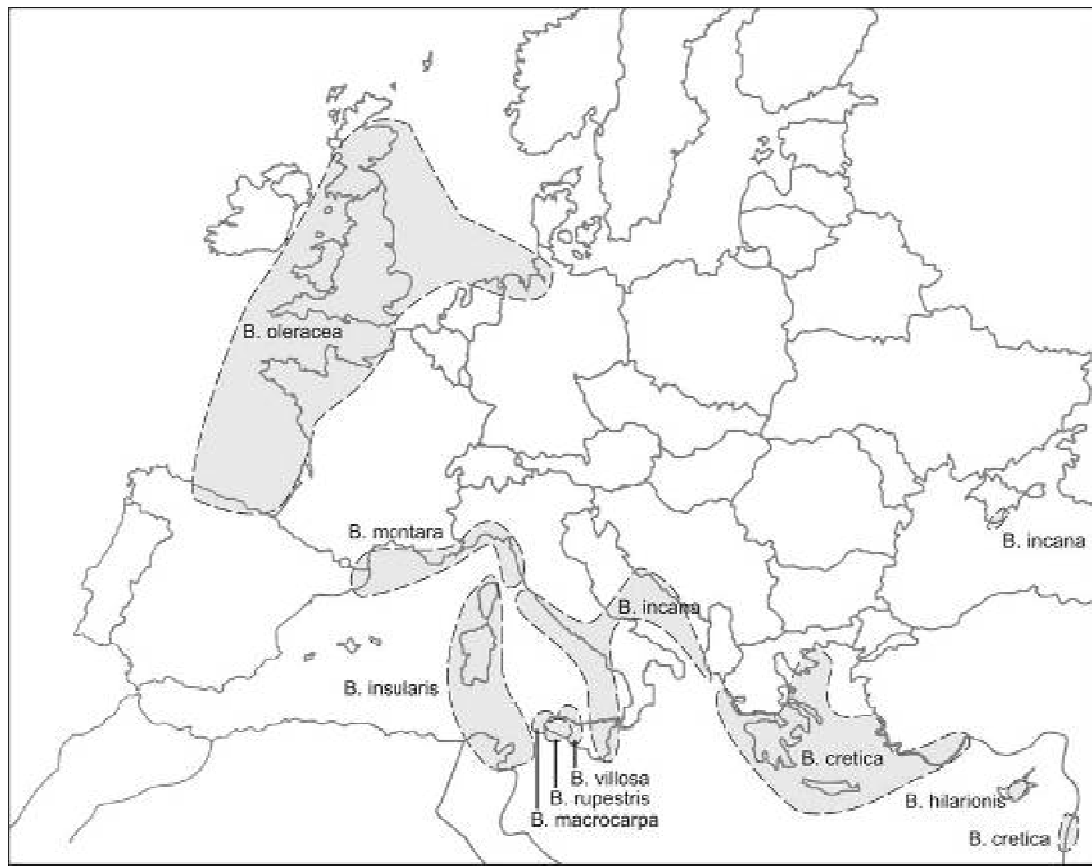


Figure 35. Distribution of wild 'species' of *B. oleracea*
 Introductions of *B. oleracea* outside its spontaneous area are not mapped
 Source: Modified from Snogerup *et al.*, 1990

111. Mutation, adaptation and selection within these populations yielded the present day forms of cabbage, savoy, kales, collard, broccoli, Brussels sprouts, cauliflower and kohlrabi. The kales, several thousand years ago, were probably the first cultivated forms. They were grown as early as 600 B.C. by the Greeks while ancient Roman writers described heading cabbage and possibly kohlrabi (Thompson, 1976). De Candolle (1886) suggested cabbage was first domesticated somewhere in Western Europe by the Celts during the first millennium B.C. Support for this conclusion comes from the respective English, German and French common names 'cabbage', 'kopf or kohl' and 'cabus', which are all probably derived from the Celtic word 'cap' or 'kap', meaning head (Prakash and Hinata, 1980). A number of authors have theorized, but lacked the research to support their views, as to which species in the *B. oleracea* complex gave rise to the various cultivated forms (Helm, 1963; Neutrofal, 1927; Schiemann, 1932; Schulz, 1936; Lizgunova, 1959). After considerable investigation, Snogerup (1980) concluded that a) headed cabbages originated from west European *B. oleracea* and savoy cabbage may have resulted from introgression with other cole crops; b) branched bush kales originated from *B. cretica* in Greece; c) stem kales probably arose from the *rupestris-incana* complex; d) the origin of the inflorescence kales such as cauliflower and broccoli is uncertain although Schulz (1936) provided some evidence that *B. cretica* could be the ancestor; and e) *B. alboglabra* originated from *B. cretica* in Greece and was carried East by traders. Today *B. oleracea* var. *alboglabra* or Chinese kale is among the ten most important market vegetables in southeast Asia, including Thailand and China (Rakow, 2004). Little is known as to when forms of *B. oleracea* arrived in Asia but Schafer (1977) noted that kohlrabi was being cultivated in Tang' times (600 to 900 A.D.).

1.3.6.5 *B. napus*

112. *B. napus* with its oilseed, forage and root forms is a relatively recent species. The Greeks and Romans knew of the Swede or rutabaga root crop, but reference to these forms does not appear in the ancient literature. Although Prakash and Hinata (1980) state that no wild *B. napus* populations have been found, Linné reported wild forms growing on the beaches of Gothland (Sweden), the Netherlands and Britain (cited by De Candole, 1886). Since the species is the result of an interspecific cross between a plant or plants of *B. rapa* and the *B. oleracea* complex, it could only have arisen in the Mediterranean or the European west coastal regions, where the two species were growing in close proximity (Figure 35). Olsson (1960) suggested that *B. napus* could have arisen several times by spontaneous hybridisation between different forms of *B. rapa* and *B. oleracea*. Evidence from chloroplast and mitochondrial DNA suggests that *B. montana* might be closely related to the maternal prototype that gave rise to *B. napus* (Song and Osborn, 1992). That *B. oleracea* was the maternal parent is supported by both Erickson *et al.* (1983) and Ohkawa (1986). However, Flannery *et al.* (2006) using SSR (simple sequence repeat) *Brassica* plastid markers noted that *B. rapa* always grouped with *B. napus* and concluded that *B. rapa* is the more likely plastid genome donor. Further, Allender and King (2010), using chloroplast and nuclear markers, concluded that it is highly unlikely that *B. oleracea* or any of the C genome species are closely related to the maternal progenitor of most *B. napus* accessions. They suggest that a *B. rapa* strain from northern Italy called 'spring broccoli raab' may be the closest extant relative of the *B. napus* maternal ancestor. However, the data also suggest that the interspecific cross may have occurred more than once with *B. napus* having multiple origins. Thus the Swede or rutabaga could have originated in medieval gardens where turnips and kale grew side by side (McNaughton, 1995b). There is general agreement that the winter or biennial form of *B. napus* originated in northern Europe. On the other hand, forage rape almost certainly evolved from the oilseed form.

113. Cultivation of oilseed rape in Europe was under way by at least the middle ages (Appelqvist and Ohlson, 1972). It is only in relatively recent times that *B. napus* oilseed forms have been introduced to other parts of the world (Figure 36). *B. napus* did not arrive in China or Japan until about 1860 to 1870, with the coming of European traders (Liu, 1985; Shiga, 1970). European immigrants introduced the forage and root crop forms into North and South America in the 17th and 18th centuries. In China, Korea and Japan *B. napus* proved to be more productive than the indigenous oilseed forms of *B. rapa*. Today most of the oilseed rape produced in China, Korea and Japan is harvested from *B. napus* cultivars that have been bred from interspecific crosses between introduced *B. napus* and the older indigenous *B. rapa* cultivars (Shiga, 1970). *B. napus* is less adapted to the Indian sub-continent due to the short days and warm growing conditions. Commercial production of the oilseed form did not occur until 1942 in Canada and 1969 in Australia.



Figure 36. Dispersal of the *B. napus* species from a proposed centre of origin
Distribution occurred throughout Europe in the 16th century, the Americas in the 17th
and 18th centuries and China and the Far East in the 19th century.

Source: modified from Liu, 1985

1.3.6.6 *B. juncea*

114. *B. juncea* appears to have a much longer history than *B. napus*, even though it is also the result of an interspecific cross (*B. rapa* × *B. nigra*). Fraction 1 protein data (Uchimiya and Wildman, 1978) and chloroplast DNA analysis established that *B. rapa* functioned as the female parent in the formation of this species (Erickson *et al.*, 1983; Palmer *et al.*, 1983, 1988; Song *et al.*, 1988a, b; Warwick and Black, 1991; Yang *et al.*, 2002). However, Qi *et al.* (2007) reported that some Chinese phenotypes may have evolved with *B. nigra* as the maternal parent. They investigated the nuclear Internal Transcribed Spacer (ITS) regions of ribosomal DNA from 15 different Chinese vegetable phenotypes and one oilseed form (Pictures of the 16 phenotypes, including two root forms, are provided in the publication). They found that four of the accessions, including the oilseed form, apparently had *B. nigra* as the maternal parent, a finding at odds with the RFLP and chloroplast DNA investigations noted above. However, the difference may be related to the limited Chinese genotypes that were available to other researchers.

115. There has been much speculation in the literature as to the centre(s) of origin for *B. juncea*. However, Prain (1898), Sinskaia (1928) and Vavilov (1949) all agree that China, where the greatest divergence of forms occurs, is one centre of origin. In addition Vavilov (1949) also identified Afghanistan and adjoining regions as a second primary centre. This observation was supported by Olsson (1960) and Mizushima and Tsunoda (1967) as well as Tsunoda and Nishi (1968) who found wild forms growing on the plateaus in Asia Minor and southern Iran. India and the Caucasus have also been put forward as secondary centres (Hemingway, 1995) (Figure 34). There is strong evidence for China as a primary site. As noted in the *B. rapa* section above, *B. juncea* has a long history in China. Leafy, vegetable forms of *B. juncea* mustard are also consumed in great quantities in China and other Asian countries (Herklots, 1972; Nishi, 1980). The greatest range in leaf types occur in Sichuan Province within varieties *rugosa*, *japonica*, *integrifolia* and *cernua*. A root forming type has also been selected and cultivated in Northern China with the variety names of *napiformis* and *tumida* (Nishi, 1980; Chen *et al.*, 2005).

116. The *B. juncea* from Afghanistan and Asia Minor is believed to have migrated south to Pakistan and India where a secondary centre of origin was established (Figure 34). The earliest direct reference to *B. juncea* is in the Indian Sanskrit literature about 1500 B.C., where it is mentioned as 'Rajika' (Prakash

and Hinata, 1980). The existence of two primary centres in China and the Middle East-India is supported by the fact that the Indian sub-continent and Chinese oilseed forms not only differ in morphological traits (Sinskaia, 1928), but also chemically and in day length requirements. The seed from Indian *B. juncea* material contains mainly 3-butenyl glucosinolate and the crop is day neutral, while the Chinese spring sown oilseed forms contain only 2-propenyl (allyl) glucosinolate and are long day requiring. The Chinese material also contains pure yellow seeded strains which are absent in the Indian material. The Russian material displays most of the same characteristics as the Chinese material and although it may also have resulted from an independent interspecific cross, more likely it was carried into Russia from China or Mongolia via the Northern Silk Road. Wu *et al.* (2009) investigated the relationships among 95 *B. juncea* accessions originating from China, India, Pakistan, France and Japan using Sequenced Related Amplified Polymorphisms (SRAPs). They found the Chinese vegetable phenotypes formed a highly diverse group with the spring and winter sown oilseed forms split into two separate groupings. The winter sown accessions exhibited more genetic diversity than the spring sown accessions but less than the vegetable group. The SRAP markers did not provide a clear cut separation between the Indian/Pakistan and Chinese winter sown mustards. Srivastava *et al.* (2001) using AFLP markers investigated the relatedness of oilseed *B. juncea* cultivars from India (7 cultivars), China (2), Tibet (1), Australia (2), Europe (6) and Canada (2). Their data separated the cultivars into an Indian/Chinese group and a second cluster of the remaining ones. Their findings and that of Wu *et al.* (2009) suggest a close relationship between the Chinese northern spring-sown oilseed cultivars with the European mustards, while the winter-sown cultivars are closely associated with the Indian form. The data from both Wu *et al.* (2009) and Qi *et al.* (2007) support the contention of Song *et al.* (1988b) that the vegetable and oilseed mustards had a polyphyletic origin and evolved separately.

1.3.6.7 *B. carinata*

117. *B. carinata*, commonly called Abyssinian or Ethiopian mustard or simply 'carinata', is an amphidiploid species derived from and containing the full genomic complement of the putative parental species, *B. nigra* (black mustard) as the female and *B. oleracea* as the male (Uchimiya and Wildman, 1978; Palmer *et al.*, 1983; Song *et al.*, 1988b; Erickson *et al.*, 1983). The plant is cultivated on a small scale on the Ethiopian plateau. *B. carinata* may have originated from a hybrid between kale, which is grown on the plateau, and wild *B. nigra* that is also present. However, this species, as with others in this group, almost certainly originated in the Mediterranean basin where the two putative parental species were growing in close proximity. It is believed that the cross occurred many eons ago when the climate on the African side of the Mediterranean was moist and lush. However, as the climate of this region became dryer and hotter, *B. carinata*, together with the plant community of the region that included castor oil plant and coffee, moved to the South and became isolated in the Ethiopian highlands. Thus, Ethiopia in effect preserved the environment of the centre of origin of *B. carinata* (Figure 34). Farmers of northeast Africa grow the plant both for its leaves, which are plucked, boiled and eaten, and for the edible oil in the seed. The local common name for the crop is *gomenzer*. The interspecific cross that created this species does not appear to have occurred elsewhere in nature or, if it did, the progeny did not survive. There is no commercial production of this species, other than in Ethiopia and neighbouring countries where the crop is grown on small holdings or in kitchen gardens. However, the species is being investigated and bred for potential commercial production in Spain, Canada, India and Australia.

1.3.6.8 *Sinapis alba*

118. *Sinapis alba* has its centre of origin in the eastern Mediterranean region (Figure 34) and wild forms are present around most of the Mediterranean littoral (Hemingway, 1995). In China *S. alba* appears to have been cultivated by the middle of the first millennium A.D. (Hemingway, 1995).

SECTION II – REPRODUCTIVE BIOLOGY

2.1 Generation Time and Duration under Natural and Managed Conditions

119. Generation and flowering times are discussed in the above sections dealing with cultivation and management.

2.2 Reproduction

2.2.1 *Floral biology* (Drawn from Downey *et al.*, 1980; Dickson and Wallace, 1986)

120. The basic floral characteristics of all the *Brassica* species included in this document are essentially the same, differing only in flower size. The floral arrangement in *Brassica* species is typically a corymbiform raceme. Flowering is indeterminate beginning at the lowest part of the main raceme and auxiliary branches, and continuing upward. The inflorescence may attain a length of 1 to 2 m. The buds begin opening under the pressure of the rapidly growing petals. The process of flower opening begins in the afternoon and is all but complete very early the following morning. The stigma is receptive from 3 days before to 3 days after the flower opens (Mohammad, 1935). Day length can play a critical role in initiating bolting of the flowering stem. Species such as *S. alba* are very day-length sensitive while some cultivars of *B. napus* and *B. juncea* are day-neutral. Both the onset of flowering and duration of the flowering period are variable and quite dependent on weather, particularly temperature. Low temperatures decrease the rate of plant development and hence the onset and rate of flowering is delayed. Low plant density results in secondary branching, thus extending the flowering period. If plants are pruned back when still green, re-growth and a second flush of flowers can be obtained. Flowers produced on re-growth are typically smaller and less productive than the first formed flowers (Downey *et al.*, 1980).

121. The flowers of the *Brassica* species are regular, bisexual and hypogenous. The differentiation of the flower proceeds through the successive development of four free sepals in two whorls, medium and transverse, six stamens, two carpels and four free diagonally placed petals (Figures 2 and 37). The flowers have one pair of lateral stamens with shorter filaments and four median stamens with longer filaments. When the anthers are a few millimetres in length, the pollen mother cells, after meiosis, give rise to the tetrads. The pollen grains are 30 to 40 μm in diameter and have three germination pores. The sutures of the anthers are introse in the bud stage, but the four long anthers become extrose as the flower opens (except in the *B. rapa* Yellow Sarson form where they remain introse).



Figure 37. Typical flower of *B. napus*

This photo is showing the typical 4 petals with the stigma in the centre surrounded by 4 median stamens and a pair of shorter lateral stamens

Source: Downey *et al.*, 1980

122. Two functioning nectaries are located at the base of the short stamens and two non-functional nectaries at the base of the pairs of the long stamens. The anthers dehisce when the petals completely unfold. The pollen is shed through two longitudinal slits on the upper side of the anthers. If the weather is warm and dry, nearly all the pollen is shed the day the flower opens. In the evening the flowers tend to close, approaching a funnel shape but open again the following morning. On the third day the flower remains almost closed and the petals and sepals begin to wilt. Studies on pollen-tube growth indicate that fertilization is effected within about 24 hours of pollination (Khanna and Chowdhury, 1974). The two carpels (although flowers on some plants may produce three or four carpels) form a superior ovary with a “false” septum and two rows of campylotropous ovules. After fertilization the ovary develops into a bivalve silique with a longitudinal septum (Figure 2). When the buds are about 5 mm long the megaspore in each ovule divides twice, producing four cells, one of which becomes the embryo sac, while the others abort. The nuclear tissue is largely displaced by the remaining embryo sac and at flower opening, the ovules mainly consist of two integuments and the ripe embryo sac.

2.2.2 *Pollination, pollen dispersal and viability*

123. *Brassica* pollen, although heavy and slightly sticky, can still become air-borne and float on the wind due to its minute size (30 to 40 μm). In addition to wind, pollen can be transferred by insects, primarily honey bees (Williams *et al.*, 1986, 1987; Scheffler *et al.*, 1993; Paul *et al.*, 1995; Timmons *et al.*, 1995; Thompson *et al.*, 1999). Physical contact between flowers of neighbouring plants also results in pollen dispersal while animals, including humans, passing through flowering *Brassica* fields can act as pollen vectors. Pollen movement can be detected using pollen traps for air-borne pollen or by using bait plants (either male sterile or emasculated) to detect outcrossing, usually through the use of marker genes such as herbicide resistance. An effective pollen trap, developed in Germany, combines a sampler that determines pollen deposition rate (Sigma-2 sampler) and a pollen mass filter apparatus that collects sufficient pollen for Polymerase Chain Reaction (PCR) analysis (VDI Richtlinien, 2007). Pollen from the Sigma-2 sampler is analysed as to species and amount under a light microscope and/or by automated imaging analysis. Strategically located bee hives can also be used to monitor pollen flow whereby pollen in honey and bee-bread samples is concentrated and analysed under a light microscope or subjected to PCR analysis (VDI Richtlinien 2006). Under natural conditions Ranito-Lehtimäki (1995) reported a gradual decrease in pollen viability over 4 to 5 days. In the laboratory, Mesquida and Renard (1982) found pollen remained viable between 24 hours to one week. However, Chiang (1974) reported that *B. oleracea* pollen stored at 4°C germinated above 20% for the first 10 days, and even after 6-7 weeks an average 4.5% of the test pollen remained viable.

124. The greatest pollen outflow from flowering *Brassica* fields is undoubtedly wind borne. Studies have shown that the vast majority of the pollen cloud travels less than 10 m and approximately half the pollen produced by an individual plant falls to the ground within 3 m (Lavigne *et al.*, 1998). In a two-year study, Bilsborrow *et al.* (1998) reported that the pollen concentrations at 10 m was reduced by 48% and 67% compared to that recorded 2 m from the field border. McCartney and Lacey (1991) found that the amount of pollen detected at 20 m from the field border was 90% less than that recorded at the field edge. Over longer distances of 360 m and 400 m, relative to the field margin, Timmons *et al.* (1995) and Thompson *et al.* (1999) reported reductions of 90% and 95%, respectively. These findings combined with outcrossing data established that *Brassica* pollen follows a leptokurtic distribution *i.e.* the presence of pollen shows a steep decline with distance, but with a long tail containing long distance events (Figure 38) (Thompson *et al.*, 1999; Staniland *et al.*, 2000). These data indicate that at a distance of 50 m from the pollen source the level of outcrossing is less than 0.5%, even when male sterile bait plants are used as pollen recipients (Figure 38).

125. Where fields are large (> 60 hectares) and/or production regions are extensive as in Canada, Australia and India, wind is considered to be the primary pollen vector since bee populations cannot service the vast number of exposed flowers. However, in the United Kingdom and other parts of Europe where field size is small and bees and pollen beetles are abundant, insects play an important role in pollen dispersal, especially over long distances (Ramsay *et al.*, 1999, 2003; Thompson *et al.*, 1999). Pollen distribution by insect can vary greatly depending on the production region, the environment and the experimental design (Gliddon, 1999; Thompson *et al.*, 1999; Ramsay *et al.*, 2003). Honey bees visiting a new field are covered with pollen from that field after visiting about four flowers, thus reducing the chances of cross fertilization between plants of the new field and fields previously visited (Cresswell, 1994). Honey bees are also more efficient pollinators than wind borne pollen over longer distances. This is to be expected since to effect fertilization, wind borne pollen must fall from the sky and land on an unfertilized stigma. Using published measurements of pollen dispersal, Hayter and Cresswell (2006) estimated that when bees are scarce, wind can contribute to pollination of fields 1 km distant at a level of up to 0.3%, but only up to 0.007% when bees are abundant. However, with a non-GM pollen source 500 m from a bee hive and a GM field 800 m from the same hive, Ramsay *et al.* (1999) detected some pollen grains from the GM field in largely non-GM pollen loads. They concluded that there was either switching between fields or a long persistence of pollen grains on the bees, or there was pollen mixing within the hive. Ramsay *et al.* (1999) also found that honey bee colonies can forage up to 2 km from their hive, indicating a potential for pollen transfer around the hive covering an area 4 km in diameter. The maximum 4 km distance for pollen dispersal by bees corresponds closely with the 4 km maximum for the wind born pollen model reported by Timmons *et al.* (1996).

126. A number of models have been developed to predict the level of gene flow that might be expected among *B. napus* fields and feral populations as well as interspecific crosses with *B. rapa* (among others, Bateman, 1947a, b; Lavigne *et al.*, 1998; Colbach *et al.*, 2005; Klein *et al.*, 2006; Devaux *et al.*, 2007; Ceddia *et al.*, 2007). However, as many biotic and abiotic factors affect gene flow, the models currently only provide an approximation. Further, the models have tended to focus on pollen dispersal and its arrival on the stigma, and have paid little attention to hybridisation and introgression.

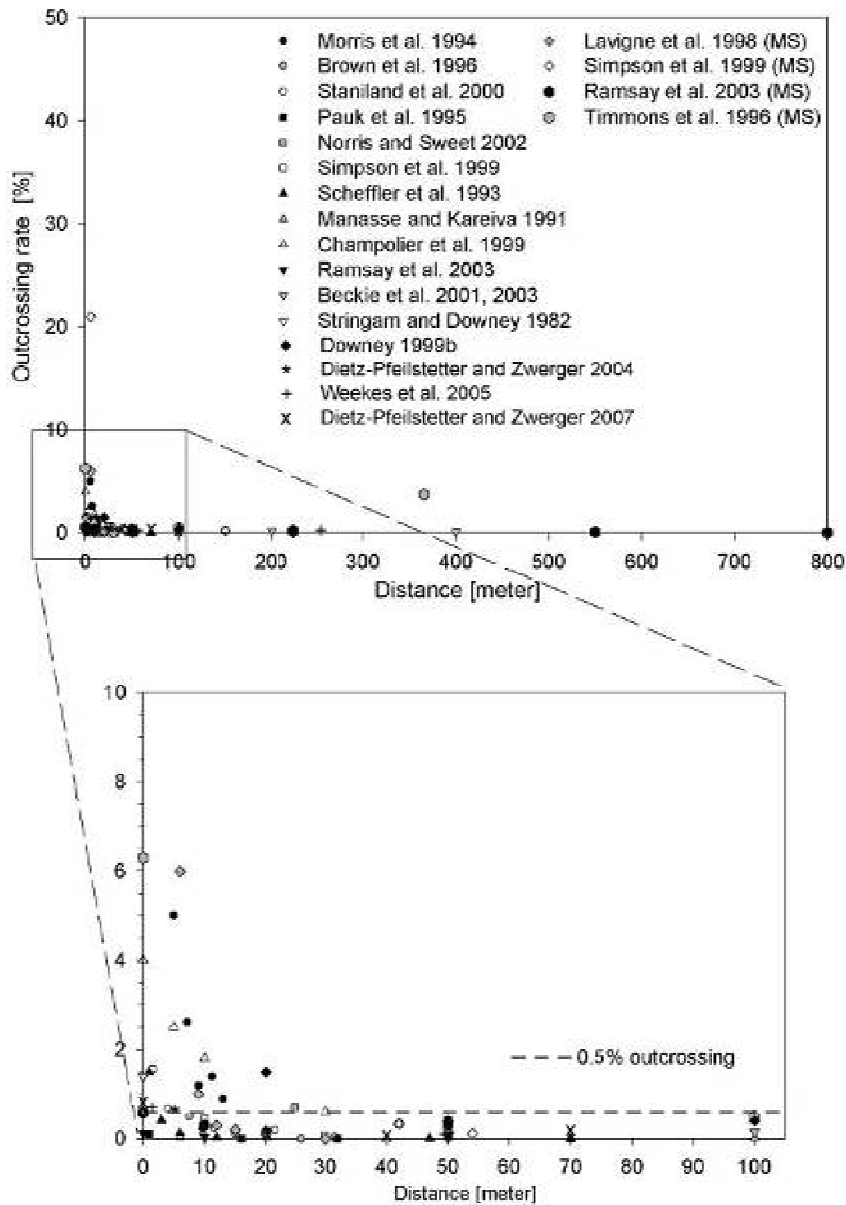


Figure 38. Outcrossing percentages as affected by distance from the pollen source
 Source: modified from Andersson and de Vicente, 2010
 (MS) indicates male sterile bait plants used.

2.2.3 Outcrossing in the field

127. Although *B. napus* is self-compatible (autogamous), pollen from neighbouring and distant *B. napus* plants compete with the plant's own pollen to effect fertilization. There are no genetic or morphological barriers to cross pollination among *B. napus* plants so crossing between fields does occur (Becker *et al.*, 1991, 1992; Rakow and Woods, 1987). The outcrossing rate within fields varies considerably, averaging between 20 to 40%, mainly depending on the environmental conditions during flowering (see Becker *et al.*, 1992 and references therein). It is estimated that one hectare of spring oilseed rape produces 9.3 ± 0.5 kg of pollen each 24 hours during a 17-day flowering period with *B. rapa* fields producing 20.2 kg/ha/day, more than twice that of *B. napus* (Szabo, 1985). Most of the crossing occurs between neighbouring plants (Rakow and Woods, 1987), but long distance pollen transfer can occur by

both wind and insects (primarily bees). The measurement of pollen flow via wind or insects, or estimating the amount of outcrossing using male sterile or emasculated bait plants, provides information on the potential for outcrossing; however, it is not an accurate indicator of the actual outcrossing level that can occur between fully fertile oilseed rape crops. In reality, male sterile plants would normally be growing in association with fully fertile plants so data from male sterile bait plants significantly over estimates the level of outcrossing that would normally be expected. Ramsay *et al.* (2003) concluded that bait plants over-estimate the outcrossing level by at least one order of magnitude.

128. Numerous experiments have been undertaken in recent years to determine the frequency of outcrossing that occurs between two populations of *B. napus*, with increasing distance between the pollen donor and recipient populations. The availability of HR genes and other markers have facilitated the detection of such genes in non-HR *B. napus* plots and fields and multiple HR types in single HR crops. However, measurement of the rate of outcrossing is complex as it can vary with the experimental design, environmental conditions, cultivars grown, synchrony of flowering, insect pollinator activity, local topography and the relative size and arrangement of the donor and recipient populations. Two types of designs have been used in these studies. In the continuous design the recipient population surrounds the donor, while in the discontinuous designs the recipient populations are distributed in locations at increasing distances from the pollen source (Hüsken and Dietz-Pfeilstetter, 2007). Using continuous designs, over short isolation distances (0 to 30 m), researchers observed a rapid decline in outcrossing rates as they sampled from the field edge into the recipient population (Scheffler *et al.*, 1993; Morris *et al.*, 1994; Brown *et al.*, 1996; Staniland *et al.*, 2000; Reboud, 2003; Dietz-Pfeilstetter and Zwerger, 2004, 2009). Examples from such studies, conducted in the United Kingdom, United States and Canada, are given in Table 6. These results underline the importance of determining outcrossing data across the whole field and not just the level at a particular spot or distance into the field. However, using commercially sized fields in a discontinuous design, Rieger *et al.* (2002) found that fields situated within 100 m of the pollen source showed very little edge effect while fields far from donor sources displayed a low and variable edge effect.

Table 6. Short distance pollen mediated gene flow from *B. napus* pollen donor to recipient field/plots in the United Kingdom, United States and Canada

Metres into recipient field	Outcrossing %	Reference, location and trial year
0.5	4.8	
1.0	1.5	Scheffler <i>et al.</i> 1993
3.0	0.4	UK, 1991
6.0	0.11	
12.0	0.016	
24.0	0.004	
36.0	0.001	
0.0	2.0/3.5	Morris <i>et al.</i> 1994
0.3	1.0/1.5	USA, 1992
0.6	0.75/1.2	east/west wind
3.0	0.65/0.6	direction
4.6	0.50/0.6	
0.0	0.70	Staniland <i>et al.</i> 2000
2.5	0.30	Canada 1994-95
5.0	0.10	Data averaged over wind directions
10.0	0.07	years
15.0	0.08	
20.0	0.07	
25.0	0.04	
30.0	0.03	

129. In most of the small plot trials, arranged in a continuous design, the area occupied by the donor population is small in relation to the recipient populations, the ratio being about 1:4. This unequal

availability of pollen tends to dilute the amount of donor pollen accessible to both wind and bee vectors. As a result, outcrossing rates reported for small plot trials with isolation distances of over 30 m tend to be lower than those recorded in larger scale investigations where the area devoted to the pollen donor are substantially greater (Table 7). Crawford *et al.* (1999) estimated that a square donor plot of at least 400 m² would be needed if a sharp decline in the effectiveness of donor pollen is to be avoided. Positioning of the donor and recipient fields can also affect the outcrossing measurements. Ingram (2000) noted that the rate of outcrossing would be higher when the long sides of donor and recipient fields faced each other. Hüsken and Dietz-Pfeilstetter (2007) statistically analysed published outcrossing results for both continuous and discontinuous designed studies. Their data indicate that with the discontinuous design the mean outcrossing rate between *B. napus* fields at 50 and 100 m would be 0.11% and at 200 m, 0.05% with lower rates for the continuous design studies (Table 8).

Table 7. *B. napus* to *B. napus* outcrossing rates, by isolation distances, reported from small plot trials and/or large fields

Isolation distance	Small Plot Trials		Large Field Trials (0.05 ha or more)	
	% Outcross	Reference	% Outcross	Reference
30-60 m	0-0.0003	Scheffler <i>et al.</i> (1993)	<0.01	Champolivier <i>et al.</i> (1999)
	0.022	Manasse & Kareiva (1991)	0.1	Simpson ²
	0.11-0.16	Sweet <i>et al.</i> (1999a)	0.2	Beckie <i>et al.</i> (2001)
	0.02-0.24	Monsanto ¹	0-0.4	Downey (1999a, b)
	0.05-0.33	Simpson <i>et al.</i> (1999) (MS)	0.1-0.65	Norris ²
	2.1	Stringam & Downey (1982)	0.02	Ramsay <i>et al.</i> (2003)
	0.02	Staniland <i>et al.</i> (2000)	<1	CETIOM (2000)
	0.05	Von Ernst <i>et al.</i> (1998)	0.05	Wilkinson <i>et al.</i> (1995)
	0	Lavigne <i>et al.</i> (1998) (MS)	0.1-0.08	Dietz-Pfeilstetter and Zwerger (2004)
	0.02-0.05	Wilkinson <i>et al.</i> (1995) (MS)	0.2-0.4	Weekes <i>et al.</i> (2005)
0.33	Ramsay <i>et al.</i> (2003)	0.00-0.09	Rieger <i>et al.</i> (2002) ³	
90-150 m	0.01-0.02	Manasse & Kareiva (1991)	0.05	Simpson ²
	0.00-0.07	Kamler (2000)	0.1	Downey (1999a, b)
	0.11-0.22	Simpson <i>et al.</i> (1999)	0.15	Beckie <i>et al.</i> (2001)
	0.01-0.13	Simpson ² (FB)	0.25-0.5	Norris ²
	0.01-0.21	Monsanto ¹	<0.5	CETIOM (2000)
	0.5	Timmons <i>et al.</i> (1996) (MS)	0.01-0.02	Weekes <i>et al.</i> (2005)
175-225 m	0.02-0.03	Simpson ² (FB)	<0.1-0.2	Norris ²
	0.017-0.6	Dietz-Pfeilstetter <i>et al.</i> (1998)	0.2	Beckie <i>et al.</i> (2001)
	0-0.9	Monsanto ¹	0.02	Ramsay <i>et al.</i> (2003)
	0.15	Scheffler <i>et al.</i> (1995)	0.00-0.005	Rieger <i>et al.</i> (2002) ³
	0.21	Ramsay <i>et al.</i> (2003) (MS)		
360-400 m	0.0038	Scheffler <i>et al.</i> (1995)	0.1	CETIOM (2000)
	0.06	Simpson ² (FB)	0.14	Beckie <i>et al.</i> (2001)
	0.6	Stringam & Downey (1982)	0.00-0.025	Rieger <i>et al.</i> (2002) ³
	0.0	Monsanto ¹		
	3.7	Timmons <i>et al.</i> (1996) (MS)		
500-800 m	0.02-0.1	Ramsay <i>et al.</i> (2003) (MS)	0.00-0.053	Ramsay <i>et al.</i> (2003)
			0.001-0.03	Rieger <i>et al.</i> (2002) ³

¹Cited by Salisbury (2002). ²(FB) indicates use of fertile bait plants, cited by Eastman and Sweet (2002), ³Ranges estimated from published graph. (MS) indicates the use of male sterile bait plants.

Table 8. Mean outcross percentages of pollen donor to *B. napus* recipient populations, for various isolation distances and two design classes

Distance from pollen source (m)	Continuous design			Discontinuous design		
	Mean	Std. ¹	n ²	Mean	Std. ¹	n ²
0-10	1.78	2.48	26	0.94	0.51	10
10-20	0.33	0.45	7	0.40	0.47	8
20-50	0.05	0.05	10	0.14	0.11	11
50-100	0.04	0.04	3	0.11	0.11	11
>200	n.d. ³	n.d. ³	n.d. ³	0.05	0.05	6

¹ Std. = standard deviation, ² n = number of data points, ³ n.d. = insufficient data

Source: Hüskén and Dietz-Pfeilstetter, 2007

130. The mean rate of outcrossing at various isolation distances is a valuable statistic, but the more important question might be "what is the maximum outcrossing that might be expected at various distances?" There is now considerable evidence that the highest rate of outcrossing that might be expected at 50 to 100 m is <0.5% and at 200 m the maximum would be <0.1% (Tables 7 and 8, Figure 38).

131. Under short isolation distances, surrounding the pollen source with a synchronous flowering recipient border may be effective in reducing pollen outflow (Staniland *et al.*, 2000; Reboud, 2003). Staniland *et al.* (2000) found that surrounding a spring *B. napus* pollen donor with a 15 and 30 m wide *B. napus* border/pollen trap, separated from the pollen donor by a cultivated 1.5 m strip, reduced the outcrossing level to 0.02% at 30 m, a level they equated to the outcrossing rates observed at 200 m by Scheffler *et al.* (1995) (Table 7). They concluded that under western Canadian conditions the current regulations, requiring a 10 m wide continuous border surrounding the pollen donor, would effectively contain the majority of pollen-mediated gene flow, but would not completely eliminate gene escape. Field size experiments by Reboud (2003), using 24 m borders, indicated that for short isolation distances gaps of bare ground between the donor and recipient plots/fields should be avoided. Outcrossing declined more rapidly when there were intervening plants, *e.g.* when the pollen donor was separated from the recipient field by a 3 to 4 m gap the level of outcrossing was similar to that found 1 m into the crop where the gap was zero. The same effect was noted by Dietz-Pfeilstetter and Zwerger (2004) when a bare gap between donor and recipient fields was increased from 0.5 to 10 m.

132. In the large field studies, not all the factors contributing to gene flow have been controlled. Weekes *et al.* (2005) found the level of outcrossing to be considerably higher in winter than in spring oilseed rape (Table 9) while Ramsay *et al.* (2003) found the opposite to be true. They attributed the low value in the winter rape trial to poor pollinating weather in May. However, Reboud (2003) and Dietz-Pfeilstetter and Zwerger (2009) observed that varieties used as pollen donors differed significantly in their outcrossing potential. The outcrossing values in some fields in the Rieger *et al.* (2002) study may have been overestimated since seed sown in the recipient fields was not tested as to the possible presence imidazolinone tolerant seeds (Salisbury, 2002). Such contaminant HR seed could have been present in seed sown in the recipient fields as a result of outcrossing or admixture during the breeding and multiplication of the donor and recipient varieties, as was observed in Canada by Downey and Beckie (2002) and Friesen *et al.* (2003). Also, it has been suggested that outcrossing levels were underestimated due to the segregation of the two genes required to provide full tolerance to the selective herbicide. Hall *et al.* (2000) identified some herbicide resistant seedlings from recipient plants situated some 650 m from an HR field. However, Downey (1999b) suggested the seed may have been transported by the farmer's swathing and harvesting equipment as observed in the Dietz-Pfeilstetter and Zwerger (2009) study. The outcrossing percentages reported by Stringam and Downey (1982) are substantially higher than recorded for other studies listed in Table 7. However, it should be noted that in the Stringam-Downey trials the pollen donors were fields of > 60 hectares which resulted in the overloading of the small 42 m² recipient plots with donor pollen. Similar high outcrossing rates were recorded by Ramsey *et al.* (2003) where blocks of 10 male-sterile plants were placed at increasing distances from a large commercial field. These results have implications for feral populations situated near commercial fields. Other observations suggest that field-to-

field crossing is likely to be highest in fields just commencing or finishing flowering when a nearby field is in full bloom.

Table 9. Predicted outcrossing rates for spring and winter oilseed rape at three isolation distances (with 95% confidence limits), based on 2000-03 multilocation UK field trials

Oilseed rape type	Percent outcrossing		
	2 m	50 m	150 m
Spring	0.46 (9.97) ¹	0.02 (0.39)	0.01 (0.14)
Winter	0.76 (12.25)	0.04 (0.84)	0.02 (0.40)

¹ 0.46 is the average % outcrossing with a 5% chance that outcrossing could be as high as 9.97%
Source: Weekes *et al.* 2005

133. Downey and Beckie (2002) and Friesen *et al.* (2003) illustrated how easily pedigree seed can be contaminated in breeding nurseries. Admixture during seeding, harvesting or cleaning was also identified as a contaminant source (Downey and Beckie, 2002). These studies alerted seed companies to the problem of contamination in Breeders seed stocks leading to tighter controls (See Section 1.3.4.8). However, the present rapid development and acceptance of *B. napus* hybrid varieties dictates that certified seed production fields will contain at least 66 to 75% male sterile plants. This increases the risk of outcrossing. In Canada, all hybrid producing seed fields are regulated and inspected to ensure that they are isolated from other rapeseed plants and fields by at least 800 m and free of certain *Brassica* weeds within the production field and the regulated isolation area. The isolation distance used by most seed companies for hybrid seed production of *B. napus* in Canada is at least 1.6 km (Wescott and Nelson, 2001). To further reduce the possibility of fertilization by foreign pollen the fields are heavily stocked with honey bees. Such fields are also saturated with leaf cutter bees [*Megachile rotundata* (Fabricius)], which have a short foraging range, to ensure the desired rapid and complete fertilization of the male sterile female parent.

2.2.4 Seed development, production, and natural dispersal

134. After fertilization the endosperm develops rapidly, while embryo growth does not start for some days. The embryo is generally still small two weeks after pollination but by 3 to 5 weeks has almost completely absorbed the endosperm and filled most of the seed coat. Nutrient reserves for germination are stored in the cotyledons which are folded one over the other so that there is a smaller inner and a larger outer cotyledon (Figure 39).

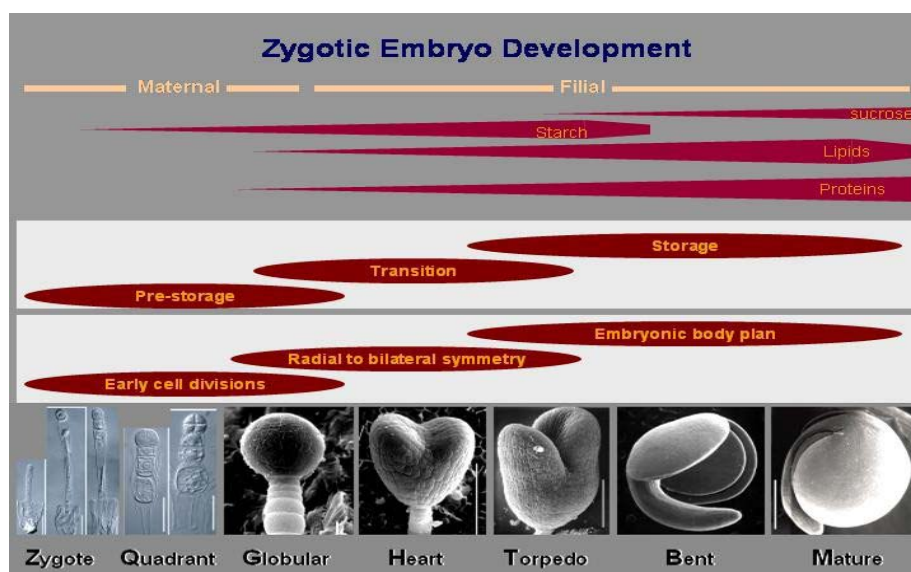


Figure 39. Development stages of the *Brassica napus* zygotic embryo
Source: courtesy Plant Biotechnology Institute, Saskatoon, Canada

135. The size of seeds can be defined by both their physical dimensions and weight. The range in seed weight among the *Brassica* crop species is given in Table 10. Typical seeds of *Brassica* species and subspecies are illustrated in Figure 40. These drawings, produced by the USDA many years ago, are still valid and can be used as a starting point to distinguish many of the species and subspecies according to the reticulation patterns on the seed surface. The different patterns are the result of variation in the size of the palisade cells that form the outer cell layer of the seed coat. Vaughan and Whitehouse (1971) investigated and described the seed surface and general features of some 200 Brassicaceous species including shape, colour, mucilage production and hilum characteristics. Koul *et al.* (2000) also examined the seed surface architecture of 78 accessions from the three subtribes, Brassicinae, Raphaninae and Moricandiinae, at both low magnification (x80) as well as the fine structure using a scanning electron microscope (x640, x1260). They noted that the seed coat patterns at high magnification were generally species-specific. However, significant seed coat pattern variations were found at the intraspecific level among the *Brassica* diploids, *B. rapa*, (two types), *B. nigra* (one type) and *B. oleracea* (two types) with the patterns of *B. rapa* and *B. oleracea* resembling each other. The seed coat patterns in most of the amphidiploids were intermediate to their putative parents, although one *B. carinata* and one *B. napus* accession exhibited patterns of their respective *B. nigra* and *B. rapa* parents. Thus employing seed coat reticulations for species identification is not foolproof, but it provides a good starting point to identify the adventitious presence of foreign species in commercial seed lots.

Table 10. Typical seed weight ranges (or averages) of *Brassica* crop plants by species and form

Species	Form	g/1000 seeds
<i>B. napus</i>	Winter oilseed rape ¹	4.5 – 5.5
<i>B. napus</i>	Spring oilseed rape ²	2.5 – 4.6
<i>B. rapa</i>	Winter turnip rape ¹	3.0 – 4.0
<i>B. rapa</i>	Spring turnip rape ¹	2.0 – 3.0
<i>B. juncea</i>	Condiment and oilseed mustard ³	2.5 – 3.0
<i>B. oleracea</i>	Cabbage ⁴	3.6
<i>B. oleracea</i>	Broccoli ⁵	2.7 – 5.8
<i>B. oleracea</i>	Brussels sprouts ⁶	2.8
<i>B. oleracea</i>	Kohlrabi ⁶	3.2

¹ Bengtsson *et al.* (1972)

² Elliott *et al.* (2008)

³ Rakow *et al.* (2009a, b)(The Indian cultivar Pusa Bold has larger than normal seed at about 5.3 g/1000)

⁴ Ohio State University (2009)

⁵ Heather and Sieczka (1991)

⁶ George (2009)

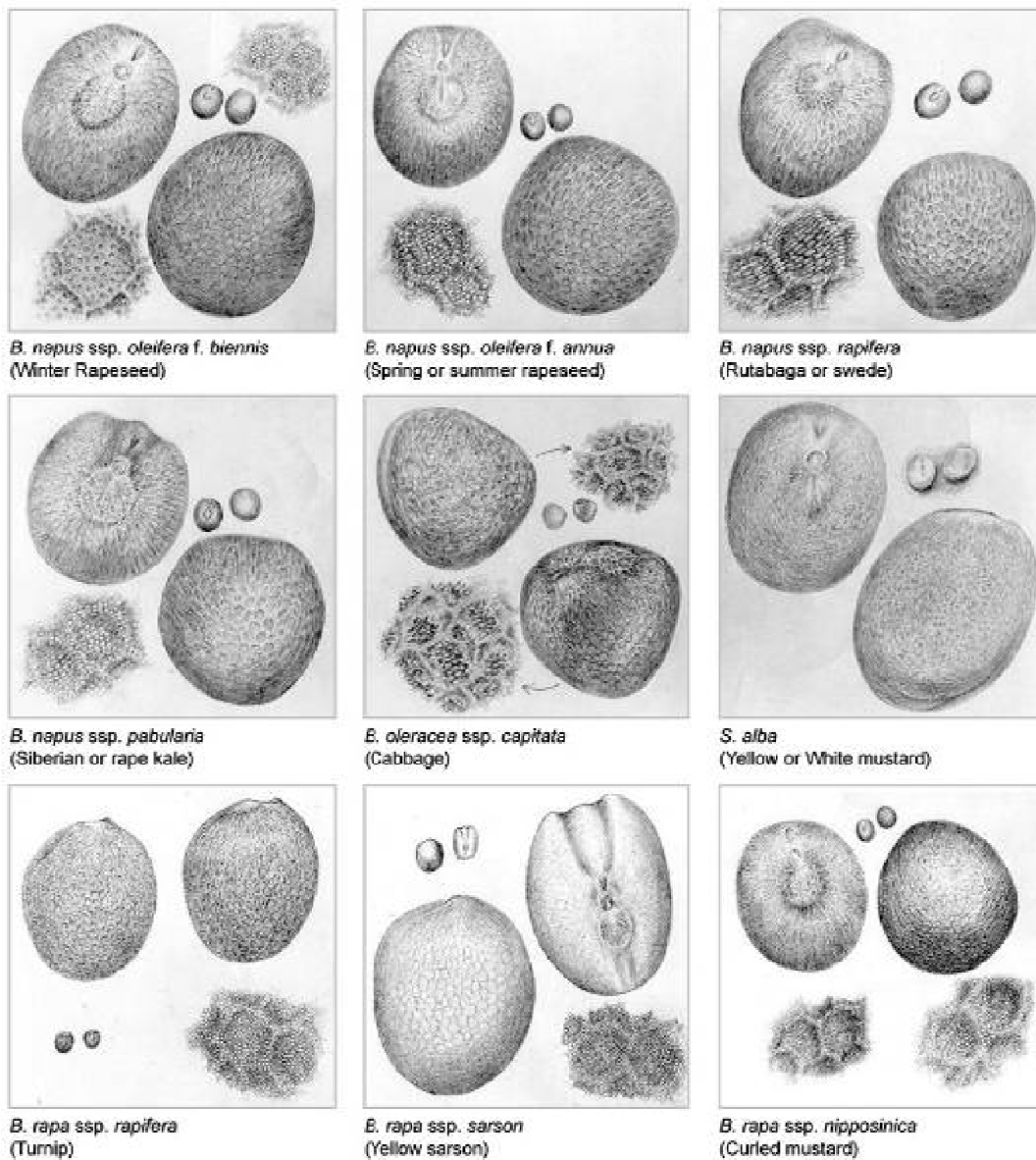


Figure 40. Distinguishing *Brassica* species by their seed coat characteristics

The small seeds shown in each compartment are about 3X natural size. The greatly enlarged surface detail is not drawn to scale but a relative proportion is maintained throughout.

Source: USDA

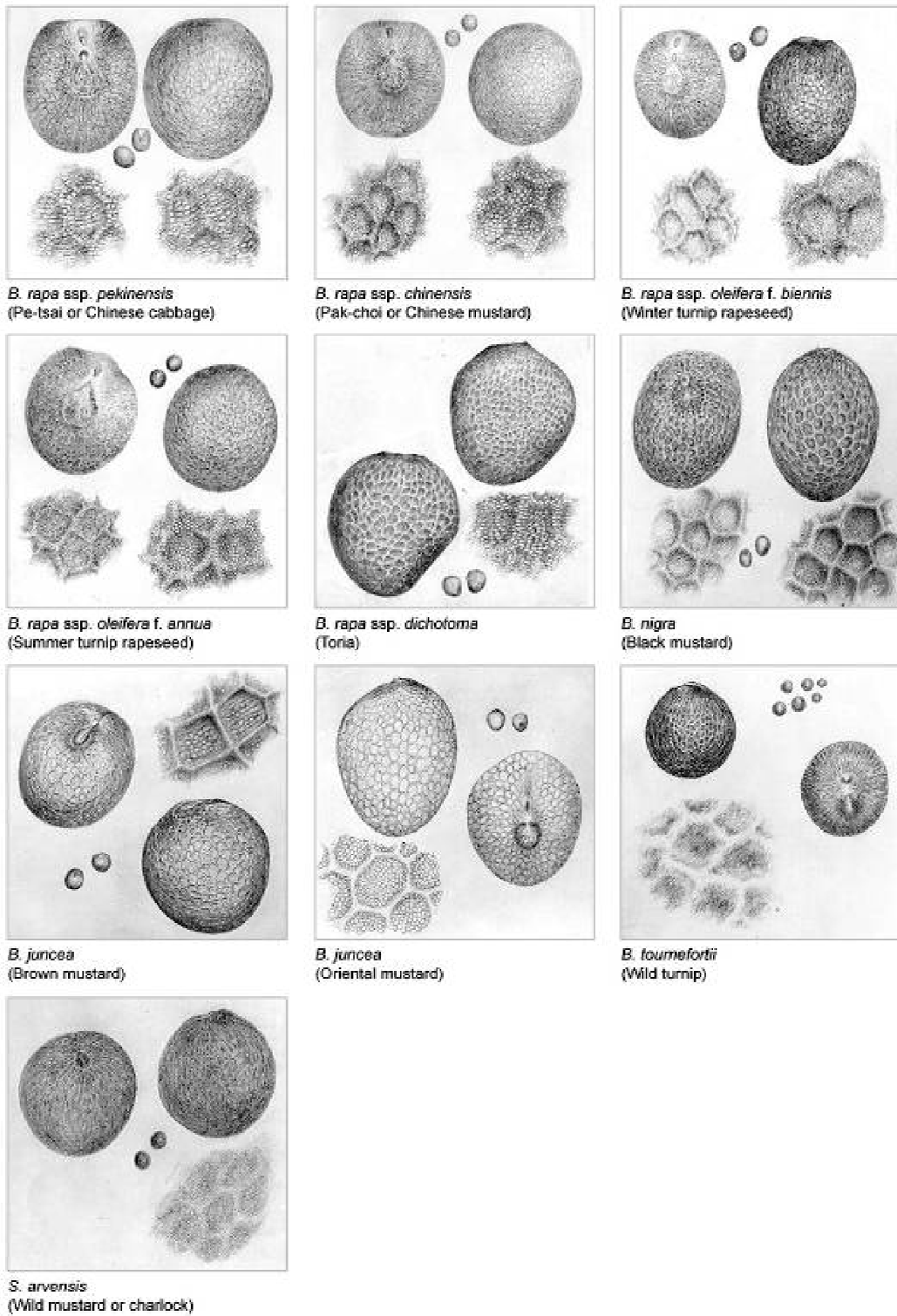


Figure 40. continued

136. The fruit of major *Brassica* crops is a glabrous silique, which is 4 to 5 mm wide and can be over 10 cm long, with two rows of seeds lying along the edges of the replum (false septum, an outgrowth of the placenta). A silique normally contains 10 to 30 seeds. Three to four weeks after the flower opens the silique attains its full diameter and length. When ripe, the silique has a tendency to dehisce and shatter, dispersing its seed. Species and varieties differ in their susceptibility to shattering. The physical forces of silique hitting silique or other plant parts causes a separation of the valve walls from the placenta, starting at the pedicel end and working toward the unattached end. The exposed seeds attached to the placenta are soon dislodged by wind action. Threshing operations easily separate the seed from the intact siliques.

137. All commercially-grown *Brassica* crops, as well as weedy species, tend to shatter their seed when ripe. However, the ease or degree of shattering varies among species. Within the oilseed crops, *B. napus* has the greatest tendency to shatter its seed, with *B. rapa* intermediate and *B. juncea* the least. Breeding work is underway to transfer the shatter resistant characteristic from *B. juncea* to *B. napus* (Wang *et al.*, 2007b). The vegetable *Brassica* species follow a similar pattern. However, with the high value F₁ seed of *B. oleracea* hybrids and the relatively small fields used for seed production, every precaution, sometimes including hand harvesting, is taken to ensure little or no seed is lost. Pod shatter is rare in the closely-related *S. alba* (Yellow or White mustard) species but some loss of intact ripe pods, due to wind or mechanical action, does occur at harvest.

138. Seed that falls to the ground can be dispersed by wind and water as well as by birds and other animals. Because the seed is small and round it is difficult to prevent some loss during transportation of farm equipment from field to field, or from field to bin and from bin to its ultimate destination. Significant losses can occur from truck containers of uncovered oilseed rape due to the wind vortex caused by the movement of the truck. The faster the truck goes the greater the loss. The distribution of seed from the truck vortex will depend on seed size and the direction and velocity of the wind prevailing at the time of loss. For spring *B. napus*, the distance such seed will travel at various wind speeds has been calculated (Table 11), although for average spring and winter *B. napus* seed, that is larger and heavier than that used to calculate the table, the wind-borne dispersal distance would be reduced.

Table 11. Estimated dispersal distances of spring *B. napus* seed released from transport vehicles at various heights above adjacent fields

Height metres	Wind speed in km/h					
	10	20	30	40	50	60
	-----Horizontal dispersal in m-----					
1.0	1.4	2.7	4.1	5.5	6.8	8.2
2.0	2.1	4.1	6.2	8.3	10.3	12.4
3.0	2.6	5.1	7.7	10.2	12.8	15.3
4.0	3.1	6.2	9.3	12.3	15.4	18.5
5.0	3.6	7.2	10.8	14.4	18.0	21.6
6.0	4.1	8.2	12.4	16.5	20.6	24.7
7.0	4.6	9.3	13.9	18.5	23.2	27.8
8.0	5.1	10.3	15.4	20.6	25.7	30.9

¹ Estimates based on small seeds of spring *B. napus*, calculated to weigh 2.2 mg with a diameter of 1.8 mm, that are the most likely to become air borne and travel the farthest.

Source: Hertz, 1999

2.2.5 Seed viability, longevity and dormancy, germination, seedling establishment

139. Well developed, fully mature *Brassica* oilseeds may remain viable for at least 25 years if dry seed is refrigerated in sealed containers (Ellis *et al.*, 1994). As of 2009, seed of oilseed *Brassica*, harvested in 1977 and stored in manila envelopes at -20°C in the Saskatoon AAFC Seed Bank, has retained its high germination (Downey, personal communication). Viability of seed lost during harvest is an important factor in determining the presence and amount of volunteer plants and populations in subsequent crops. Harvest losses can be substantial and the survival and persistence of this seed is greatly influenced by environment, seed dormancy as well as crop and field management.

2.2.5.1 Contribution of *B. napus* harvest losses to persistence

140. Harvest losses in the United Kingdom, when the winter *B. napus* crop is straight combined under ideal conditions, ranged from 2 to 5%, but under unfavourable harvest environments could amount to 50% (Price *et al.*, 1996). Pekrun *et al.* (1998a) placed these losses between 200-300 kg/ha or about 5,000 to 7,000 seeds/m². Lutman *et al.* (2005) in the United Kingdom and Gruber *et al.* (2004) in Germany recorded average harvest losses of 3,000 to 3,500 seeds/m². Similarly, French studies estimated harvest losses to be between 1.5 and 8.5% of the average yield. This calculates to 50 to 300 kg/ha of seed remaining on the field after harvest or 1,100 to 6,700 seeds/m² (CETIOM, 2000; Messéan *et al.*, 2007). In Canada, Gulden *et al.* (2003a) reported that spring *B. napus*, harvest losses averaged 5.5% or about 3,590 seeds/m², while Légère *et al.* (2001) estimated the losses at 2,000/m². Similarly, Warwick *et al.* (2003) reported spring *B. napus* harvest losses averaging 5.5% or about 3,590 seeds/m². Salisbury (2002) estimated Australian losses would be similar to those found in Canada. However, a vast majority of the seed remaining in the field after harvest will not survive the first year. The *Brassica* oilseed density of the seed bank in western Canada is reported to drop 10 fold in the first year and to decline slowly thereafter, due to replenishment of the seed bank by uncontrolled volunteer plants. However, where post harvest tillage is shallow and delayed and volunteers in subsequent crops are controlled, very few plants are found four years after a spring *B. napus* crop (Gulden *et al.*, 2003b).

2.2.5.2 Seed dormancy

141. Seed dormancy can play an important part in determining the amount and persistence of volunteer *Brassica* plants in subsequent crops. There are two main types of seed dormancy, primary and secondary. Primary dormancy is when seed germination is prevented during the seed maturation process and for some time after the seed has been removed from its parent (Karszen, 1980/81; Hilhorst and Toorop, 1997). To overcome primary dormancy, a period of after-ripening is usually required. Secondary dormancy is a reduction in seed germinability that develops after the seed is separated from the parent plant and may, in some cases, be induced prior to the complete alleviation of primary dormancy. Primary dormancy does not occur in ripe seeds of any of the cultivated *Brassica* oilseed, vegetable or condiment crops. For seed certification status, these crops require a minimum germination of at least 90%. However, during seed maturation, germination percentages may be low in spring and winter *B. napus* but increase with maturity (Finkelstein *et al.*, 1985) to where at harvest no primary dormancy occurs (Schlink, 1995). However, secondary dormancy can be induced in *B. napus* and cultivated *B. rapa* under certain conditions (Hails *et al.*, 1997; Pekrun *et al.*, 1998b; Adler *et al.*, 1993). An exception to the rule occurs in the weedy forms of *B. rapa*, where primary dormancy is present as a recessive trait in weedy *B. rapa*. Thus crossing between weedy and cultivated *B. rapa*, as well as between weedy *B. rapa* and *B. napus*, will produce seed that does not exhibit primary dormancy (Linder, 1998; Landbo and Jorgensen, 1997; Adler *et al.*, 1993).

142. The main factors contributing to secondary dormancy of *B. napus* seed are elevated temperatures, darkness, osmotic stress and limited oxygen (Gulden *et al.*, 2004b; Pekrun *et al.*, 1997a). Studies in Europe

(Pekrun *et al.*, 1997b; Gruber *et al.*, 2004) and China (Momoh *et al.*, 2002) suggested that genotypes differ in their predisposition to undergo secondary dormancy. Indeed, it has been clearly shown that genotype is the principal factor controlling its potential in *B. napus* (Gulden *et al.*, 2004a; Pekrun *et al.*, 1997; Gruber *et al.*, 2004, 2009). Gulden *et al.* (2004a) found seed size was of secondary importance, with large seed more likely to undergo secondary dormancy, while maturity and pre- and post-harvest environment had little influence. The occurrence of secondary dormancy is reduced by alternating temperatures (Pekrun *et al.*, 1997b; Momoh *et al.*, 2002), while cold stratification readily releases secondary dormancy as does exposure to continuous light (Schlink, 1995). Exogenous applications of gibberellic acid (0.2 mg l^{-1}) will also reverse secondary dormancy (Pekrun *et al.*, 1998b).

143. In Germany, Gruber *et al.* (2004) evaluated the persistence and secondary dormancy in the seed of four winter oilseed rape varieties. They found that of the 3,000 to 3,500 seeds/m² lost during harvest, 60 to 75% of that seed either died or was scavenged within a few months. Similar levels of seed disappearance were observed by Gruber *et al.* (2003) when investigating the effect of different tillage treatments on seed persistence. Six months after harvest no seed of the variety Artus could be detected in the soil seed bank while the other three varieties. Bristol, Liberator and Capital, respectively contributed 4.3, 9.3 and 11% of their lost seed to the seed bank. Laboratory tests for the presence of secondary dormancy closely corresponded to that observed in the field. Gruber *et al.* (2009) also laboratory tested seed from over 40 varieties for their tendency to undergo secondary dormancy. The seed was harvested from one site for three years and second site for two years. They found that, over several years, varieties consistently ranked high, medium or low in percentage of seed exhibiting secondary dormancy. However, the rate of secondary dormancy varied significantly with harvest years, dry years having the lowest incidence. They concluded variety rank, rather than the actual percentage of secondary dormancy, should be used to characterise a variety. Thus selection for varieties without secondary dormancy could be easily achieved and would greatly reduce the incidence of *B. napus* volunteers in subsequent crops. It should probably be made mandatory for all new *B. napus* varieties to be free of the secondary dormancy trait.

144. At shallow burial depths, *B. napus* and closely related species exhibit low seed bank persistence (Schlink 1995; Pekrun and Lutman 1998; Sparrow *et al.* 1990; Gulden *et al.* 2003a). At 10 cm depth Gulden *et al.* (2004b) found seed bank populations shifted from a germinable to an ungerminable state and no seedling recruitment was observed. Masden (1962) reported that 1% of buried *B. napus* seed germinated after 5 years, and that trace amounts of *B. rapa* seed emerged after 10 years. Schlink (1998) and Lutman *et al.* (2003) found that approximately 1% of *B. napus* seed in undisturbed soil could survive for 10 years. Jørgensen *et al.* (2007), sampling a deep soil layer, identified viable seeds of a variety sown in the field 17 years earlier. In Canada, Beckie and Warwick (2010) reported a small population of volunteers resistant to the herbicide bromoxynil in a field that had not grown oilseed rape since the sowing of a bromoxynil-resistant variety seven years previously. The volunteers persisted in low lying areas of the field which were too wet to plant or spray with herbicides between 2001 and 2007. No volunteers were detected in either 2008 or 2009. There is general agreement that secondary dormancy will be induced in a significant percentage of deeply buried *B. napus* seed.

2.2.5.3 Persistence

145. Very few seeds of oilseed rape survive in the seed bank compared with their wild relatives (Chadoeuf *et al.*, 1998). Most seeds of the cultivated *Brassica* crops, if left on or near the soil surface, will germinate and be killed by frost or cultivation or be eaten by rodents, birds and insects. Nevertheless, a small proportion may not germinate and secondary dormancy may be induced, particularly if the seed is buried. Studies in Europe with winter *B. napus* found that when seeds were buried immediately after seed shed, 30% of the seed bank survived one winter compared to only 0.1% when seeds were left on the undisturbed soil surface (Pekrun and Lutman, 1998). Similarly for spring *B. napus*

in western Canada, Gulden *et al.* (2003a) found spring seedlings, from fall sown seeds buried at a 1 cm depth, to be only 0.1 to 1.5% of the original seed bank. In Canada, oilseed rape is typically grown on the same land once in four years with most of the volunteers occurring in the year following oilseed rape production. However, volunteers can occur 4 to 5 years after production (Légère *et al.*, 2001; Simard *et al.*, 2002; Beckie and Owen, 2007). Harker *et al.* (2006) found that if first year volunteers were prevented from producing seed, the densities of volunteers in subsequent years were reduced to levels that would not require herbicidal intervention. Surveys in southern Australia by Baker and Preston (2008), where zero and minimum till are practiced, found zero germination of seed sampled from fields 3.5 years after the last *B. napus* harvest. But in Germany, Förster and Diepenbrock (2002) reported more than 0.5 plants/m² of winter *B. napus* three years after the last oilseed rape harvest. However, no information on timing or type of post-harvest cultivation was provided. In France two conventional oilseed rape varieties, one of which was dwarf, were planted on fields that had grown three different HR varieties three to eight years before (Messéan *et al.*, 2007). The percentage of GM HR seed occurring in the harvest of the conventional varieties was determined. HR seed from two of the GM varieties never exceeded 0.9% of the conventional harvested seed. However, one GM variety that was grown five years previous made up 4 to 18% of the conventional harvest, with the highest values occurring in the seed harvested from the dwarf variety. Since all oilseed rape volunteers were removed from the rotation crops in the intervening years, the volunteers must have arisen from dormant seed in the seed bank. The results illustrate the importance of breeding varieties without the secondary dormancy trait, not only for GM varieties, but more generally for the production of pure seed stocks and segregation of specialty oil types.

146. In the United Kingdom, Lutman *et al.* (2005) recorded a large average harvest seed loss (3,575 seeds/m²) from four *B. napus* winter varieties grown in multiple-site, multi-year trials. Within six months the number of seeds present declined by an average of 63% with a slower decline recorded at 18 and 30 months. Appreciably more seeds were found on sites that were ploughed immediately after harvest compared to sites where cultivation was delayed by about four weeks. These data support the recommendations of Pekrun *et al.* (1998a) and Gulden *et al.* (2003a) that cultivation of *B. napus* stubble should be avoided for several weeks after harvest. Regression models applied to the Lutman *et al.* (2005) data predicted that it would take an average of 9 years to reduce the seed in the soil bank by 95%. However, other studies (Lutman *et al.*, 2003) indicate that the 95% reduction would occur in 3 to 4 years. Indeed, Beismann and Roller (2003) in Germany reported that no viable *B. napus* seeds could be found in soil sample cores taken from sites where transgenic plots were sown 5 and 6 years before.

147. Studies in the United Kingdom and Canada with winter and spring forms of *B. napus*, indicate that seed bank persistence is less in lighter than heavier, clay containing, soils (López-Granados and Lutman, 1998; Gulden *et al.*, 2004b). In general, if post harvest tillage is delayed and volunteers are controlled in the intervening years, the evidence indicates the presence of volunteers from the seed bank decline by at least 90% by the fourth year (Lutman *et al.*, 2003; Gulden *et al.*, 2003b; Baker and Preston, 2008). Failure to follow the above-recommended practices can extend the presence of seed bank volunteers by several years (Lutman *et al.*, 2005).

148. Linder and Schmitt (1995) assessed the persistence, in field and greenhouse trials, of GM *B. napus* lines with elevated levels of stearate and laurate fatty acids in their seed oils. They concluded the risk of persistence of the high stearate and high laurate genotypes, compared with their parental non-GM types, was low. No interspecific hybrid seed could be obtained from hand-crossing GM high stearate *B. napus* × wild *B. rapa*. Greenhouse trials using seed from the high laurate *B. napus* × *B. rapa* cross indicated that such hybrids “will not possess seed bank dynamics promoting reproduction”.

SECTION III – GENETICS

3.1 Relevant Detailed Genetic Information

3.1.1 Cytology

149. Mitotic metaphase chromosomes of the Brassicaceae are very small. Conventional cytological protocols condense *Brassica* meiotic chromosomes to tiny rods or dot-like shapes. Their small size, lack of distinctive cytological features and the difficulties of pachytene investigations make cytological identification of individual chromosomes almost impossible. Although the small chromosome size of the Brassicaceae family has limited the direct cytology approach, the sequencing of the *Arabidopsis thaliana* (The *Arabidopsis* Genome Initiative 2000), *B. rapa* (Wang, 2010; The *B. rapa* Genome Sequencing Project Consortium, 2011), *B. oleracea*, and *B. napus* genomes (Bayer CropScience, 2009) are providing a much clearer picture of species interrelationships. However, the *B. napus* sequence has not been released to the public thus far. A publicly available sequence of *B. oleracea* is expected to be available in late 2012 (I. Parkin, personal communication).

150. Comparative mapping, using more than 20 linkage maps for *B. oleracea*, *B. rapa*, *B. nigra*, *B. napus* and *B. juncea*, has contributed greatly to the understanding of chromosome homology and colinearity (Lysak and Lexer, 2006). In addition, great strides have been made in determining the extent of genome colinearity, and rates and modes of evolution in the Brassicaceae family. Comparative cytogenetic studies now employ a wide array of techniques including among others, rDNA probes, nucleolus organizer regions (NORs), variation in centromeric satellite repeats, genome *in situ* hybridisation (GISH), fluorescence *in situ* hybridisation (FISH), combined with bacterial artificial chromosomes (BAC FISH) and large scale comparative chromosome painting (CCP). Such techniques have helped to unravel the genomic evolution of *A. thaliana*, *B. oleracea*, *B. rapa*, *B. juncea* and *B. napus* as well as the time frame in which the species arose.

151. Research into the genome microstructure of the Brassicaceae species indicates the family originated from an ancestral karyotype that evolved after the monocot/dicot split. The ancestral karyotype had a basic chromosome number of $x=4$ and underwent a genome duplication some 65 million years ago (Mya) followed by diploidisation (Song *et al.*, 1988a; Rana *et al.*, 2004) (Figure 41). From this progenitor, the ancestral Brassicaceae form evolved with $x=8$ chromosomes (Lysak *et al.* 2006). This was followed by the divergence about 20 Mya of the ancestral genera of *Arabidopsis* and tribe Brassiceae. Genome triplication via allohexaploidy occurred about 14 to 16 Mya (Lysak and Lexer, 2006), followed by diploidisation and chromosome number reduction resulting in the evolution of the ancestral Brassiceae karyotype with $x=6$ chromosomes (Lysak *et al.*, 2005; Yang *et al.*, 1999). It is estimated that the separation of the Nigra and Rapa/Oleracea lineages took place about 7.9 Mya (Lysak *et al.*, 2005). The *B. oleracea* and *B. rapa* divergence is estimated to have occurred about 4 Mya (Inaba and Nishio, 2002), with the interspecific crosses, forming *B. napus*, *B. juncea* and *B. carinata*, taking place less than 10,000 years ago (Song *et al.*, 1988a; Rana *et al.*, 2004; Lysak *et al.*, 2005).

152. A slightly different scenario of the polyploidy events in the evolution of the Brassiceae genomes has been put forward by Mun *et al.* (2009), following a *B. rapa* and *A. thaliana* genome-wide comparative analysis. They suggest that a whole genome duplication (WGD) occurred twice, once about 55-63 Mya and again at 23-30 Mya, between the existence of an ancient ancestral species and the evolution of the ancestral

Brassicaceous karyotype. They suggest that the second WGD resulted in the divergence of *Arabidopsis* from the *Brassicaceae* lineage about 13 to 17 Mya. This was followed by a whole genome triplication in the Brassicaceae about 11 to 12 Mya with the divergence of *B. rapa* from *B. oleracea* taking place about 8 Mya. Their data also suggest that the allopolyploidisation that resulted in the species *B. napus* occurred only 0.7 to 1 Mya.

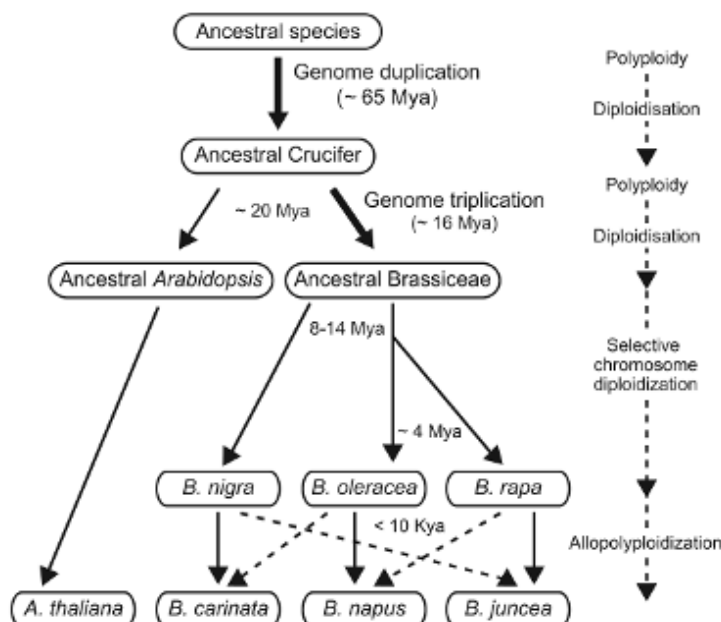


Figure 41. Illustration of major events in the evolution of selected *Brassica* species and *Arabidopsis thaliana*

Source: Modified from Song *et al.*, 1988a, b; Rana *et al.*, 2004

The dotted lines indicate the species believed to be the maternal parent in the interspecific cross.

153. Genome mapping of *B. rapa* and *B. oleracea* has shown the gross organisation of their genomes to be highly collinear (Lagercrantz and Lydiate, 1996) but their genome size and complexity differ. The genome size of *B. rapa* is *ca.* 500 Mb compared to the much larger and more complex genome of *B. oleracea* at *ca.* 600 Mb (Arumuganathan and Earle, 1991). Comparative studies have shown that within the amphidiploids species, *B. napus*, *B. juncea* and *B. carinata*, the chromosomes within the respective putative diploid genomes have remained more or less intact (Parkin *et al.*, 1995; Sharpe *et al.*, 1995; Axelsson *et al.*, 2000). DNA sequence data indicate that the A genome of *B. rapa* and the C genome of *B. oleracea* are very closely related while *B. nigra*, with its B genome, is from an earlier divergent lineage (Mizushima, 1972; Song *et al.*, 1988b; Prakash and Chopra, 1991). Song *et al.* (1995) reported there was rapid genome change after polyploidisation in *B. napus* and *B. juncea*, which suggests that the micro structural changes observed in the *Brassica* lineage happened shortly after genome duplication followed by a slow but on-going rate of change (Rana *et al.*, 2004).

154. The techniques of fluorescence *in situ* hybridisation (FISH) facilitates the integration of genetic and physical chromosome maps as it allows chromosomal location of labelled DNA probes to be directly determined (Snowdon *et al.*, 2007a). Since molecular markers can now be ordered and physical distances measured, it is possible to construct molecular karyotypes and distinguish individual chromosomes of the A and B genomes that make up *B. napus* (Fukui *et al.*, 1998; Armstrong *et al.*, 1998; Snowdon *et al.*, 2002). Snowdon *et al.* (2007b) provides a consensus genetic linkage map of molecular markers for *B. napus* where linkage groups (LGs) N1 to N10 correspond to the *B. rapa* A genome LGs of A1 to A10, and LGs N11 to N19 correspond to *B. oleracea* C genome LGs of C1 to C9.

3.1.2 Nuclear genome size

155. The genome size of the *Brassica* diploids (approximately 500-700 Mbp) are more than four times that of the related Brassicaceous species *A. thaliana* (approximately 157 Mbp) (Table 12). The gene content of *A. thaliana* is believed to be very similar to *Brassica* diploids with more than 87% sequence identity in the coding regions (Parkin *et al.*, 2005). Although it is believed that the diploid *Brassica* evolved through a common hexaploid ancestor (Parkin *et al.*, 2005), the necessary genome triplication would be insufficient to explain the differences in genome size. Therefore, this important difference in genome size is likely to reflect a different rate of non-coding DNA accumulation.

Table 12. Ploidy level, chromosome number, genome size and map length of *A. thaliana* and *Brassica* species of "Triangle of U"¹

Species	Ploidy level	Chromosome number	1C nuclear DNA ² content (Mb) ³	Observed map length (cM) ⁴
<i>A. thaliana</i>	2	10	157	437 and 501
<i>B. nigra</i>	2	16	634-765	855
<i>B. oleracea</i>	2	18	696-765	820-1738
<i>B. rapa</i>	2	20	528-784	1455
<i>B. carinata</i>	4	34	1280-1548	–
<i>B. juncea</i>	4	36	1070-1500	2073
<i>B. napus</i>	4	38	1127	1441-1765

¹ The Triangle of U is a theory about the evolution and relationships between members of the plant genus *Brassica* (Source: Wikipedia, the Free Encyclopedia).

² The C value refers to the haploid DNA content of the species

³ Data adopted by Lysak and Lexer (2006) compiled from Bennett and Leitch (2004) and Johnston *et al.* (2005). 1 pg = 980 Mb

⁴ From Lysak and Lexer (2006), choice based on map marker coverage

3.1.3 Possible extent of repetitive or non-coding DNA sequences

156. Transposable elements (TEs) constitute a major fraction of non-coding DNA in plant species. Good estimates of TE distribution and density are presently only available for the *B. oleracea* genome, based on a partial draft genome sequence (Zhang and Wessler, 2004). Class 1 (retro) elements were the most abundant TE class with long terminal repeat (LTR) and non-LTR elements comprising the largest fraction of the genome. However, several families of class 2 (DNA) elements have amplified to very high copy number in *B. oleracea* compared to *A. thaliana* and have contributed significantly to genome expansion. Approximately 20% of the *B. oleracea* genome was estimated to be composed of class 1 and class 2 TEs.

157. Lim *et al.* (2005) describe the morphology and molecular organisation of heterochromatin domains in the interphase nuclei and mitotic and meiotic chromosomes of the 10 chromosomes of *B. rapa*, using DAPI staining and FISH of rDNA and pericentromere tandem repeats. They characterised the centromeric repeat sequences, which fell into two classes, CentBr1 and CentBr2, occupying the centromeres of eight and two chromosomes, respectively. The centromere satellites encompassed about 30% of the total chromosomes, particularly in the core centromere blocks of all the chromosomes. Interestingly, centromere length was inversely correlated with chromosome length.

3.1.4 Main genetic diversity or variability

158. Considerable genetic diversity has been found within the six cultivated *Brassica* species using nuclear RFLP (Restriction Fragment Length Polymorphisms) markers (Song *et al.*, 1988b). These results

suggested that 1) *B. rapa* and *B. oleracea* have multiple centres of origin, 2) *B. nigra* originated from one evolutionary pathway whereas *B. rapa* and *B. oleracea* came from another pathway, and 3) amphidiploid *B. napus* and *B. juncea* arose from different combinations of diploid morphotypes indicating polyphyletic origins may be a common mechanism for natural occurrence of amphidiploids in *Brassica*.

159. The genetic diversity within *B. napus* is considerably less than that found within either of the diploid ancestral species. This is probably a result of *B. napus* being a relatively modern species, fixed as a product of human civilisation and with no truly wild populations. Most of the diversity within *B. napus* has been introduced from its diploid progenitors. Variation in the A genome has been increased by natural *B. napus* × *B. rapa* crosses whereas variation in the C genome is more limited. Recent molecular marker analysis has identified more extreme genetic variation in exotic vegetable and fodder genotypes as well as newly re-synthesized *B. napus* lines (Snowdon and Friedt, 2004 for review). In *B. juncea* the A genome is mostly conserved and the C genome is significantly changed, more so than the considerably altered C genome in *B. carinata*. Similar genetic information, with much duplication is contained in all three genomes (Slocum *et al.*, 1990, 1989; Chyi *et al.*, 1992; Jackson *et al.*, 2000; Parkin *et al.*, 2003). However, the chromosomal organisation and the genetic distribution within the genome is different (Truco *et al.*, 1996). New high throughput and very informative simple sequence repeat (SSR) and single nucleotide polymorphism (SNP) molecular markers are now being used routinely to expedite the introduction of novel genetic variation in *Brassica* breeding programmes.

3.1.5 Maternal and/or paternal inheritance of organelle genomes

160. Analysis of the chloroplast DNA of the cultivated diploid *Brassica* species, and their close relatives, divided the subtribe Brassicinae into two ancient evolutionary lineages (Warwick and Black, 1997), the ‘Nigra’ lineage which contained the diploid *B. nigra* and the related wild mustard *Sinapis arvensis*, and the ‘Rapa/Oleracea’ lineage which contained the diploid progenitors of *B. napus* (Figure 33). There has been little work studying the origins of the cultivated amphidiploids *B. carinata* or *B. juncea*. However, studies of organellar and nuclear DNA of *B. napus* and related species suggested that a species closely related to *B. montana* gave rise to the cytoplasm of both *B. rapa* and *B. oleracea* (Song and Osborne, 1992). The same study and an earlier study on chloroplast evolution in amphidiploid *Brassica* species (Palmer *et al.*, 1983) suggested that oilseed rape (*B. napus*) evolved from multiple hybridisations between *B. oleracea*, and the closely related n=9 species, *B. montana* and *B. rapa*. Some of these lineages may have been subject to introgression from post-hybridisation with their diploid progenitor.

3.1.6 Self incompatibility, ‘S’ alleles

161. Self-incompatibility (SI) occurs in many flowering plants and is one of the most important systems to prevent inbreeding (Takayama and Isogai, 2005). SI is defined as the inability of plants to produce functional gametes to effect fertilization upon self-pollination or when crossed with certain relatives (De Nettancourt, 1971). Although the amphidiploid *Brassica* species, *B. napus*, *B. juncea* and *B. carinata* are largely self-pollinating (autogamous), the diploid species, with some exceptions, are self-incompatible and are obligatory out crossers. Among *Brassica* species and their close relatives, 50 out of 57 species are self-incompatible (Hinata *et al.*, 1994). The self-/non-self recognition in most species is controlled by a single locus, termed the ‘S locus’ that inhibits the self pollen from penetrating the style when the same S-allele specificity is expressed by both the pollen and pistil. In the *Brassica* incompatibility system over 30 *B. rapa* alleles and 50 *B. oleracea* alleles have been identified, S₁, S₂, S₃...S₅₀₊ (Nou *et al.*, 1993; Ockendon, 2000). Self-compatible (S_i) alleles are also known.

162. Among angiosperms there are two major types of physiological SI systems, gametophytic (GSI) and sporophytic (SSI) (Briggs and Knowles, 1967). In a GSI system the pollen reaction is controlled by the genotype of the individual pollen grain, *i.e.* a plant heterozygous at the S-locus would produce

two possible types of pollen with each microspore receiving one of the two possible S-alleles. However, in the SSI system that is present in the Brassicaceae, all pollen released by a plant has the same phenotype with respect to the compatibility reaction, regardless of the genotype of the individual pollen grain. The S-locus consists of at least three tightly linked transcriptional units arranged in pairs, with one functioning as the female determinant and the other the male. This multi gene complex at the S-locus is inherited as one segregating unit so the gene complexes are called "S-haplotypes". Self-/nonself recognition operates at the level of protein-protein interaction of the two determinants (Takayama and Isogai, 2005). When the SI system is activated in a Brassicaceae species, a recognition reaction occurs between the papilla cells of the stigma and the pollen (Hinata and Nishio, 1980).

163. There are three highly polymorphic genes involved in the SI response. The two female determinants consist of the S-Locus Glycoproteins (SLGs) and the S-locus Receptor Kinase (SRK). SRK consists of an SLG-like extracellular domain, a transmembrane domain and an intercellular serine/threonine domain. SLG and SCR expression occurs just before the flower opens, primarily in the stigma papilla cells. They also exhibit allelic sequence diversity (Takayama and Isogai, 2005). The male determinant genes named *SP11* (*S-locus protein 11*) or *SCR* (*S-locus cysteine rich*) code for the secretion of small, cysteine-rich proteins, SP11/SCR, in anther tapetum cells and gametophytically in the microspores (Takayama *et al.*, 2000). These genes are tightly linked and behave as a single Mendelian locus, displaying multiple allelic versions (Takayama and Isogai, 2003). The SI-response occurs when stigma and pollen share at least one allele. Upon pollination, SP11, carried in the pollen coat, penetrates the papilla cell wall and binds with SRK. The binding induces autophosphorylation of SRK starting a signalling cascade that causes the rejection of self pollen by preventing hydration and further development of the pollen tube (Takayama and Isogai, 2005). SLGs are not present or active in all members of the mustard family (Kusaba *et al.*, 2001). If there is a compatible reaction, the papilla cells provide moisture for pollen germination, however with self-pollination the absorption of water and germination are disrupted (Dickinson, 1995) and a callus deposition may occur at the attachment site (see Hinata *et al.*, 1994 for review). If the incompatible pollen is able to germinate, the pollen tube growth is slowed or inhibited due to the inability of the pollen tube to grow through the papilla cell wall.

164. For vegetable crops, the National Vegetable Research Station at Wellesbourne, England, maintains a collection of all known S alleles together with their internationally accepted nomenclature (Dickson and Wallace, 1986). The genotypes of most self-incompatible *Brassica* plants will be heterozygous at the S locus since cross fertilization is mandated by the self-incompatibility specificities of the S alleles present. Dominant and recessive interactions occur between S-haplotypes (Thompson and Taylor, 1966). The interaction is complex with the S-haplotypes classified as class I or class II, based on the nucleotide sequences of *SGL* and *SRK* alleles (Nasrallah *et al.*, 1991). The class I S-haplotypes are normally dominant over class II S-haplotypes in the pollen. The S allele specificities of the pollen and the stigma can be co-dominant, which occurs more frequently than the dominance/recessive. Dominance/recessive relationships occur more frequently in the pollen than the stigma and are not identical for S alleles between the stigma and pollen (Watanabe and Hinata, 1999). Among the *SP11/SCR* alleles in Class I S-haplotypes, the dominance relationship is non-linear whereas Class II S-haplotypes exhibit linear dominance (Takayama and Isogai, 2003; Hatakeyama *et al.*, 1998). The molecular mechanism of the dominance relationship in the stigma is an active area of investigation and is not fully understood (Takayama and Isogai, 2003; Fujimoto *et al.*, 2006). Selfed seed of most incompatible plants can be obtained through bud pollination *i.e.* applying pollen to the stigma 1 to 4 days before the flower opens since the SGLs and SRK are not expressed until just prior to the flower opening (Takayama and Isogai, 2005). Various other methods have been utilized to overcome the SI system including stigma mutilation, stigma treatment with various organic acids, solvents, oils and ionic solutions, thermally aided pollination as well as elevated carbon dioxide treatment and momentary high temperature application (Hinata *et al.*, 1994).

165. The SI system of *S*-haplotypes has been used by vegetable breeders to capture heterosis by producing top cross, double, or three way F₁ hybrids. However, from the perspective of intra- and interspecific outcrossing in the field, it has been noted that the incidence of interspecific crossing in mixed species populations is likely to increase as the number of plants in the self-incompatible species decreases, due to scarcity of pollen of the same species and increasing pollen competition from other nearby species.

166. Although nearly all the mono-genomic *Brassica* species are self-incompatible, the natural amphidiploids species, *B. napus*, *B. juncea* and *B. carinata* are all self-compatible (Takahata and Hinata, 1980). Okamoto *et al.* (2007) note that interspecific crosses between *B. rapa* and *B. oleracea* are difficult to make and, when the chromosome complement is doubled, produce self-incompatible amphidiploids plants (Beschoner *et al.*, 1995; Nishi, 1968). They suggest that a single mutation in a dominant *S*-haplotype could result in a self-compatible *B. napus* plant that could reproduce itself through the production of self seed. Amphidiploid plants without such a mutation would be forced to cross with one or the other diploid parent and rapidly be assimilated into one or the other parent species. Fujimoto *et al.* (2006) provide evidence for such mutations in *B. rapa* and *B. oleracea*.

SECTION IV – INTERSPECIFIC HYBRIDISATION AND INTROGRESSION

4.1 Introduction

167. With the introduction of genetically modified (GM) *B. napus*, the potential for inserted genes to transfer and introgress into related Brassicaceae species has been the subject of much speculation and research. There are many conditions which have to be met for such an event to occur. First, the cross of interest must occur. However, crossing success depends on a series of preconditions that include physical proximity of the parents, pollen movement and longevity, synchrony of flowering, breeding system of the parents, flower characteristics, pollen-style compatibility, and competitiveness of foreign pollen. If all these pre-fertilization conditions are met the next series of hurdles include sexual compatibility, embryo-endosperm imbalance as well as hybrid fertility and viability in nature. In addition, the hybrid must have sufficient fitness to backcross with the recipient parent producing fertile progeny through several generations. For example, Wei and Darmency (2008) found crosses between male sterile *B. napus* and *B. juncea*, *B. nigra*, *H. incana* and *R. raphanistrum* produced only small seed, resulting in poor seedling establishment of the hybrids under field conditions. Even if all the conditions are met, introgression will not occur unless there is pairing between a chromosome of the recipient parent and a donor parent chromosome segment that carries the inserted gene. Gene transfer cannot occur in nature if any one of these requirements is not met. However, it has been speculated that strong selection pressure over many backcross generations could result in the transgene existing in a stable strain carrying an extra chromosome pair (Chèvre *et al.*, 2001).

168. Modern researchers have overcome many of the natural barriers to interspecific and intergeneric crosses within the tribe Brassicaceae. Techniques such as ovule, ovary and embryo culture, as well as protoplast fusion have produced hybrids that would otherwise fail due to sexual barriers. Success has also been achieved by crossing induced polyploids from one or both parents. Such techniques have been used to try to integrate important agronomic or quality traits from a foreign species into a cultivated crop. However, success using such techniques is no indication that the same result could occur through sexual crossing in nature.

169. The development of male sterile *B. napus* parental lines, for the production of commercial varieties, has also provided a means to investigate intraspecific, interspecific and intergeneric crossing on a field scale, without pollen competition. The results have shown that where male sterile plants were used, the frequency of interspecific crosses was significantly higher as indicated in the following species cross reports below. Thus, the presence of male sterile *B. napus* plants in commercial fields was seen as increasing the incidence and/or risk of unwanted species hybrids. Some of the first developed hybrid *B. napus* varieties used a seed production system termed 'synthetic hybrids'. Commercial production fields growing such hybrids consisted of about 80 to 90% male sterile hybrid plants with the remaining fully fertile plants (10 to 20%) providing the pollen cloud necessary to fertilize the male sterile plants in the rest of the field. Fortunately, this 'synthetic hybrid' system has been replaced with new systems that reverse the ratio of fully fertile to male sterile plants in commercial hybrid fields. Today only a small percentage (15 to 20%) of male sterile plants may occur as off types in these hybrid varieties. Such plants would be saturated with pollen from the surrounding *B. napus* plants, thus greatly reducing the risk of pollination by a foreign pollen source.

170. Chèvre *et al.* (2004) identified 14 species related to *B. napus* to which gene introgression from *B. napus* could be of concern to oilseed rape growing countries in Europe and North America. The reports of interspecific and intergeneric sexual crossing attempts between these species and *B. napus* are summarized in Table 13. Each species cross is discussed in the following paragraphs. Warwick *et al.* (2009b) have compiled a complete list of reports on interspecific and intergeneric hybridisation within the Brassicaceae that includes studies that use sexual as well as special techniques to effect a cross.

Table 13. Interspecific and intergeneric sexual crossing attempts, degree of success and potential for gene introgression¹

Interspecific cross	Sexual cross	Field cross	Seeds /cross	BC ♂	BC ♀	Potential		References
						Natural cross	Introg resion	
<i>Brassica napus</i>								
<i>B. napus</i> × <i>B. carinata</i> <i>B. carinata</i> × <i>B. napus</i>	Y F, Y,	NR NR	8 <1		Y Y	L L	L L	U 1935; Roy 1977, 1980; Alam <i>et al.</i> 1992; Gupta 1997; Rashid <i>et al.</i> 1994; Fernandez-Escobar <i>et al.</i> 1988; Sacristan & Gerdemann 1986 ; Navabi <i>et al.</i> 2010
<i>B. napus</i> × <i>B. juncea</i> <i>B. juncea</i> × <i>B. napus</i>	Y Y	Y Y	4 0.54	Y Y	Y Y	H H	H H	Bing <i>et al.</i> 1991, 1996; Alam <i>et al.</i> 1992; Frello <i>et al.</i> 1995; Jørgensen <i>et al.</i> 1998, 1999; GoshDastidar & Varma 1999; Choudary & Joshi 1999; Kirti <i>et al.</i> 1995; Davey 1959; Sharma & Singh 1992; Heyn 1977; Roy 1980; Dhillon <i>et al.</i> 1985; Shpota & Podkolzina 1986; Sacristan & Gerdemann 1986; Roy 1984; Wei and Darmency, 2008
<i>B. napus</i> × <i>B. fruticulosa</i> <i>B. fruticulosa</i> × <i>B. napus</i>	Y Y	NR NR	0.008 F			VL VL	VL EL	Heyn 1977; Plümper 1995; Siemens 2002; Salisbury 2002
<i>B. napus</i> × <i>B. maurorum</i> <i>B. maurorum</i> × <i>B. napus</i>	Y					EL	EL	Bijral <i>et al.</i> 1995
<i>B. napus</i> × <i>B. nigra</i> <i>B. nigra</i> × <i>B. napus</i>	Y	Y F	0–0.09 0.01	Y F	F F	L VL	L L	Bing <i>et al.</i> 1991; This <i>et al.</i> 1990; Brown and Brown 1996; Struss, <i>et al.</i> 1991; Daniels <i>et al.</i> 2005; Wei and Darmency, 2008
<i>B. napus</i> × <i>B. oleracea</i> <i>B. oleracea</i> × <i>B. napus</i>	Y Y	NR Y		Y	Y	VL VL	VL VL	Gupta 1997; Ford <i>et al.</i> 2006
<i>B. napus</i> × <i>B. rapa</i> <i>B. rapa</i> × <i>B. napus</i>	Y Y	Y Y	Many Many	Y Y	Y Y	H H	H H	Bing <i>et al.</i> 1991, 1996; Brown & Brown 1996; Gupta 1997; Jørgensen & Andersen 1994; Landbo & Jørgensen 1997; Mikkelsen <i>et al.</i> 1996; Vijayakumar <i>et al.</i> 1994; Metz <i>et al.</i> 1997; Choudhary & Joshi 1999; Daniels <i>et al.</i> 2005
<i>B. napus</i> × <i>B. tournefortii</i> <i>B. tournefortii</i> × <i>B. napus</i>	Y F	NR	0.69			L VL	L VL	Nagpal <i>et al.</i> 1996; Gupta 1997; Lokanadha & Saria 1994; Liu <i>et al.</i> 1996; Salisbury 2002
<i>B. napus</i> × <i>D. catholica</i> <i>D. catholica</i> × <i>B. napus</i>	Y NR	NR NR	NR NR	NR NR	NR NR	VL EL	VL EL	Bijral & Sharma 1998
<i>B. napus</i> × <i>D. muralis</i> <i>D. muralis</i> × <i>B. napus</i>	Y NR	NR NR	0.28			L	VL	Bijral & Sharma 1996a
<i>B. napus</i> × <i>Eruca sativa</i> <i>Eruca sativa</i> × <i>B. napus</i>	Y NR	NR NR				L	VL	Bijral & Sharma 1996b
<i>B. napus</i> × <i>Erucastrum gallicum</i> <i>Erucastrum gallicum</i> × <i>B. napus</i>	Y F	F F	0.1 0	Y	Y	VL VL	VL VL	Lefol <i>et al.</i> 1997; Batra <i>et al.</i> 1989; Warwick <i>et al.</i> 2003.
<i>B. napus</i> × <i>H. incana</i> <i>H. incana</i> × <i>B. napus</i>	Y Y	Y Y	2 2×10^{-5}	Y	Y Y	H H	L L	Chadoeuf <i>et al.</i> 1998; Lefol <i>et al.</i> 1991, 1995, 1996b; Eber <i>et al.</i> 1994; Chèvre <i>et al.</i> 1996; Kerlan <i>et al.</i> 1992, 1993 ; Wei and Darmency 2008

ENV/JM/MONO(2012)41

<i>B. napus</i> × <i>R. raphanistrum</i> <i>R. raphanistrum</i> × <i>B. napus</i>	Y Y	Y F	Y $10^{-4,-8}$	Y Y	Y Y	H VL	L VL	Baranger <i>et al.</i> 1995; Chadoeuf <i>et al.</i> 1998; Darmency <i>et al.</i> 1998; Eber <i>et al.</i> 1994; Lefol <i>et al.</i> 1997; Rieger <i>et al.</i> 1999; Chèvre <i>et al.</i> 1997a, 1998; Wei and Darmency 2008
<i>B. napus</i> × <i>R. sativus</i> <i>R. sativus</i> × <i>B. napus</i>	Y NR	NR F	0.6 0	NR NR	NR NR	VL VL	EL EL	Gupta 1997; Ammitzbøll & Jørgensen 2006
<i>B. napus</i> × <i>S. alba</i> <i>S. alba</i> × <i>B. napus</i>	Y F	NR NR	Y			VL EL	EL EL	Bijral <i>et al.</i> 1993; Ripley & Arnison 1990; Mathias 1991; Lelivelt <i>et al.</i> 1993; Chèvre <i>et al.</i> 1994; Brown <i>et al.</i> 1997; Sridevi & Saria 1996.
<i>B. napus</i> × <i>S. arvensis</i> <i>S. arvensis</i> × <i>B. napus</i>	Y Y	F F	0.18 F		F	L EL	VL EL	Bing <i>et al.</i> 1991; Moyes <i>et al.</i> 2002; Inomata 1988; Brown <i>et al.</i> 1996; Sweet <i>et al.</i> 1997; Eastham and Sweet 2002; Daniels <i>et al.</i> 2005; Lefol <i>et al.</i> 1996b
<i>B. napus</i> × <i>D. erucoides</i> <i>D. erucoides</i> × <i>B. napus</i>	NR Y	NR NR			Y		VL VL	Ringdahl <i>et al.</i> 1987

¹Y = Successful cross by hand pollination or in the field, F = Cross attempted but failed, NR= Not reported.

Probability of crossing in nature and/or gene introgression H = High, L = Low, VL = Very low, EL = extremely low.

4.1.1 *B. napus* – *Raphanus raphanistrum*

171. *R. raphanistrum* is an economically damaging weed with a worldwide distribution but its range is limited to areas with acid soils. Hand crosses between *B. napus* and *R. raphanistrum* have produced reciprocal hybrids with a higher number of hybrids obtained with *B. napus* as the female (Kerlan *et al.*, 1992; Chèvre *et al.*, 1996). In France, when *R. raphanistrum* served as the female, only three hybrids have been identified even though tens of thousands of seeds were examined (Eber *et al.*, 1994; Baranger *et al.*, 1995; Chèvre *et al.*, 1997b, 1998, 2000; Darmency *et al.*, 1998; Darmency and Fleury, 2000). Chèvre *et al.* (2000) estimated the hybridisation frequency to be 10^{-7} to 10^{-5} while Australian and Canadian studies reported respective frequencies of 4×10^{-8} (Rieger *et al.*, 2001) and 3×10^{-5} (Warwick *et al.*, 2003). Guéritaine *et al.* (2003) found that under field conditions the F₁ hybrid emergence was lower and slower and seedling survival significantly less both parents. A six-year United Kingdom monitoring programme of natural populations of *R. raphanistrum* growing near fields of HR *B. napus*, showed no evidence of intergeneric crossing (Eastman and Sweet, 2002). Similarly in the United Kingdom, Daniels *et al.* (2005) found no *R. raphanistrum* × *B. napus* plants or progeny when they sampled *R. raphanistrum* plants growing in or near four fields sown to glufosinate resistant *B. napus*. Further, no hybrids were found in a Swiss survey (Thalmann *et al.*, 2001). When *R. raphanistrum* was the female, no hybrids were found in any of these studies. The frequency of hybridisation can vary depending on the *B. napus* parental variety and the population source of *R. raphanistrum*. When *B. napus* male sterile plants were used as females, the frequency of hybrids was greatly increased, ranging from <0.2% (Chèvre *et al.*, 1996; 2000) to as high as 90% in Danish and French field trials (Eber *et al.*, 1994; Baranger *et al.*, 1995; Ammitzbøll and Jørgensen, 2006). These findings would be of concern if the use of synthetic hybrids became standard as the vast majority of plants in commercial oilseed rape fields would be male sterile. However, as indicated earlier this hybrid system has now been phased out.

172. In the *B. napus* by *R. raphanistrum* cross, the majority of the F₁ hybrids had half the chromosomes of each species (ACRr, 2n=28) while one hybrid had all the chromosomes of *R. raphanistrum* and half the *B. napus* chromosomes (RrRrAC, 2n=37) (Chèvre *et al.*, 2000). Thus the fertility of the hybrids is very low (Baranger *et al.*, 1995; Chèvre *et al.*, 1996, 1998; Darmency *et al.*, 1998; Pinder *et al.*, 1999; Thalmann *et al.*, 2001; Warwick *et al.*, 2003). However, Rieger *et al.* (2001) reported two fertile amphidiploids hybrids with a genome complement of AACCRrRr, 2n=56. Chèvre *et al.* (2000) also reported four fertile amphidiploids but questioned their genetic stability due to the presence of univalents and multi/quadrivalents at meiosis. The fitness of F₁ hybrids produced on *B. napus* male sterile plants was assessed in the field by Guéritaine *et al.* (2003). They found that the hybrids were slower to emerge and less likely survive than either parent, particularly when subjected to crop competition. The hybrids also flowered later than either parent, which limited the opportunities for backcrossing to *R. raphanistrum*. It should also be noted that if crossing between these species were to occur, it would most likely take place in a field of oilseed rape. Thus, most of the crossed seed would be harvested and only a very small proportion of the original hybrid seed would remain (Rieger *et al.*, 2001). The few surviving hybrids would germinate among *B. napus* volunteers with backcrosses to *B. napus* much more likely than with wild radish.

173. When *B. napus* herbicide resistant (HR) hybrids were surrounded by *R. raphanistrum* plants in the field, the seed set was less than one seed per hybrid plant (Darmency *et al.*, 1995). Despite the low fertility and poor fitness of the hybrids, the fertility and fitness of the backcross progeny improved with each backcross generation but the percentage of HR plants decreased (Chèvre *et al.*, 1997b, 1998, 1999; Darmency *et al.*, 1998; Benabdelmouna *et al.*, 2003; Guéritaine *et al.*, 2003). In each generation the progenies were selected for herbicide tolerance and only HR plants advanced to the next backcross (BC). None of the HR plants in the BC₃ to BC₅ had the chromosome number of *R. raphanistrum* (2n=18) indicating that no genomic introgression had occurred (Chèvre *et al.*, 1998; Guéritaine *et al.*, 2002).

Backcrossing to *R. raphanistrum* was continued up to BC₇ followed by random mating and selection pressure in generations (G) G8 through G11 (Al Mouemar and Darmency, 2004). Root tip cytology of HR G9 plants established that all 32 plants were either carrying extra chromosomes or, as indicated by the non Mendelian segregation of the progeny, did not have the HR gene stably introgressed into the *R. raphanistrum* genome. The authors concluded that “the prospect of stable introgression of herbicide tolerance to wild radish in nature seems remote”.

4.1.2 *B. napus* - *B. rapa*

174. *B. rapa*, a widespread weed of cultivated and disturbed lands, is also grown as a vegetable and oilseed crop. The weedy type differs from the cultivated oilseed form only in the primary seed dormancy trait. Plant breeders of *B. rapa* and *B. napus* have known for many years that these two species readily cross in nature and they were not surprised that natural interspecies gene flow was demonstrated in several countries, including Denmark (Landbo *et al.*, 1996; Hansen *et al.*, 2001), Canada (Warwick *et al.*, 2003; Beckie *et al.*, 2003; Yoshimura *et al.*, 2006), the United Kingdom (Daniels *et al.*, 2005; Allainguillaume *et al.*, 2006), the United States (Halfhill *et al.*, 2002) and the Czech Republic (Bielikova and Rakousky, 2001).

175. Normally the highest hybrid frequencies occur when individual, self-incompatible plants of *B. rapa* are present in *B. napus* fields (Jørgensen *et al.*, 1996). In the field, more hybrids are produced on *B. rapa* plants than on *B. napus* plants (Jørgensen and Andersen, 1994; Hauser *et al.*, 1997; Jørgensen *et al.*, 1998), primarily due to their respective self-incompatible and self-compatible breeding systems. However, in reciprocal hand crosses, more hybrids per cross are found when *B. napus* is the female (Downey, *et al.*, 1980). Natural interspecific hybridisation between *B. rapa* and *B. napus* varies widely, depending on the environment under which the plants develop and the design of the experiment, particularly the ratio of *B. rapa* to *B. napus* plants. In Danish trials, up to 95% hybrids were found in *B. rapa* progeny (Mikkelsen *et al.*, 1996), while in New Zealand Palmer (1962) reported a range of 10-88%. In contrast, others in Canada (Bing *et al.*, 1991) and England (Wilkinson *et al.*, 2000) found less than 1% hybridisation. In Canadian field experiments (two in the East and one in the West), *B. rapa* plants were grown at various positions within and alongside HR *B. napus* plots. Approximately 7% of the harvested *B. rapa* seed was found to be triploid hybrids (AAC, 2n=29) (Warwick *et al.*, 2003). Similarly, in commercial *B. napus* fields, containing sparse populations of weedy *B. rapa*, the hybrid frequency was approximately 13.6%. However, the frequency of hybrids from weedy *B. rapa* growing in a harvested corn field with HR *B. napus* volunteers was only 0.023% (Warwick *et al.*, 2003). In New Zealand field studies, with ratios of *B. rapa* to *B. napus* plants of 1:400 and 1:1, the hybrid frequencies ranged from 2.1 to 0.06% with the total for the experiment of 0.46% (Jenkins *et al.*, 2001). A study of *B. rapa* populations growing outside *B. napus* fields in the United Kingdom found few hybrids (0.4 to 4.5%) in 7% of the populations, and no hybrids in the remaining 93% (Scott and Wilkinson, 1998).

176. Hybridisation also occurs with *B. napus* as the female; however most of the hybrid seed that is formed will be removed from the field at harvest. Any hybrids that volunteer the following year are almost certain to be surrounded by *B. napus* volunteers. Thus any backcrosses will quickly revert to *B. napus* form and chromosome number.

177. Compared to the parent species, natural interspecific hybrids have reduced fertility and poor seed set, averaging 2-5 seeds per pod (Jørgensen and Andersen, 1994). The survival rate of hybrid seedlings is also low with <2% survival (Scott and Wilkinson, 1998), reducing the rate of introgression (Jørgensen *et al.*, 1996; Sweet *et al.*, 1999b). Interspecific vegetative and reproductive competition strongly impacts the relative and absolute fitness of the hybrids (Hauser *et al.*, 2001). When Mikkelsen *et al.* (1996) sowed interspecific hybrids within a *B. napus* population, no *B. rapa* × hybrid BC progeny were found among 2,000 offspring raised from 30 *B. rapa* plants. Further, the hybrids lacked primary seed dormancy (Linder,

1998). This may explain why Landbo and Jørgensen (1997) found interspecific hybrids in feral *B. rapa* populations, but no hybrid seed in the seed banks at those sites. Introgression of HR transgenes from *B. napus* to *B. rapa* has occurred in Europe (Jørgensen, 1999; Hansen *et al.*, 2001; Norris and Sweet, 2002). However, no evidence of introgression was found in seed samples taken from *B. rapa* plants in the field, indicating there may be selection pressure against backcross individuals (Norris and Sweet, 2002).

178. The rate of introgression of a *B. napus* trait into the *B. rapa* genome will greatly depend on the selection pressure exerted on the gene (Scott and Wilkinson, 1998; Sweet *et al.*, 1999a; Snow and Jørgensen, 1999). The introgression of a gene into the *B. rapa* genome might be slowed by positioning it in the C genome of *B. napus* but the findings of Stewart *et al.* (2002), where 12 independent *B. napus* transformations distributed across both the A and C genomes all generated backcrosses at similar rates, suggests this theory may not be valid. Leflon *et al.* (2006) found that the transmission rate of the C chromosomes depended on which C chromosome was involved, and that a gene carried on a C chromosome is less likely to be transferred in a *B. rapa* background than if it was on an A chromosome. The presence of an introgressed HR gene in *B. rapa* did not increase its fitness or weediness relative to conventional non-GM *B. rapa* including glufosinate resistant BC₃ hybrids (Snow *et al.*, 1999) or BC₂F₂ glyphosate hybrids (Warwick, 2007). It should be kept in mind that, if introgression of an R gene does occur, the resulting HR *B. rapa* plant(s) can be controlled with other herbicides or cultivation. In Canada, with 16 years of experience growing millions of hectares of HR *B. napus* each year, no significant agronomic problems with HR *B. rapa* have been encountered (Beckie *et al.*, 2006).

4.1.3 *B. napus* – *Hirschfeldia incana*

179. *H. incana* is an important weed in some European countries and eastern Australia, but not in Canada or the Indian sub-continent. Hand crosses between *B. napus* and *H. incana* produced 1.3 and 3.1 hybrids per 100 pollinations when *H. incana* and *B. napus*, respectively, were used as the female (Kerlan *et al.*, 1992). In the field, when male sterile *B. napus* was used as the female, 1.9 hybrids were recorded per pollinated flower (Eber *et al.*, 1994). However, in three years of field trials, isolated *H. incana* plants growing in *B. napus* plots only produced 0.6 hybrid seeds per plant. Most F₁ plants had reduced fitness with seedling emergence over three years being <1% (Chadoeuf *et al.*, 1998). However, some hybrids were at least as competitive as the wild parent (Eber *et al.*, 1994; Lefol *et al.*, 1996a)

180. When F₁ plants were backcrossed to *H. incana* and only HR progeny were selected for further backcrossing, fewer seeds were produced in each generation. BC₃ produced only one seed with no viable seeds obtained in BC₄ (Darmency and Fleury, 2000). It is suggested that a *H. incana* gene inhibits homeologous pairing resulting in an expulsion of *B. napus* chromosomes (Kerlan *et al.*, 1992; Lefol *et al.*, 1996a). Thus, although interspecific F₁ hybrids will frequently occur in areas where *H. incana* is prevalent, their persistence will be short and the possibility of gene introgression from *B. napus* remote.

4.1.4 *B. napus* – *B. juncea*

181. *B. juncea* is primarily a crop plant grown in China, Russia and on the Indian sub-continent as a major source of edible oil, and in Canada and a few other countries as a condiment crop. However, it is present as a weed in parts of Europe and Australia. Since *B. juncea* (AABB) and *B. napus* (AACC) have a common genome, the chance of interspecific crossing is enhanced. In Canadian co-cultivation experiments, Bing *et al.* (1996) identified five interspecific hybrids in seed harvested from 469 *B. napus* plants and three out of 990 plants when *B. juncea* was the female. Jørgensen *et al.* (1998) noted that as the ratio of *B. juncea* to *B. napus* plants increased from 1:3 to 1:15 the hybridization frequency on *B. juncea* plants decreased from 2.3 to 0.3%. Warwick (2007) reported gene flow from HR *B. napus* to neighbouring

fields of *B. juncea* at a rate of 0.245% at the adjacent *B. juncea* field border and 0.030, 0.021 and 0.005% at 50, 100, and 200 m, respectively.

182. The viability of F₁ pollen is reported to be low (18-26%) (Frello *et al.*, 1995; Choudhary and Joshi, 1999; GoshDastidar and Varma, 1999), but spontaneous backcrossing with improved fertility has been reported (Alam *et al.*, 1992; Bing *et al.*, 1991, 1996; Jørgensen, 1999). Given this background of results, the introgression of genes from *B. napus* could be expected to occur where these two species are widely grown.

4.1.5 *B. napus* – *Sinapis arvensis*

183. *S. arvensis* is a serious weed in all oilseed rape growing countries. In a five year study of *S. arvensis* growing in and around GM *B. napus* crops in the United Kingdom, Sweet *et al.* (1997) and Norris *et al.* (unpublished, cited in Eastham and Sweet, 2002) failed to detect any hybridisation with *S. arvensis*. Also in the United Kingdom Daniels *et al.* (2005) tested 60,768 progeny from 818 *S. arvensis* plants, growing in or close to 23 glufosinate resistant *B. napus* fields. No resistant plants were found in the parents or their progeny. Similarly, Warwick *et al.* (2003) found no interspecific hybrids among 43,828 *S. arvensis* progeny from plants growing in HR *B. napus* fields in western Canada. Bing *et al.* (1996) also found no hybrids in Canadian co-cultivation experiments involving the assessment of 7,500 *S. arvensis* seeds. Similar results were reported from UK trials where 9,688 *S. arvensis* seedlings were screened (Moyes *et al.*, 2002) and in France, Lefol *et al.* (1996b) found no hybrids among the 2.9 million *S. arvensis* seeds tested. However, when male sterile or emasculated *B. napus* plants were pollinated with *S. arvensis* pollen, either naturally or artificially, a small number of hybrids were obtained. Chèvre *et al.* (1996) found 0.18 hybrids per 100 pollinations while Lefol *et al.* (1996a) detected six hybrids in 50,000 seeds analysed. In hand crosses, using *S. arvensis* females from different UK and French populations, Moyes *et al.* (2002) detected one completely sterile hybrid. No such hybrid had previously been reported without embryo rescue or ovule culture (Inomata, 1988; Kerlan *et al.*, 1992; Bing *et al.*, 1991, 1996; Chèvre *et al.*, 1996; Lefol *et al.*, 1996a). All hybrids produced were weak, largely or completely sterile and unlikely to survive in nature (Moyes *et al.*, 2002). None of the hybrids were able to backcross to *S. arvensis*.

184. Daniels *et al.*, (2005) identified a single plant in the United Kingdom that they believed to be a *S. arvensis* × *B. napus* hybrid. It was growing in a patch of *S. arvensis* plants adjacent to a field that had grown a crop of glufosinate resistant *B. napus* the previous year. The hybrid classification was based on a null reaction to the application of glufosinate to a single leaf followed by a positive DNA test for the glufosinate resistance gene. However, only morphological characteristics were used to classify the plant as a *S. arvensis* × *B. napus* hybrid. The lack of any information on chromosome number and/or markers, and in the light of previous studies, the question remains as to whether the plant was indeed a *S. arvensis* × *B. napus* hybrid rather than another interspecific cross such as *B. rapa* × *B. napus*. In the words of the report's reviewer "such a finding needs to be interpreted with caution."

185. Despite the one hybrid produced by Moyes *et al.* (2002) on an emasculated *S. arvensis* plant, there is general agreement among researchers that the possibility of gene flow between *B. napus* and *S. arvensis* is extremely low (Moyes *et al.*, 2002) to nonexistent (Downey, 1999a,b).

4.1.6 *B. napus* – *Raphanus sativus*

186. *R. sativus* is a vegetable crop in many parts of the world, but when grown for seed it can escape from cultivation and colonise disturbed sites such as roadsides, fields and coastal sand dunes (Snow *et al.*, 2001). Daniels (2005) reported flowering of *R. sativus* plants could coincide with either winter or spring *B. napus*. In *R. sativus* plants growing in or near a field of glufosinate resistant *B. napus* in the United

Kingdom, Daniels *et al.* (2005) found no *R. sativus* × *B. napus* hybrids. Further, progeny from the sampled *R. sativus* plants were all susceptible to glufosinate. Hybrids between *B. napus* and *R. sativus* have been obtained in several studies with the aid of ovule culture or embryo rescue (Lelivelt *et al.*, 1993; Paulmann and Röbbelen, 1988; Sundberg and Glimelius, 1991; Metz *et al.*, 1995; Takeshita *et al.*, 1980) and also by hand pollination (Gupta, 1997). All artificially produced hybrids were male sterile. However, in natural crosses Ammitzbøll and Jørgensen (2006) obtained an average of 0.6 seeds per pod when male sterile *B. napus* plants were used as the female and a radish cultivar as the pollen parent. Huang *et al.* (2002) in hand crosses also produced many hybrids on Ogura male sterile plants. All seeds produced proved to be F₁ triploid hybrids with low pollen fertility (0–15%). It is highly probable that the presence of radish cytoplasm in the male sterile *B. napus* parent greatly facilitated *R. sativa* pollen penetration of the stigma. Further studies need to be made with this cross since *R. sativa* crosses easily with *R. raphanistrum* (Snow *et al.*, 2001).

4.1.7 *B. napus* – *Erucastrum gallicum*

187. *E. gallicum* is a self-compatible, annual or winter annual with very small seeds. It is a minor weed of cultivated fields and waste places in many oilseed rape growing countries. Batra *et al.* (1989) obtained three hybrids from the cross *E. gallicum* × *B. napus* using embryo rescue. Lefol *et al.* (1997), using reciprocal hand crosses, obtained one slow growing *B. napus* × *E. gallicum* F₁ hybrid with pollen viability of 28%. Indications were that the F₁ would not survive in competition with a *B. napus* crop. No seed was produced when *E. gallicum* served as the female parent. The F₁ hybrid was backcrossed in all combinations and many seeds were obtained when *E. gallicum* was the male and a few when *B. napus* was the female. Backcross seed from the hybrid produced plants identical to *E. gallicum*, suggesting that the *B. napus* chromosomes were lost. A survey of 22,000 seedlings of *E. gallicum* from western Canadian *B. napus* fields yielded no hybrids, indicating that the possibility of hybridisation between *B. napus* and *E. gallicum* is very low ($< 5 \times 10^{-5}$) (Warwick *et al.*, 2003).

4.1.8 *B. napus* – *B. nigra*

188. *B. nigra* is a minor weed and an occasional crop in warmer, shorter day length locations of oilseed rape growing regions. Interspecific hand crosses between *B. napus* and *B. nigra* have been difficult to obtain with some success using oilseed rape as the female (Davey, 1959; Heyn, 1977; Diederichsen and Sacristan, 1988; Nishiyama *et al.*, 1991; Bing *et al.*, 1991, 1996; Kerlan *et al.*, 1992; Struss *et al.*, 1992; Zhu *et al.*, 1993). The F₁ hybrids were moderately to highly sterile but a few F₂ and BC seeds were obtained (Bing *et al.*, 1991; Zhu *et al.*, 1993). Using controlled crosses hybridization levels were extremely low (Raybould and Gray, 1993; Scheffler and Dale, 1994). In the cross *B. napus* × *B. nigra*, Brown and Brown (1996) observed the pollen tubes of *B. nigra* were short and twisted with only a few penetrating the style. No hybrids were found in natural crosses when *B. nigra* was the female (Bing *et al.*, 1991; Leckie *et al.*, 1993; Daniels *et al.*, 2005).

4.1.9 *B. napus* – *B. oleracea* and *Brassica* vegetables

189. Gene flow from oilseed rape to *B. napus* vegetables (Swedes, rutabaga, Siberian kale) is possible since they are all within the same species. Similarly, gene flow to *B. rapa* vegetables (*e.g.* turnip, Chinese cabbage etc.) is possible since they have the A genome in common. However, *B. napus* and *B. rapa* vegetables are not considered weedy. In addition, they are generally harvested prior to flowering.

190. Hand crosses between *B. napus* and *B. oleracea* have been successful but at a very low frequency (Chiang *et al.*, 1977) and natural crosses have only been successful with the assistance of embryo rescue (Ayotte *et al.*, 1987; Takeshita *et al.*, 1980; Quazi, 1988; Habman *et al.*, 2010). However, amphidiploid F₁

hybrids were fertile and readily backcrossed to either parent (Sundberg and Glimelius, 1991; Kerlan *et al.*, 1992; Chèvre *et al.*, 1996).

191. No spontaneous hybrids between *B. napus* and *B. oleracea* were found in two United Kingdom surveys of wild *B. oleracea* populations (Scheffler and Dale, 1994; Wilkinson *et al.*, 2000). However, a later United Kingdom survey of two wild *B. oleracea* populations, growing within 25 m of *B. napus* fields, identified one triploid F₁ hybrid and nine introgressants based on flow cytometry and crop specific microsatellite markers (Ford *et al.*, 2006). The fertility of these plants has not been reported.

4.1.10 *B. napus* – *Sinapis alba*

192. *S. alba* is commercially grown as a condiment crop but weedy forms occur in the Mediterranean region and in some countries where *S. alba* is used as a green manure crop. The cross *B. napus* × *S. alba* is difficult to make even with hand pollination, usually requiring embryo or ovule culture (Ripley and Arnison, 1990; Mathias, 1991; Bijral *et al.*, 1993; Lelivelt *et al.*, 1993; Chèvre *et al.*, 1994; Brown *et al.*, 1997; Sridevi and Saria, 1996). No field crosses have been reported (Daniels *et al.*, 2005) and the possibility of such an occurrence is very low.

4.1.11 *B. napus* – Other weedy species

193. Hand crosses have been made in enclosed environments between *B. napus* and a number of weedy species within the tribe Brassiceae (*e.g.* *B. fruticulosa*, *B. tournefortii*, *B. maurorum*, *Diplotaxis muralis*, *D. tenuifolia*, *Rapistrum rugosum*, *Eruca sativa*) while protoplast fusion and embryo or ovule rescue have produced F₁ plants in *B. napus* crosses with *B. oxyrrhina*, *B. barrelieri*, *B. elongata*, *B. gravinae*, *B. souliei* and *Diplotaxis tenuisiliqua*. No field interspecific or intergeneric hybrids have been reported between *B. napus* and the above species (Salisbury, 2002).

SECTION V – ECOLOGY

5.1 Interactions in Natural and Agricultural Ecosystems

5.1.1 Glucosinolates and their ecological interaction

194. Virtually all plants of the Brassicaceae produce sulphur compounds called glucosinolates (Kjaer, 1960). Although there are some 250 of these allelochemicals that occur in 16 botanical families of the order Brassicales (Verkerk *et al.*, 2009), only about 20 are commonly found in *Brassica* species (Sarwar and Kirkegaard, 1998). A single species will usually contain significant amounts of four different glucosinolates but a single plant may contain as many as 15 different glucosinolates. They are present in varying amounts in all tissues of the plant and directly or indirectly impact their biological environment (Brown and Morra, 1997). They are the source of the flavour and odour of the *Brassica* vegetables and the hot component in mustards. The kind and quantity of glucosinolate varies within and among species and even between stages of plant development as well as between plant parts *e.g.* cotyledon, leaf, root, flower buds and seed. The highest concentration of glucosinolates is normally found in flower buds and seeds.

195. All glucosinolates have the same basic structure consisting of a β -D-thioglucose group, a sulphonated oxime group and a side chain 'R', derived from one of the amino acids, methionine, phenylalanine, tryptophane or a branched-chain amino acid (Figure 42). Glucosinolates accumulate in plant cell vacuoles. They can be broken down (hydrolyzed) by the enzyme myrosinase which is located separately in the idoblast cells. When plant cells are crushed or broken, and moisture is present, the glucosinolates and myrosinase are released and the enzyme catalyses the hydrolysis of the glucosinolates into glucose, sulphate and thiocyanates, isothiocyanates and nitriles plus sulphur (Figure 42). The intact glucosinolates have little biological activity but their thiocyanate and isothiocyanate breakdown products have broad biocidal activity (Brown and Morra, 1997).

196. The glucosinolates serve as an advance-prepared system of protection that is activated only when plant tissue is damaged by a disease or insect attack. The destruction of the plant cells results in the hydrolysis of the glucosinolates by the myrosinase enzyme, thus releasing the volatile isothiocyanates that have a wide spectrum of anti-microbial effects and act as attractants or repellents to some insects and herbivores (Vašák, 2002; Brown and Morra, 1997; Fenwick *et al.*, 1983).

197. *Brassica* crops are also used as biofumigants based on the release of the bioactive isothiocyanates in the soil when seed meal amendments or green manure are incorporated, or *Brassica* crops are used in the rotation (Brown and Morra, 1997). It is also suspected that the volatile isothiocyanates, from residue of *Brassica* crops, result in inhibitory effects on some subsequent crops (see section 5.5 Allelopathy).

198. The glucosinolates also impact on the health and nutrition of animals and humans as well as the quality and usefulness of products from *Brassica* crops. These aspects are discussed in the following section.

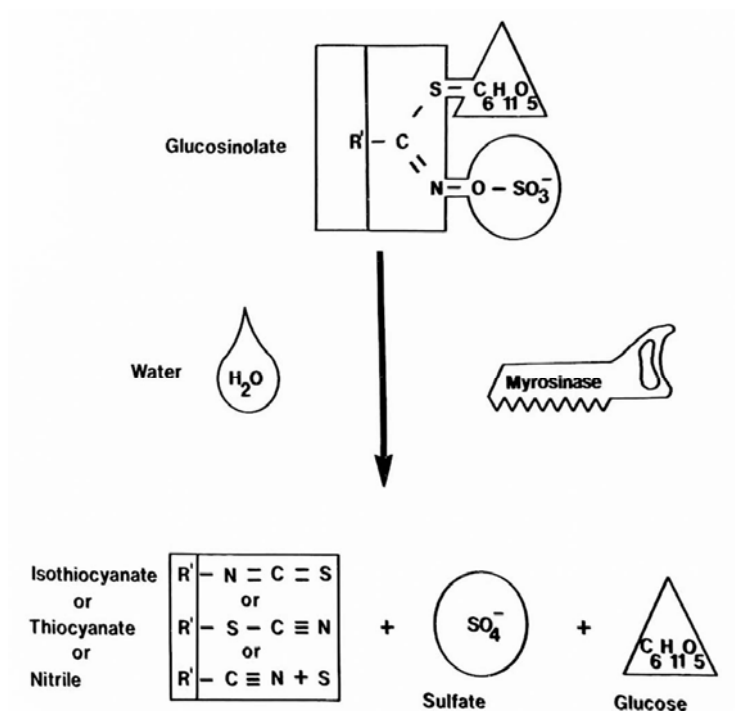


Figure 42. Glucosinolate chemical structure and enzymatic breakdown products formed in broken *Brassica* plant cells with moisture

5.1.2 Damaging insects

199. *Brassica* species are important components of temperate climate ecosystems. They provide forage for many insects as well as wild life. The complex of insects that feed upon the Brassicas is one of the important factors limiting the production of commercial *Brassica* crops (Ekbohm, 1995; Lamb, 1989). Brassicaceous plants produce a family of sulphur compounds called glucosinolates whose breakdown products are attractants and stimuli for feeding and oviposition but, on the other hand, act as deterrents or toxins for herbivores not adapted to plants of the Brassicaceae. A list of insects important to *Brassica* plants is given in Annex, Table A.1. Some of the more important insects are discussed below.

5.1.2.1 *Phyllotreta* spp. - Flea beetles

200. Flea beetles feed on spring sown seedlings and in some years the second generation may attack green foliage and pods in the fall. Several species of flea beetles occur in different *Brassica* growing areas of the world. Damage by these small beetles is characterised by feeding holes in cotyledons and first true leaves and is most severe under warm, dry conditions. Some Brassicaceous species (e.g. *Sinapis alba*, *B. villosa*) avoid damage due to the presence of hairs (trichomes) on cotyledons, leaves and stems. Attempts at biological control have not been successful, but research is underway to develop *B. napus* plants expressing large numbers of trichomes as a means of defence (Gruber *et al.*, 2006). The primary control measure is insecticidal seed dressings.

5.1.2.2 *Psylliodes chrysocephala* – Cabbage stem flea beetle

201. This beetle is one of the most important pests of oilseed rape in Europe (Ekbohm, 1995). Eggs, laid by adults in the soil at the base of seedlings, produce larvae that eat into leaf stocks and later into the stem and base of the biennial plants, where the larvae overwinter. Feeding damage results in weakened plants, resulting in reduced yield and winter kill. Control is dependent upon insecticide sprays.

5.1.2.3 *Ceutorhynchus* spp. – Stem weevils

202. Both *C. napi* and *C. quadridens* are important pests in continental Europe. The weevils overwinter as adults and lay their eggs on leaf petioles of overwintered *Brassica* plants. The larvae eat into and feed in the stems resulting in weakened and broken plants. Insecticide sprays are used for control.

5.1.2.4 Aphid species

203. Three species of aphids can be of economic importance on Brassicaceous plants (Ekbohm, 1995). *Lipaphis erysimi* and *Brevicoryne brassicae* prefer Brassicaceae hosts while *Myzus persicae* is polyphagous. On the Indian sub-continent *L. erysimi* is a very serious pest capable of reducing oilseed mustard yields by 50%. In temperate zones *B. brassicae* is a common pest of vegetable Brassicas and occasionally of oilseed crops. Suction feeding causes a direct loss of vigour and yield. *M. persicae* also causes indirect damage as a vector of beet western yellow virus (BWYV) (Hill *et al.*, 1991). Insecticide sprays can be used for control but care must be taken not to kill beneficial insects present during flowering.

5.1.2.5 *Lepidoptera* species

204. The lepidopteron pests occur sporadically and can have more than one generation per year. The eggs are laid on the leaves where the larvae feed. In Canada, a second generation of the diamondback moth (*Plutella xylostella*) may also attack pods of oilseed rape. The diamondback and *Pieris brassicae* (the large cabbage white butterfly) are also important pests of vegetable crops where their leaf damage affects market value. Chemical control is applied where populations warrant.

5.1.2.6 *Meligethes* species

205. Pollen beetles are important pests of both spring and winter oilseed rape in Europe. Adult beetles move onto the crop from unrelated early flowering plants to feed on pollen from open flowers and to lay eggs in unopened buds. The larvae emerge and eat the stamens, causing buds to abort. The final instar larvae fall to the ground and pupate with the new generation emerging in July into August (Ekbohm, 1995). Pyrethroids are the most commonly used chemical control.

5.1.2.7 *Ceuthorhynchus assimilis* – Seed pod weevil

206. This weevil occurs in both Europe and North America. It has one generation per year, emerging from overwintering sites in the late spring to feed on the crop. The main damage is done to the seed pods. The adults make small holes in the pods to feed on the seeds within and to lay eggs. The larvae eat their way out of the pods, drop to the ground and pupate. The weevil has only recently invaded the oilseed rape growing area of western Canada. Control is by chemical sprays.

5.1.2.8 *Dasineura brassicae* – Pod midge

207. The pod midge is a European pest that uses the small holes in the pods, made by the seed pod weevil, for oviposition. The larvae eat the developing seed and cause the pods to open losing their seed. The larvae overwinter in cocoons in the soil, pupate in the late spring and fly to the plants to oviposition, living only a few days (Ekbohm, 1995). Early spraying for the pollen beetle can provide control of pod midge and other pod pests.

5.2 Beneficial Insects

208. The interaction between bees, both farmed and wild, and *Brassica* plants are mutually beneficial. The bees aid fertilization and receive nectar and pollen in return. Where grown, oilseed rape and mustard provide productive bee pasture while the fertilizing activities of the bees are essential for the production of hybrid seed and tend to increase seed yields of commercial fields.

5.3 Animal Interaction

209. Succulent *Brassica* plants attract many foraging animals including rabbits, rodents and deer to name a few. Winter oilseed rape is an important winter pasture for wild deer and other animals. Ruminant animals, both wild and domestic, under certain circumstances, can become ill from grazing kale or winter oilseed rape crops (Marquard and Walker, 1995). The toxic compound responsible is dimethyldisulphide that arises from the breakdown products of glucosinolates and S-methylcysteine sulphoxide (SMCO), also known as the kale anaemia factor (Maxwell, 1981). Birds often feed on fall germinating seedlings and on the developing seed in the pod.

5.4 Soil Microbial Interaction

210. The genetic makeup of crop plants can influence the composition of the soil microbial community in which they grow. However, the interaction between plants and their residues with the soil microflora is not well understood (Dunfield and Germida, 2004). The soil microbial communities associated with the growing of conventional spring oilseed rape (both *B. napus* and *B. rapa*) and transgenic HR *B. napus* were investigated in western Canada plot trials. The soil microflora in the plots of the glyphosate resistant variety Quest differed significantly from that found in both conventional and transgenic glufosinate resistant varieties, particularly at the flowering stage. However, although the microbial diversity was altered, the effects varied by test site and plant growth stage. In addition, the change in the microbial community was temporary as no differences were found the following spring (Dunfield and Germida, 2001, 2003, 2004; Siciliano and Germida, 1999).

5.5 Allelopathy

211. There have been numerous reports of inhibitory effects by *Brassica* residues on the following planting of pasture, cereal and oilseed crops (Campbell, 1959; Bell and Muller, 1973; Rice, 1984; Mason-Sedun *et al.* 1986; Horricks, 1969; Vera *et al.*, 1987). The allelopathic effects include germination inhibition, reductions in root growth, plant height, dry weight, tiller number and seed yield. Species involved in the inhibition included marrow stem kale (*B. oleracea*), oilseed and turnip rape (*B. napus*, *B. rapa*) and condiment and black mustard (*B. juncea* and *B. nigra*). The inhibiting compound(s) are leached by water from dead or decaying stems and leaves of *Brassica* vegetation. The compound(s) appear to reside in the upper soil layer for a short period and then dissipate. Mason-Sedun *et al.* (1986) compared the effect of water extracts from dry residues of four *Brassica* species on coleoptile growth of common wheat (*Triticum aestivum*). All residues significantly reduced grain yield, plant dry weight, plant height and tiller production, with greatest level of inhibition resulting from *B. juncea* residues followed by *B. nigra*, *B. napus* and *B. rapa*. Laboratory studies indicated that when stored, dry residues became less toxic over time. Waddington and Bowren (1978) found that rapeseed residue was no more toxic to barley, brome grass or alfalfa than comparable amounts of wheat residue. Normally *Brassica* residue will have been rained on well before seeding, resulting in no inhibition. Indeed there is good evidence that cereal crops are more productive following oilseed rape than another cereal (Almond *et al.*, 1986). Vera *et al.* (1987) suggested that the primary cause of the observed inhibition in western Canada may be the release of a chemical compound from volunteer oilseed rape seedlings that

are killed by cultivation at seeding time. The chemical was thought to be the indole glucosinolate, glucobrassicin, present in high concentrations in tissues of young seedlings (Röbbelen and Thies 1980).

5.6 Pathogens

212. The *Brassica* crops and their wild allies are subject to a broad range of pathogens and adverse conditions or disorders associated with non-infectious causes. Although many of the *Brassica* species have many diseases in common there are also significant differences in susceptibility among and within species. The Compendium of *Brassica* Diseases (Rimmer *et al.*, 2007) provides an authoritative and practical reference guide to disease problems in *Brassica* crops the world over. Colour plates and text describe the infectious diseases caused by fungi, oomycetes, bacteria, mollicutes, viruses and nematodes. In addition, non-infectious disorders such as those related to environmental effects, herbicide injury, and nutritional deficiencies are also described. The American Phytopathological Society (APS) also provides a listing by common and scientific name of known *Brassica* diseases and conditions at its website as reproduced in the Appendix, Table A.2. (APS, 2001)

213. Of the many *Brassica* field crops diseases listed in Table A.2, three stand out as particularly troublesome as they are pandemic and have the potential to cause major crop injury: blackleg or stem canker (*Leptosphaeria maculans*); Sclerotinia stem rot (*Sclerotinia sclerotiorum*); and clubroot (*Plasmodiophora brassicae*). To date there are few control measures for these pathogens that are fully effective and economical. Varieties with single race resistance have been developed, but the multi race pathogenicity of these fungi has made it difficult to breed varieties with long lasting resistance. However, it is anticipated that with the location of resistance genes on marker saturated genome maps, breeders will be able to bring together multiple resistance genes from both within and outside the genus that will provide long-lasting disease resistance.

SECTION VI – BREEDING IMPROVED VARIETIES

6.1 Introduction

214. The objective of all plant breeding programmes is to produce plants of greater value to the producer, the industry and the consumer. The objective is achieved by building on past advances, through the incorporation of desirable traits that impart increased yield, pest resistance, superior quality and/or utility to new varieties. To accomplish the task many related disciplines are essential, including genetics, biotechnology, agronomy, cytology, chemistry, pathology, entomology, physiology and statistics. Within the biotechnology component gene transfer and the production of transgenic varieties has attracted public attention but the discipline is much broader and includes, among others, tissue culture, protoplast fusion, dihaploid production, gene identification and cloning.

215. The essential requirement for success is genetic variation for the trait or traits of interest. The breeder will normally search for the desired trait within adapted genotypes and then the crop's world germplasm collection. If it is not present within the species but present in a related species, interspecific and intergeneric crosses and/or protoplast fusion may be attempted. If those approaches fail, induced mutation may be explored. Generally gene transfer, because of regulatory hurdles, is the last resort.

216. Valuable, new gene-controlled traits are added with each improved variety. The breeder evaluates the need and the genetic variability available and stacks desirable traits, be they large or small advances, into the genetic base that previous breeders have built. Gene stacking is the very essence of plant breeding. Breeding techniques vary with the crop being bred and its mode of pollination and reproduction. Among the commercial *Brassica* crops, both self-compatible and self-incompatible species are present so that a wide array of techniques are employed, as described below, depending on the species and the trait or traits to be introduced.

217. The application of conventional genetic manipulation in plants can have major beneficial impact on the nutritional quality and quantity of the world's food supply. A very successful example, described below, is the conversion of *Brassica* oilseed crops from a problematic commodity to the high quality productive crop we now define as canola.

218. Lipids not only make our food taste better but are required dietary ingredients. They are essential cell membrane components, regulating cell permeability and are responsible for vitamin transport as well as the starting point for hormone biosynthesis. Oils and fats are predominantly (~98%) triacylglycerols (TAGs) that consist of a three carbon chain with fatty acids attached to each carbon. The fatty acid composition of an oil determines its value, use and nutritional worth.

219. Oils from *B. juncea*, *B. rapa* and later *B. napus* have been part of the Asian diet for centuries but in Europe and the Americas they are relatively recent edible oil additions. Prior to and during the Second World War, rapeseed oil was primarily used as a lubricant for steam engines and as a lamp oil, but following the war *B. napus* and *B. rapa* oils became an important constituent of margarine. Researchers became interested the nutritional value of *Brassica* seed oils because they differed from most other edible oils in having a high percentage of long carbon chain monoenoic fatty acids, eicosenoic (C20:1) and erucic (C22:1) (Table 14). Small animal feeding studies in the late 1950's and throughout the

60's indicated the nutritional value of rapeseed oil could be substantially improved if the long chain fatty acids could be reduced to < 5% of the fatty acid total (Kramer *et al.*, 1983). Breeding and selection within the world's germplasm was successful in developing plants of *B. napus* (Stefansson *et al.*, 1961), *B. rapa* (Downey, 1964) and later *B. juncea* (Kirk and Oram, 1981) that produced oils with less than 2% erucic acid. This oil was found nutritionally superior to the high erucic oil (Kramer *et al.*, 1983) and proved to be an excellent liquid and salad oil, as well as a suitable ingredient for margarine and shortening manufacture. This new natural oil is called 'canola oil' in most countries of the world and is defined as oils from *B. napus*, *B. rapa* or *B. juncea* containing less than 2% erucic acid of the fatty acid total. The genetic blocking of the biosynthesis of eicosenoic and erucic acids resulted in an increased percentage of oleic and linoleic acids (Table 14).

Table 14. Fatty acid composition of rapeseed, canola, soybean, sunflower and linseed oils

Fatty acid	Symbol ¹	Rapeseed	Canola	Soybean	Sunflower	Linseed
		Fatty acid composition %				
Palmitic	C16:0	4.0	4.7	11.5	7.5	7.0
Stearic	C18:0	1.5	1.8	3.5	4.5	4.0
Oleic	C18:1	17.0	61.5	23.0	16.0	20.0
Linoleic	C18:2	13.0	21.0	43.0	71.0	17.0
Linolenic	C18:3	9.0	11.0	8.0	1.0	52.0
Eicosenoic	C20:1	14.5	<1.0	0.0	0.0	0.0
Erucic	C22:1	41.0	<1.0	0.0	0.0	0.0

¹ The first number denotes the number of carbon atoms in the fatty acid chain and the second number, the number of double bonds in the chain

220. When nutritionists recommended that dietary intake of saturated fat be reduced, the nutritional value of canola oil gained widespread recognition since it contains the lowest level of saturated fatty acids of any edible oil (Grundy and Denke, 1990; Gurr, 1992; Hu *et al.*, 1997) (Figure 43). Further, in 1985 Mattson and Grundy reported on the nutritional desirability of the so called 'Mediterranean diet' pointing out the health advantages of oils with a low level of saturates and high content of oleic acid. The fatty acid composition of canola oil met or exceeded the nutritional requirements of a superior edible oil, with the lowest saturate content (6 to 7%) of any edible oil and a high (58 to 60%) level of oleic (18:1n-9) that reduces the undesirable low density lipoproteins (LDLs) without reducing the desirable high density lipoproteins (HDLs).

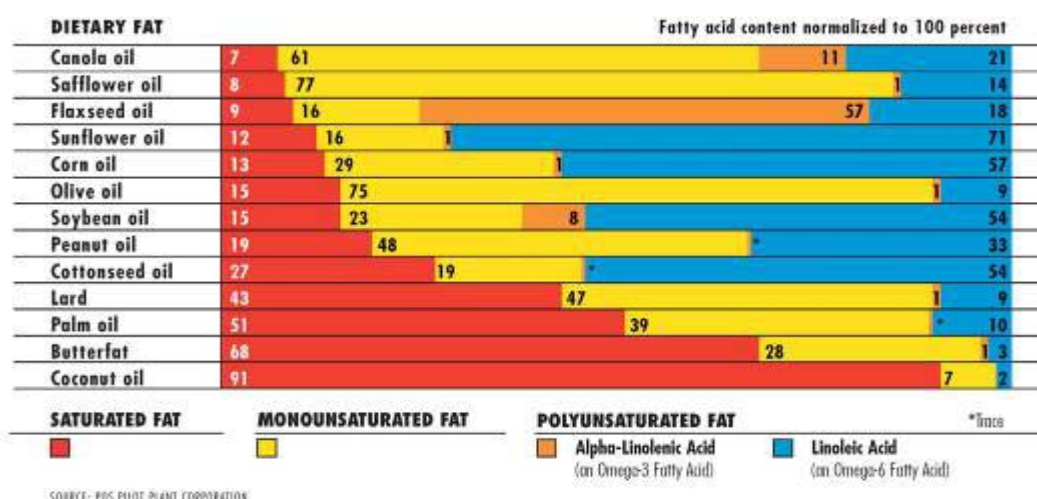


Figure 43. Canola oil compared to other edible vegetable oils as to total saturated fat content and other fatty acids

Source: analyses conducted by POS Pilot Plant Corporation, Saskatoon, Canada, data courtesy of Canola Council of Canada.

221. Plant breeders have now developed varieties that produce canola oils with less than 3% α -linolenic acid which improves the oxidative stability of the oil and reduces the development of unpleasant flavours and cooking odours (Scarth *et al.* 1988, 1995; Eskin *et al.*, 1989; Przybylski *et al.*, 1993). More recently plant breeders have combined the low linolenic trait with a reduced level of linoleic acid to provide an oil with over 70% oleic acid (Table 15) (Downey, 1996). The high oleic acid level further increases the oil's stability so that little or no hydrogenation of the oil is required, which would otherwise result in undesirable *trans* fatty acids. Canola varieties that produce this latter fatty acid composition now occupy about 10% of Canada's oilseed rape growing area.

Table 15. Fatty acid composition of canola and specialty *B. napus* varieties grown in Canada

Oil type	Fatty acid composition %							
	C16:0	C16:1	C18:0	C18:1	C18:2	C18:3	C20:1	C22:1
Canola	4.7	<1.0	1.8	61.5	21.0	11.0	<1.0	<1.0
High erucic	2.0	<1.0	2.0	13.0	12.0	9.0	7.0	54.0
Low linolenic	4.0	<1.0	2.0	64.0	27.0	2.0	1.0	<1.0
High oleic	4.0	<1.0	2.0	75.0	9.0	8.0	<1.0	<1.0

222. Oil extraction of *Brassica* oilseeds yields about 40% oil and some 60% high protein meal. The meal is used as a high quality protein supplement in diets for animals, poultry and fish. Unfortunately the plant translocates and concentrates the glucosinolates in the seed. As a result rapeseed and mustard can contain over 120 mg/g of glucosinolates per whole seed. This high concentration of glucosinolates, and their breakdown products, greatly limited the amount of traditional rapeseed meal that could be fed to non-ruminant animals, such as swine and poultry. Glucosinolates and their breakdown products reduced the palatability of the meal but more importantly they interfered with the iodine uptake by the thyroid gland and are active goitrogens. Feeding rapeseed meal to non-ruminant animals frequently resulted in poor feed efficiency and weight gains as well as reproductive difficulties (Bell, 1993). Thus the amount of seed that could be processed was determined by the limited size of the meal market. A partial solution was the inactivation of the myrosinase enzyme as the first step in the oil extraction process but enzymes in the animal gut, although less efficient, were also able to hydrolyse the glucosinolates. The answer to this problem was to breed plants with little or no glucosinolates in their seed.

223. Analytical advances in the 1960's allowed breeders to identify plants with only 10 to 12 μ moles of aliphatic glucosinolate per gram oil free meal. These plants were crossed with low erucic acid varieties to produce 'double low' or 'canola quality' varieties of *B. napus* (Stefansson, 1983), *B. rapa* (Downey and Rakow, 1987) and *B. juncea* (Love *et al.*, 1990). The reduction in glucosinolate levels now allowed canola meal to be fed at maximum economic levels to non-ruminants and canola meal became the preferred protein supplement for dairy cattle. Canola is defined as seeds of the genus *Brassica* (*Brassica napus*, *Brassica rapa* or *Brassica juncea*) from which the oil shall contain less than 2% erucic acid in its fatty acid profile and the solid component shall contain less than 30.0 micromoles of any one or any mixture of 3-butenyl glucosinolate, 4-pentenyl glucosinolate, 2-hydroxy-3-butenyl, or 2-hydroxy-4-pentenyl glucosinolate, per gram of air-dried, oil free solid (Canola Council of Canada).

6.1.1 Breeding methods

224. The amphidiploids, *B. napus*, *B. juncea* and *B. carinata*, are largely self-pollinating with the self-pollinated progeny exhibiting very little, if any, loss in vigour. Thus methods developed for highly inbred crops, such as the cereal grains, have been adapted for these partially outcrossing species. In the oilseed forms of these species complete homozygosity is normally not the objective,

although varietal distinctness, uniformity and stability are still a requirement. However, with cole crops and hybrids, high levels of homozygosity are required. Regardless of the breeding technique employed, success is dependent upon the identification of suitable parents that, when crossed, will yield progenies that express the desirable traits of both parents.

6.1.1.1 Mass selection

225. This early plant breeding technique relied on the identification and harvesting of seed from the most productive or desirable plants within a population for sowing in the following year. The system is one of population improvement based on plant phenotype and is best suited to self-fertilized crops and where gene action is additive. It lacks the efficiency of present day techniques but a variation is used today to preserve the identity of established varieties whereby off-types are removed from elite lines and breeder seed plots.

6.1.1.2 Pedigree method

226. In the past, most *B. napus* and *B. juncea* commercial varieties were developed using the pedigree method. Crosses are made between parents exhibiting the traits to be combined and the F₁s are selfed or intercrossed. The progeny are selfed or allowed to interpollinate and selection of the best F₄ rows is done within the best F₃ families. By the F₅₋₆ the vast majority of loci will be homozygous and the characteristics of the breeding line are fixed. The pedigree method may be modified in various ways depending on the inheritance of the trait or traits being introduced or combined. The method is well suited to the mainly self pollinating species *B. napus* and *B. juncea*, because the seed multiplication rate, unlike cereal grains, is high (ca. 1,000:1). In the self-incompatible *Brassica* vegetables and oilseed *B. rapa*, inbreeding leads to a rapid loss in vigour and reduced fertility. However, it is sometimes used to produce inbred lines destined for production of hybrid vegetables. Some cauliflower varieties are exceptions, being natural self-pollinators that do not exhibit the usual vigour and fertility losses.

6.1.1.3 Single seed descent

227. As with the pedigree method, the first step in single seed descent (SSD) breeding is the careful choice of parents for hybridisation. However, unlike the pedigree method, selection is not practiced until a high degree of homozygosity is reached. The object is to advance generations as rapidly as possible and subsequently select among the randomly derived lines. The size of the segregating population is kept at a manageable level by planting only one randomly chosen single seed from each plant in the previous generation. Since the degree of homozygosity is not as critical in *B. napus* and *B. juncea* as it is in cereals, this method has not been widely used in *Brassica* breeding programmes.

6.1.1.4 Backcross method

228. The backcross method is designed to introduce one or more specific trait(s) into an otherwise highly-desirable parent or variety. The donor parent, containing the trait(s) to be incorporated, is crossed onto plants of an adapted, desirable, recurrent parent. Depending on the inheritance of the trait(s) and the ease or efficiency of selection, the F₁ or selected BCF₁ plants will be backcrossed to the recurrent parent. By the 4th to 6th backcross, the genetic makeup of the recurrent parent is expected to have been reconstituted with the new trait incorporated. However, linkage between the desirable trait and one or more undesirable characteristics may require selection within large populations to identify plants or lines with an uncoupled linkage. Frequently, in the self-pollinating species, only one or two backcrosses are made followed by pedigree selection. Figure 44 illustrates the combined use of the backcross and pedigree methods.

229. In the self-incompatible species, backcrossing can also be effective for the incorporation of specific traits. However, crosses in oilseed *B. rapa* need to be made with sufficient numbers of recurrent parent plants to ensure heterozygosity of the self-incompatibility alleles of the recurrent parent is maintained in the backcross generations. To overcome this potential problem the “recurrent selection” breeding system is widely used. Backcrossing is also effective in the self-incompatible vegetable species. Dickson and Wallace (1986) outline a complete backcross breeding programme for cabbage improvement.

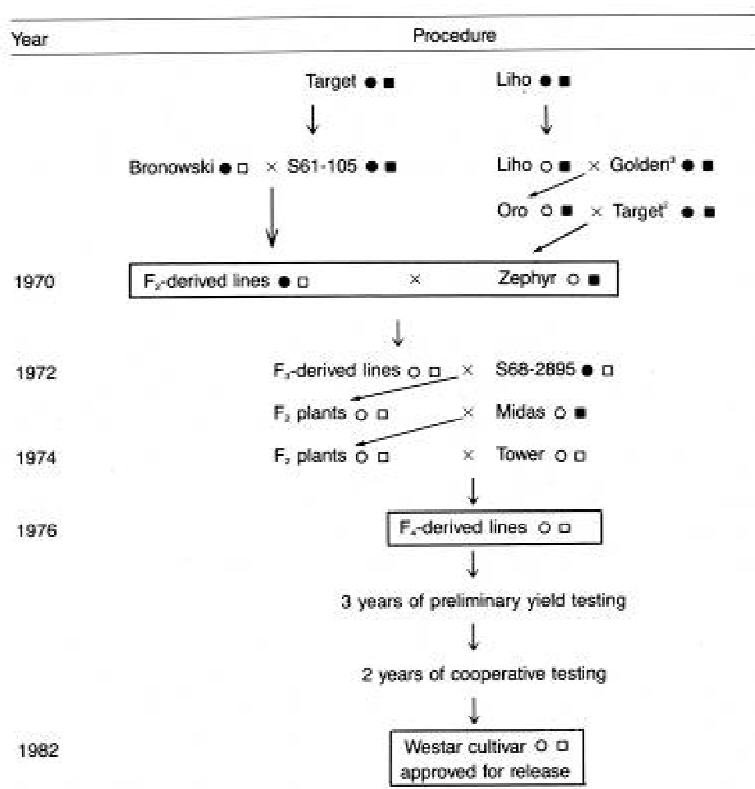


Figure 44. Breeding scheme combining the backcross and pedigree selection systems to develop a low erucic, low glucosinolate variety with high seed and oil yield

This breeding scheme uses agronomically superior parents that contribute either high (●) or low (○) erucic acid levels and high (■) or low (□) glucosinolate content. (Source: Downey and Rakow, 1987)

6.1.1.5 Recurrent selection method

230. This method is standard procedure for improving populations of self-incompatible species. Any type of segregating population may be a candidate for improvement. Normally open-pollinated seed is harvested at random from individual plants within the population, and a progeny row or rows sown from each plant. However, some seed from each plant is held in reserve. The progeny rows are evaluated and the best performing identified. An equal amount of the reserve seed from the best single plants, based on the performance of their progeny, is bulked. The first cycle of recurrent selection is complete when the new seed composite is sown in an isolation plot, and the second cycle begins with the harvesting of random single plants within the new composite. A bulk sample from the remaining plants can be harvested and planted in replicated trials to measure the response to selection. Recurrent selection is continued as long as it is anticipated that there will be a reasonable response to selection. With every additional trait under selection, the intensity of selection increases exponentially, thus it is difficult to improve a population for

several traits simultaneously. This constraint is overcome by having specialised composites for different traits that are brought together after the original objectives for each have been met.

6.1.1.6 Synthetic varieties

231. Allard (1960) defines a synthetic variety as one “that is maintained from open pollinated seed following its synthesis by hybridisation in all combinations among a number of selected genotypes”. This method, which is widely used in breeding forage crops, is also effective for the breeding of oilseed *B. rapa* (Falk *et al.*, 1994). Equal amounts of seed from varieties or recurrent lines that arise from widely different gene pools are mixed and sown in Syn.-0 isolation plots. Seed harvested from the Syn.-0 plot constitutes the Syn.-1 generation. Syn.-1 seed from a two component synthetic will consist of 25% from each parental genotype and 50% hybrid seed. Thus, if the parental lines are good combiners, a significant amount of heterosis can be captured. The method (Figure 45) has also been explored in *B. napus* (Becker *et al.*, 1999) but breeding programmes in this species are now directed to F₁ hybrid varieties. Normally, despite the high multiplication rate (1,000: 1), there is insufficient Syn.-1 seed for commercialisation so that Syn.-1 seed is sown to provide commercial Syn.-2 seed. This procedure has been used in Canada to produce the first commercial *B. rapa* synthetic varieties, Hysyn 100 and Hysyn 110. Because of the large number of genotypes within the parental lines there is very little loss in heterosis between the Syn.-1 and Syn.-2 generations (Falk and Woods, 2003). If the market is very large a Syn.-3 generation could be added.

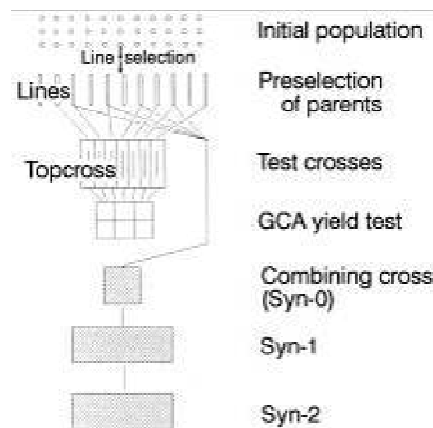


Figure 45. Breeding scheme for development of commercial synthetic varieties of oilseed *Brassica* crops (Source: Becker *et al.*, 1999)

6.1.1.7 Diallel and polycross methods

232. In vegetable crops uniform maturity, head size and appearance are critical to the success of a variety and seed yield is of secondary importance. Further, the numbers of parents that make up a variety are few and the market price of seed is substantially greater than the commodity oilseed crops. Thus breeding methods used for vegetables can be more intensive than the large population breeding methods used in oilseed improvement programmes. For example if a deleterious trait is controlled by a recessive gene, it is difficult to completely eliminate it from a self-incompatible plant population. However, within a small population of potential elite parents, diallel crossing, *i.e.* hand crossing each parent with all other potential parents, followed by progeny assessment, can eliminate the heterozygous parent(s). Although labour-intensive, this technique is suitable for most vegetable Brassicas because individual plants (parents) can be vegetatively maintained over many generations. Vegetative propagation also makes possible the use of the polycross breeding method used to identify desirable parents with good general combining ability. In this method, parental clones are space planted in a field design that assures

each parent is equally exposed to pollen from all the other parents in the nursery. Progeny evaluation then identifies the best parents for inter-pollination to produce seed of a new variety.

6.1.1.8 Hybrid varieties

233. The vigour, yield and uniformity advantages associated with hybrids in both oilseed and vegetable *Brassica* crops have been demonstrated by many breeders. The main constraint to their commercial exploitation has been an effective pollen control-fertility restoration system. Vegetable breeders have utilized the variations in SI alleles, which control the self-incompatible system, to produce single and double cross hybrids. Kuckuck (1979) illustrates how lines, selected for general combining ability and specific S alleles, are programmed to produce double cross cabbage hybrids (Figure 46). The self-incompatible parent can be maintained through bud pollination, micro propagation or by overcoming the SI barrier by exposing flowering plants to high CO₂ concentrations. Nuclear male sterility in oilseed rape has also been used commercially in China but the segregating male fertile progeny have to be removed by hand (Fu *et al.*, 1997), thus making the system expensive in many regions.

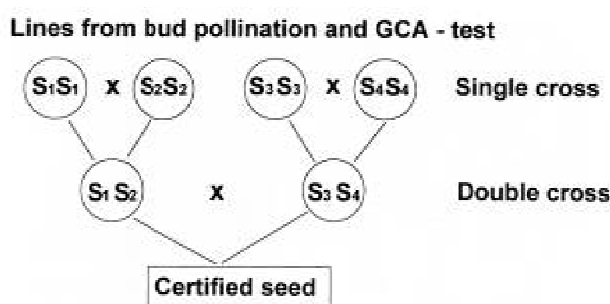


Figure 46. Self-incompatibility (SI) scheme for breeding cabbage hybrid seed production

Source: Kuckuck, 1979 modified by Becker *et al.*, 1999

234. The most practical and efficient system is that of cytoplasmic male sterility (CMS). More than 17 different male sterile forms have been investigated in *Brassica* species (Stiewe *et al.*, 1995; Prakash *et al.*, 1995). Only a few have been developed to the commercial stage, but varietal development programmes worldwide are rapidly moving to the use of CMS-restorer systems for hybrid seed production. The CMS systems are based on genetic miscommunication between cytoplasmic mitochondria and nuclear genes, resulting in the disruption of normal anther and/or pollen development. There are three components to the system: the *A line*, carrying the cytoplasmic mitochondrial genome that results in male sterility, the *B line* that is fully fertile and maintains the *A line*, and the *R line* with a nuclear gene that restores fertility. The *R line* should be highly heterotic to the *A line* to produce a high yielding, fully fertile F₁ commercial crop (Figure 47). China developed the first *B. napus* commercial CMS system, known as the Polima system (Fu *et al.*, 1997). However, the Polima system is rapidly being replaced in western breeding programmes with the *ogu*-INRA system, the Male Sterile Lembke (MSL) system and others under development (Stiewe *et al.*, 1995; Prakash *et al.*, 1995; Downey and Rimmer, 1993). A transgenic pollen control-restorer system, developed by Plant Genetic Systems and commercialised by Bayer CropScience, is in widespread use in Canada and the USA. Details of how this system functions are outlined by Downey and Rimmer (1993).

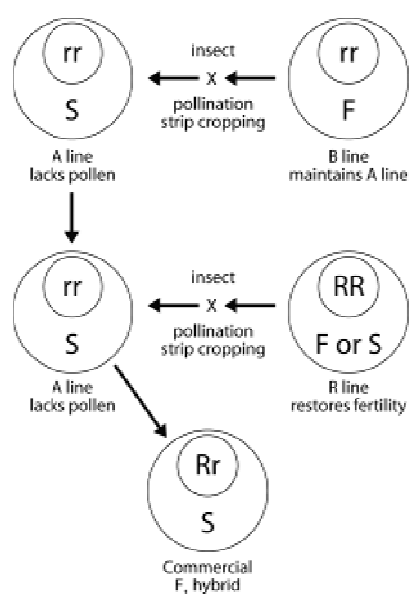


Figure 47. Production system for Cytoplasmic Male Sterile (CMS) hybrid seed of oilseed rape

The small circle represents the nucleus showing the fertility restorer genes r and R and the larger circle the cell with cytoplasm containing fertile (F) or sterile (S) mitochondrial genes.

6.2 Improvement through "Interspecific Hybrids" and "Cybrids"

235. Interspecific and intergenomic crosses are important options for the introduction of desired traits that are not available, or cannot be found, within the primary gene pool of a crop species. Normally such crosses are difficult to make. As noted previously, there are many natural barriers, both pre- and post-fertilization, that protect the integrity of a species. Further, even if such crossing is successful, chromosome pairing and alien gene introgression into the genome of the target species must occur. However, in the Brassicaceae a number of desirable nuclear genes from different genera and species have been transferred to targeted crop species. A list of traits that have been transferred to *B. napus*, *B. juncea* and/or *B. oleracea* from other Brassicaceae species is presented in Table 16 (Prakash *et al.*, 2009).

236. The development of protoplast fusion technology has been highly successful in circumventing the natural sexual barriers that separate the Brassicaceae species and genera. The technology has the potential to access desirable genes present in distant relatives (Glimelius, 1999; Christey, 2004; Navrátilová, 2004; Liu *et al.*, 2005). Prakash *et al.* (2009) has compiled a list of intertribal somatic hybrids in the Brassicaceae and the desirable traits to be transferred (Table 17). Additional intergenomic hybrids have been produced but failed to establish in soil *e.g.* *Camelina sativa* + *B. carinata* (Narasimhulu *et al.*, 1994), *C. sativa* + *B. oleracea* (Hansen, 1998) and *Barbarea vulgaris* + *B. napus* (Fahleson *et al.*, 1994b). With some exceptions, the somatic hybrids so far obtained have exhibited a high degree of sterility and/or morphological abnormalities that have limited their use. However, the importance of somatic hybridisation is not so much the direct use of the resulting amphidiploids, containing both parental genomes, but rather to utilize the somatic hybrids as a bridge to transfer desirable traits to target species (Glimelius, 1999).

237. Cell fusion not only brings together the nuclear contents of both parents but also combines the cytoplasm and organellar content of fused cells. Frequently, to improve the outcome, the nucleus of one parent is eliminated by X-ray, centrifugation or chemical treatment before fusion but the fused cell contains the cytoplasm of both parents. The resulting plant is termed a 'cybrid'. This technique allows cytoplasmic substitution which frequently results in cytoplasmic male sterility (CMS). Cell fusion among the Brassicaceae, where the cytoplasm of both parents are combined, can also generate novel cytoplasmic variability, bringing about organellar reassortment and DNA rearrangement, which is not possible using

sexual hybridisation. Chloroplast segregation is independent of mitochondrial segregation and while mitochondrial recombination has been frequently observed in the Brassiceae (Glimelius, 1999), recombination is rarely found in the chloroplasts. It is also rare to have a mixture of the two chloroplasts occurring in the same hybrid. In general, the chloroplasts are usually contributed by crop species. This may occur because many of the fusions are with the allopolyploid crop species that contribute large numbers of chloroplasts per cell (Butterfass, 1989).

Table 16. Introgression of nuclear genes conferring desirable traits to Brassica crops (Source: Prakash *et al.*, 2009)

Trait	Donor species	Recipient species	Reference
Yellow seed coat	<i>B. rapa</i>	<i>B. napus</i>	Chen and Heneen 1991
	<i>B. juncea</i> / <i>B. carinata</i>	<i>B. napus</i>	Rashid <i>et al.</i> 1994
	<i>B. carinata</i>	<i>B. napus</i>	Qi <i>et al.</i> 1995
	<i>B. rapa</i> / <i>B. carinata</i>	<i>B. napus</i>	Meng <i>et al.</i> 1998
	<i>B. rapa</i> / <i>B. juncea</i>	<i>B. napus</i>	Rahman 2001; Potapov and Osipova 2003
CMS fertility Restoration	<i>Raphanus sativus</i>	<i>B. napus</i> CMS (<i>Ogu</i>)	Heyn 1977; Rousselle <i>et al.</i> 1985
	<i>Raphanus sativus</i>	<i>B. napus</i> CMS (<i>Kosena</i>)	Sakai <i>et al.</i> 1996
	<i>B. juncea</i>	<i>B. napus</i> CMS (<i>Polima</i>)	Fan <i>et al.</i> 1985
	<i>B. tournefortii</i>	<i>B. napus</i> CMS (<i>Tour</i>)	Stiewe and Röbbelen 1994
	<i>Trachystoma ballii</i>	<i>B. juncea</i> CMS (<i>Trachy</i>)	Kirti <i>et al.</i> 1997
	<i>Moricandia arvensis</i>	<i>B. juncea</i> CMS (<i>Moricandia</i>)	Prakash <i>et al.</i> 1998
	<i>Erucastrum canariense</i>	<i>B. juncea</i> CMS (<i>Canariense</i>)	Prakash <i>et al.</i> 2001
		<i>B. napus</i> CMS (<i>Canariense</i>)	Banga <i>et al.</i> 2003b
<i>Enarthrocarpus lyratus</i>	<i>B. rapa</i> CMS (<i>Lyratus</i>)	Deol <i>et al.</i> 2003	
	<i>B. juncea</i> / <i>B. napus</i> CMS (<i>Lyratus</i>)	Banga <i>et al.</i> 2003a	
Chlorosis removal	<i>Raphanus sativus</i>	<i>B. napus</i> CMS (<i>Ogu</i>)	Paulmann and Röbbelen 1988
Beet cyst nematode Resistance	<i>Sinapis alba</i>	<i>B. napus</i>	Lelivelt <i>et al.</i> 1993
	<i>Raphanus sativus</i>	<i>B. napus</i>	Lelivelt and Krens 1992; Voss <i>et al.</i> 2000; Peterka <i>et al.</i> 2004; Budahn <i>et al.</i> 2006
Club root resistance	<i>B. napus</i>	<i>B. oleracea</i> var. <i>capitata</i>	Chiang <i>et al.</i> 1977,1980
	<i>B. rapa</i>	<i>B. napus</i>	Gowers 1982
Blackleg resistance	<i>B. juncea</i>	<i>B. napus</i>	Roy 1984; Dixelius 1999; Sacristán and Gerdemann-Knörck 1986
	<i>B. nigra</i>	<i>B. napus</i>	Struss <i>et al.</i> 1996; Chèvre <i>et al.</i> 1996, 1997b; Plieske <i>et al.</i> 1998; Dixelius 1999
	<i>Arabidopsis thaliana</i>	<i>B. napus</i>	Bohman <i>et al.</i> 2002; Ogbonnaya <i>et al.</i> 2003; Saal <i>et al.</i> 2004
	<i>Sinapis arvensis</i>	<i>B. napus</i>	Snowdon <i>et al.</i> 2000; Winter <i>et al.</i> 2003
	<i>Coincya monensis</i>	<i>B. napus</i>	Winter <i>et al.</i> 2003
<i>B. rapa</i>	<i>B. napus</i>	Chèvre <i>et al.</i> 2003	
Alternaria leaf spot Resistance	<i>Sinapis alba</i>	<i>B. napus</i>	Primard <i>et al.</i> 1988
	<i>Diplotaxis eruroides</i>	<i>B. napus</i>	Klewer <i>et al.</i> 2003
	<i>Sinapis alba</i>	<i>B. oleracea</i>	Sigareva <i>et al.</i> 1999
Black rot resistance	<i>B. juncea</i>	<i>B. napus</i>	Tonguc and Griffiths 2004
Soft rot resistance	<i>B. oleracea</i> var. <i>italica</i>	<i>B. oleracea</i> ; <i>B. rapa</i> subsp. <i>pekinensis</i>	Ren <i>et al.</i> , 2001a,b
Altered oil quality	<i>Orychophragmus violaceus</i>	<i>B. napus</i>	Hu <i>et al.</i> 2002; Hua and Li 2006
Earliness	<i>B. rapa</i>	<i>B. napus</i>	Shiga 1970; Namai <i>et al.</i> 1980
	<i>B. oleracea</i>	<i>B. napus</i>	Habman <i>et al.</i> 2010
Low erucic acid	<i>B. juncea</i>	<i>B. carinata</i>	Getinet <i>et al.</i> 1994
Low glucosinolate	<i>B. napus</i>	<i>B. carinata</i>	Getinet <i>et al.</i> 1994

Table 17. Intertribal somatic hybrids in Brassiceae for the integration and incorporation of desirable traits into *Brassica* crops
(Source: Prakash *et al.*, 2009)

Somatic hybrid	Desirable trait for introgression	Reference
<i>Arabidopsis thaliana</i> (n=5) + <i>B. nigra</i>	Resistance to flea beetles, cold tolerance, short life cycle	Siemens and Sacristán 1995*
<i>Arabidopsis thaliana</i> (n=5) + <i>B. oleracea</i>	Plastome transformation	Nitovskaya and Shakhovskiy 1998; Yamagishi and Nakagawa 2004; Nitovskaya <i>et al.</i> 2006a
<i>Arabidopsis thaliana</i> (n=5) + <i>B. rapa</i>	Experimental demonstration	Gleba and Hoffmann 1979, 1980
<i>Arabidopsis thaliana</i> (n=5) + <i>B. juncea</i>	Phosphinothricin resistance	Ovcharenko <i>et al.</i> 2004
<i>Arabidopsis thaliana</i> (n=5) + <i>B. napus</i>	Herbicide resistance, Blackleg resistance	Bauer-Weston <i>et al.</i> 1993*; Forsberg <i>et al.</i> 1994, 1998*; Yamagishi <i>et al.</i> 2002*
	Transposable element <i>Spm/dSpm</i>	Ovcharenko <i>et al.</i> 2005*
<i>Armoracia rusticana</i> (n=16) + <i>B. oleracea</i>	Clubroot resistance	Navrátilová <i>et al.</i> 1997
<i>Barbarea vulgaris</i> (n=8) + <i>B. oleracea</i>	Cold tolerance	Ryschka <i>et al.</i> 1999
<i>Barbarea vulgaris</i> (n=8) + <i>B. rapa</i>	Cold tolerance	Oikarinen and Ryöppy 1992
<i>Barbarea vulgaris</i> (n=8) + <i>B. napus</i>	Cold tolerance	Fahleson <i>et al.</i> 1994b
<i>Barbarea stricta</i> (n=8) + <i>B. rapa</i>	Cold tolerance	Oikarinen and Ryöppy 1992
<i>Camelina sativa</i> (n=20) + <i>B. oleracea</i>	Alternaria resistance	Hansen 1998; Sigareva and Earle 1999
<i>Camelina sativa</i> (n=20) + <i>B. carinata</i>	Alternaria resistance	Narasimhulu <i>et al.</i> 1994
<i>Capsella bursa-pastoris</i> (n=16) + <i>B. oleracea</i>	Resistance to flea beetles, alternaria blight	Nitovskaya <i>et al.</i> 1998; Sigareva and Earle 1999
<i>Crambe abyssinica</i> (n=45) + <i>B. napus</i>	High erucic acid content, insect resistance	Wang <i>et al.</i> 2003*, 2004a*
<i>Lepidium meyenii</i> (n=32) + <i>B. oleracea</i>	Glucosinolate content	Ryschka <i>et al.</i> 2003
<i>Lesquerella fendleri</i> (n=6) + <i>B. napus</i>	High lesquerolic acid content, drought tolerance	Skarzhinskaya <i>et al.</i> 1996**, 1998;
	<i>Lesquerella</i> chloroplasts	Schröder-Pontoppidan <i>et al.</i> 1999; Nitovskaya <i>et al.</i> 2006b
<i>Lunaria annua</i> (n=14) + <i>B. napus</i>	High nervonic acid content	Craig and Millam 1995
<i>Matthiola incana</i> (n=7) + <i>B. oleracea</i>	Oil quality	Ryschka <i>et al.</i> 1999
<i>Orychophragmus violaceus</i> (n=12) + <i>B. napus</i>	High linoleic and palmitic acid content	Hu <i>et al.</i> 1999; Hu <i>et al.</i> 2002*
	Phosphinothrin resistance	Sakhno <i>et al.</i> 2007
	Chlorosis correction	Vasilenko <i>et al.</i> 2003
<i>Thlaspi perfoliatum</i> (n=21) + <i>B. napus</i>	High nervonic acid content	Fahleson <i>et al.</i> 1994a**
<i>Thlaspi caerulescens</i> (n=7) + <i>B. napus</i>	Zinc and cadmium tolerance	Brewer <i>et al.</i> 1999
<i>Thlaspi caerulescens</i> (n=7) + <i>B. juncea</i>	High metal accumulation	Dushenkov <i>et al.</i> 2002

*Denotes asymmetric hybrids and ** both asymmetric and symmetric hybrids identified.

6.3 Biotechnology in *Brassica* Breeding

6.3.1 Introduction

238. Although the above breeding procedures have been very effective in combining important agronomic and nutritional traits in superior cultivars, the process of identifying the desired genotype in genetically stable, uniform and high yielding varieties takes many years. Further, the small chromosome size plus their lack of distinctive features has been an additional limitation on the selection of superior genotypes. However, beginning in the mid to late 1980's developments in tissue culture, embryo rescue, cell fusion, molecular markers and genetic mapping have not only reduced the time from cross to market but have given breeders powerful tools to quickly identify and assemble desirable traits in a single genotype. In addition, these biotech tools have greatly expanded the size and variation of the available gene pool, well beyond species boundaries.

6.3.2 Doubled Haploid breeding

239. The Doubled Haploid (DH) breeding technique is now widely used in *B. napus* and *B. juncea* breeding programmes (Ferrie and Keller, 2004). This breeding tool not only eliminates the several generations needed to attain genetic stability and uniformity in breeding lines, but also significantly reduces the size of populations needed to find a desired genotype. For example in *B. napus* two genes code for the level of the fatty acid erucic in the seed oil, and an additional six genes code for the content of glucosinolates in the seed. Thus when making a high by low cross, to produce progeny that have both low erucic acid and low glucosinolate (double low or canola quality), large segregating populations must be examined since the desired genotype must have all eight genes in the recessive state. Table 18 illustrates the DH technique's increased selection efficiency, particularly when the selected plants are completely homozygous individuals that can be used directly as pure breeding varieties or as hybrid parents.

Table 18. Minimum population size required to select the least frequent homozygote at 95% probability

No. of genes	Minimum F ₂ population	
	Diploid	Haploid
1	11	5
2	47	11
4	766	47
5	3,067	95
6	12,269	191
7	49,077	382
8	196,259	766
10	3,123,923	3,067

Source: Rajhathy, 1976

240. The technique involves inducing large numbers of immature pollen grains (microspores) from *Brassica* species to develop into plants with the gametic or half the somatic chromosome number. Such plants are termed haploids and are sterile. By applying colchicine to the developing haploid plant, cell division is temporarily arrested bringing about chromosome duplication. The result is a doubled haploid (DH) or dihaploid plant that is fully fertile and totally homozygous. Thus complete homozygosity is reached in a single generation, and all seeds arising from self fertilization of that plant will be genetically identical. It is this single step to homozygosity that reduces the number of generations and time required to develop a new variety or hybrid parent. However, in a breeding programme, large populations of DH lines must be generated and evaluated since no prior selection has taken place. DH lines are usually derived

from F₁ donors, although the use of F₂ and F₃ donor plants allows for more recombination and some preselection.

6.3.3 *Molecular markers and their application*

241. Marker assisted selection and chromosome mapping came into general use in the 1980s with the development of RFLP (Restriction Fragment Length Polymorphisms) techniques that resulted in the first linkage maps for *B. oleracea* (Slocum *et al.*, 1990), *B. rapa* (Song *et al.*, 1991) and *B. napus* (Landry *et al.*, 1991). This technique was important in identifying genomes and their chromosomes, locating genes and Qualitative Trait Loci (QTLs), which are DNA regions containing a gene or genes that regulate traits of agronomic or quality interest. The discovery of the Polymerase Chain Reaction (PCR) by Mullis and Faloona (1987) resulted in new types of genetic markers such as Amplified Fragment Length Polymorphisms (AFLPs) that are more sensitive than RFLPs and simultaneously detect various polymorphisms in different genomic regions. Additional marker systems have since been added to the tool box including: Random Amplified Polymorphic DNAs (RAPDs); Sequence Tagged Sites (STS); Simple Sequence Repeats (SSRs) or microsatellites and Single Nucleotide Polymorphisms (SNPs). Breeders use these molecular markers to produce densely marked chromosome maps that can then be used to: 1) characterise germplasm and its genetic variability, 2) estimate the genetic distance between gene pools, inbreds and populations, 3) detect and locate QTLs and monogenic traits of interest, 4) select genotypes based on the presence or absence of specific markers and 5) identify useful candidate genes for sequencing (For more detailed information on genome mapping and molecular breeding in *B. napus*, see Snowdon *et al.* 2007a, b). The marker systems differ in their ease of use, cost and other characteristics. It is expected that the SNPs system will become the marker system of preference, despite its initial high cost, due to its ease of use, low cost per analysis, and high level of reproducibility (Korzun, 2003).

6.3.4 *Comparative genomic gene identification*

242. The distantly-related and intensively-studied species *Arabidopsis thaliana* provides information that is highly relevant for gene isolation and characterization in *Brassica* crops. However, the genomes of *Brassica* species are much more complex (Snowdon *et al.*, 2007b). A comprehensive comparative RFLP linkage map of *A. thaliana* and *B. napus* genomes indicated the five *Arabidopsis* chromosomes could be allocated to a minimum of 22 conserved, duplicated and rearranged blocks throughout the *B. napus* genome (Parkin *et al.*, 2005). Such information highlights the complexity of genome rearrangements between the two species, but also the great potential the model genome offers for comparative genetic analysis of the *Brassica* crops (Snowdon *et al.*, 2007b).

6.3.5 *TILLING technique*

243. The technique of TILLING (Targeted Induced Local Lesions in Genomes) can be used to identify a series of mutations (alleles) in a target gene by heteroduplex analysis (McCallum *et al.*, 2000). The method combines a standard technique of mutagenesis with a chemical mutagen such as ethyl methanesulfonate (EMS)², with a sensitive DNA screening-technique that identifies single base mutations (also called point mutations) in a target gene. This technique is available from the Canadian TILLING Initiative (CAN-TILL) at the University of British Columbia on a fee-for-service basis. The CAN-TILL facility is currently developing a large scale mutant population for *Brassica napus* as part of a Genome Canada project and has completed projects on *B. oleracea* and *Arabidopsis thaliana*. *B. rapa* TILLING services are available from RevGenUK in the United Kingdom (John Innes Centre).

² ethyl methanesulfonate (EMS): http://en.wikipedia.org/wiki/Ethyl_methanesulfonate

6.3.6 *Gene transfer*

244. The transfer of a gene(s) from an unrelated species is undertaken only when the desired trait cannot be found or induced by traditional methods. Because of the huge costs and time required to comply with multiple regulations in multiple countries, only those traits that have a potentially large and valuable market are considered for commercial exploitation.

REFERENCES

- AAFC (1994), "Agriculture and Agri-Food Canada – Regulatory Directive DIR 94-09: The Biology of *Brassica napus* L. (Canola/ Rapeseed)", pp. 11.
- AAFC (1995a), "Agriculture and Agri-Food Canada – Decision Document DD95-01: Determination of Environmental Safety of AgrEvo Canada Inc.'s Glufosinate-ammonium-tolerant Canola", pp. 7.
- AAFC (1995b), "Agriculture and Agri-Food Canada – Decision Document DD95-02: Determination of Environmental Safety of Monsanto Canada Inc.'s Roundup® Herbicide-tolerant *Brassica napus* Canola Line GT 73", pp. 6.
- AAFC (1995c), "Agriculture and Agri-Food Canada – Decision Document DD95-03: Determination of Environmental Safety of Pioneer Hi-Bred International Inc.'s Imidazolinone-tolerant Canola", pp. 6.
- AAFC (1995d), "Agriculture and Agri-Food Canada – Decision Document DD95-04: Determination of Environmental Safety of Plant Genetic Systems Inc (PGS) Novel Hybridization System for Canola (*Brassica napus* L.)", pp. 8.
- AAFC (1996a), "Agriculture and Agri-Food Canada – Decision Document DD96-07: Determination of Environmental Safety of Monsanto Canada Inc.'s Roundup® Herbicide-tolerant *Brassica napus* Line GT200", pp. 6.
- AAFC (1996b), "Agriculture and Agri-Food Canada – Decision Document DD96-011: Determination of Environmental Safety of AgrEvo Canada Inc.'s Glufosinate-ammonium-tolerant Canola Line HCN28", pp. 5.
- Abel, S. and H.C. Becker (2007), "The Effect of Autopolyploidy on Biomass Production in Homozygous Lines of *Brassica rapa* and *Brassica oleracea*", *Plant Breed*, 126, pp. 642-643.
- Adler, L. S., *et al.* (1993), "Potential for Persistence of Genes Escaped from Canola: Germination Cues in Crop, Wild, and Crop-wild Hybrid *Brassica rapa*", *Functional Ecology*, 7, pp. 736-745.
- Agrisearch (2001), *A Physical Survey of Representative Australian Roadside Vegetation to Evaluate the Incidence and Distribution of Canola and Key Brassicaceae Weeds*, pp. 54.
- Alam, M., *et al.* (1992), "Cross Compatibility Studies within the Genus *Brassica* 1. Amphidiploid Combinations", *Sci. Khyber*, 5, pp. 89-92.
- Alam, Z. (1945), "Nomenclature of Oleiferous Brassicas Cultivated in Punjab", *Indian J. Agr. Sci.*, 15, pp. 173-181.
- Al Mouemar, A. and H. Darmency (2004), "Lack of Stable Inheritance of Introgressed Transgene from Oilseed Rape in Wild Radish", *Environ. Biosafety Res.*, 3, pp. 209-214.
- Al-Shehbaz, L.A. (1984), "The Tribes of Cruciferae (Brassicaceae) in the Southeastern United States", *J. Arnold Arber.*, 65, pp. 343-373.
- Al-Shehbaz, L.A., *et al.* (2006), "Systematics and Phylogeny of the Brassicaceae (Cruciferae): an Overview", *Pl. Syst. Evol.*, 259, pp. 89-130.
- Allainguillaume, J., *et al.* (2006), "Fitness of Hybrids between Rapeseed (*Brassica napus*) and Wild *Brassica rapa* in Natural Habitats", *Mol. Ecol.*, 15, pp. 1175-1184.
- Allard, R.W. (1960), *Principles of Plant Breeding*, J. Wiley, New York, pp. 485.
- Allender, C.J. and G.J. King, (2010), "Origins of the Amphiploid Species *Brassica napus* L. Investigated by Chloroplast and Nuclear Molecular Markers", *BMC Plant Biology*, 10, pp. 54.

- Almond, J.A., *et al.* (1986), "Aspects of Crop Husbandry", in R.D.H. Scarisbrick and R.W. Daniels (eds.), *Oilseed Rape*, Collins, London, pp. 127-175.
- Ammitzbøll, H. and R.B. Jørgensen (2006), "Hybridization between Oilseed Rape (*Brassica napus*) and Different Populations and Species of *Raphanus*", *Environ. Biosafety Res.*, 5, pp. 3-13.
- Andersson, M.S. and M.C. De Vicente (2010), "Canola, Oilseed Rape", in M.S. Andersson and M.C. de Vicente (eds.), *Gene Flow between Crops and their Wild Relatives*, John Hopkins Univ. Press, pp. 73-123.
- Aono, M., *et al.* (2006), "Detection of Feral Transgenic Oilseed Rape with Multiple-herbicide Resistance", *Japan Environ. Biosafety Res.*, 5, pp. 77-87.
- AOSCA (Association of Official Seed Certifying Agencies), "Crop Certification", www.aosca.org/crop%20certification.htm, accessed 27 Oct 2011.
- Appelqvist, L.-Å., and R. Ohlson (1972), *Rapeseed: Cultivation, Composition, Processing and Utilization*, Elsevier, Amsterdam and New York, pp. 393.
- APS (American Phytopathological Society), (2001), "Diseases of Rapeseed = Canola (*B. napus* L. and *Brassica rapa* L. (= *B. campestris* L.))", www.apsnet.org/publications/commonnames/Pages/Rapeseed.aspx, accessed 27 Oct 2011.
- Armstrong, S.J., *et al.* (1998), "Physical Mapping of DNA Repetitive Sequences to Mitotic and Meiotic Chromosomes of *Brassica oleracea* var. *alboglabra* by Fluorescence *in situ* Hybridization", *Heredity*, 81, pp. 666-673.
- Australian Bureau of Statistics, *Agricultural Census 2001*
- Axelsson, T., *et al.* (2000), "Amphidiploid *Brassica juncea* Contains Conserved Progenitor Genomes", *Genome*, 43, pp. 679-688.
- Ayotte, R., *et al.* (1987), "The Transfer of Triazine Resistance from *Brassica napus* L. to *B. oleracea* L. I. Production of F₁ Hybrids through Embryo Rescue", *Euphytica*, 36, pp. 615-624.
- Bailey, L.H. (1930), "The Cultivated Brassicas: Second Paper", *Gentes Herb.*, 2, pp. 211-267.
- Baker, J and C. Preston, (2008), "Canola (*Brassica napus* L.) Seedbank Declines Rapidly in Farm Managed Fields in South Australia", *Australian J. Agric. Res.*, 59, pp. 780-784.
- Banga, S.S., *et al.* (2003a), "Alloplasmic Male Sterile *Brassica juncea* with *Enarthrocarpus lyratus* Cytoplasm and the Introgression of Gene(s) for Fertility Restoration from Cytoplasmic Donor Species", *Theor. Appl. Genet.*, 106, pp. 1390-1395.
- Banga, S.S., *et al.* (2003b), "Alloplasmic Line of *Brassica napus* L. with *Erucastrum canariense* Cytoplasm is Male Sterile", *Proc. 11th Int. Rapeseed Congr.*, Copenhagen, Denmark, 6-10 July, pp. 324-325.
- Baranger, A., *et al.* (1995), "Effect of Oilseed Rape Genotype on the Spontaneous Hybridization Rate with a Weedy Species: an Assessment of Transgene Dispersal", *Theor. Appl. Genet.*, 91, pp. 956-963.
- Bateman, A.J. (1974a), "Contamination in Seed Crops I. Insect Pollination", *J. Genet.*, 48, pp. 257-275.
- Bateman, A.J. (1974b), "Contamination in Seed Crops III. Relation with Isolation Distance", *Heredity*, 1, pp. 303-336.
- Batra, V., *et al.* (1989), "Hybrids of Wild Species *Erucastrum gallicum* and Crop brassicas", *Proc. 6th Int. Congr. SABRAO*, 1: pp. 443-446.
- Bauer-Weston, W.B., *et al.* (1993), "Production and Characterization of Asymmetric Somatic Hybrids between *Arabidopsis thaliana* and *Brassica napus*", *Theor. Appl. Genet.*, 86, pp. 150-158.
- Bayer CropScience (2009), "Bayer CropScience First to Sequence the Entire Genome of Rapeseed/Canola", www.bayercropscience.ca/our-company/news/Bayer-CropScience-first-to-sequence-the-entire-genome-of-rapeseed-canola/, accessed 12 Oct 2009.

- Becker, H.C., *et al.* (1991), "Genotypic and Environmental Variation for Outcrossing Rate in Rapeseed (*Brassica napus*)", in D.I.McGregor (ed.), *Proc. 8th Int. Rapeseed Congr.*, I., Saskatoon, Canada, 5, pp. 1455-1459.
- Becker, H.C., *et al.* (1992), "Environmental Variation for Outcrossing Rate in Rapeseed (*Brassica napus*)", *Theor. Appl. Genet.*, 84, pp. 303-306.
- Becker, H.C., *et al.* (1999), "Breeding: An Overview", in C. Gómez-Campo(ed.), *Biology of Brassica Coenospecies*, Elsevier Science, pp. 413-460.
- Beckie, H.J. and M.D.K. Owen, (2007), "Herbicide-resistant Crops as Weeds in North America", *CAB Reviews: Persp. Agric. Veter. Sci. Nutr. Natl. Resour.*, 2, No.044, pp. 22.
- Beckie, H.J. and S.I. Warwick (2010), "Persistence of an Oilseed Rape Transgene in the Environment", *Crop Prot.*, 29(5), pp. 509-512.
- Beckie, H.J., *et al.* (2001), "Impact of Herbicide-resistant Crops as Weeds in Canada", *Proc. Brighton Crop Protection Conf. Weeds*, British Crop Protection Council (BCPC), pp. 135-142.
- Beckie, H.J., *et al.* (2003), "Gene Flow in Commercial Fields of Herbicide-resistant Canola (*Brassica napus*)", *Ecol. Appl.*, 13, pp. 1276-1294.
- Beckie, H.J., *et al.* (2004), "Multiple Herbicide-resistant Canola can be Controlled by Alternative Herbicides", *Weeds Sci.*, 52, pp. 152-157.
- Beckie, H.J., *et al.* (2006), "A Decade of Herbicide-resistant Crops in Canada", *Can. J. Plant Sci.*, 86, pp. 1243-1264.
- Beismann, H. and A. Roller (2003), "Assessing the Number of Transgenic Oilseed Rape Seeds in the Soil Seedbank of Former Release Sites", *Aspects of Applied Biology*, 69, pp. 209-215.
- Bell, J.M. (1993), "Factors Affecting the Nutritional Value of Canola Meal", *A review. Can. J. Anim. Sci.*, 73, pp. 679-697.
- Bell, D.T. and C.H. Muller, (1973), "Dominance of California Annual Grasslands by *Brassica nigra*", *Am. Midl. Nat.*, 90, pp. 277-299.
- Benabdelmouna, A., *et al.* (2003), "Genome Discrimination in Progeny of Interspecific Hybrids between *Brassica napus* and *Raphanus raphanistrum*", *Genome*, 46, pp. 469-472.
- Bengtsson, L., *et al.* (1972), "Botany of Rapeseed", in L.-Å Appelqvist and R. Olson (eds.), *Rapeseed: Cultivation, Composition, Processing and Utilization*, Elsevier, Amsterdam and New York, pp. 36-48.
- Bennett, M.D. and I.J. Leitch (2004), *Angiosperm DNA C-values database* (release 5.0, Dec. 2004), www.rbgekew.org.uk/cval/homepage.html.
- Beschorner, M., *et al.* (1995), "Analysis of Self-incompatibility Interactions in 30 Resynthesized *Brassica napus* Lines. I. Fluorescence Microscopic Studies", *Theor. Appl. Genet.*, 90, pp. 665-670.
- Bielikova, L. and S. Rakousky (2001), "Survey on Oil-seed Rape Cultivation and Weed Relatives in the Czech Republic", *European Sci. Foundation Meeting of a Working Group on: Interspecific Gene Flow from Oilseed Rape to Weedy Species*, June 2001, Rennes, France, pp. 9.
- Bijral, J.S. and T.R. Sharma (1996a), "Cytogenetics of Intergeneric Hybrids between *Brassica napus* L. and *Eruca sativa* Lam", *Cruciferae Newslett. Eucarpia*, 18, pp. 12-13.
- Bijral, J.S. and T.R. Sharma (1996b), "Intergeneric Hybridization between *Brassica napus* and *Diplotaxis muralis*", *Cruciferae Newslett. Eucarpia*, 18, pp. 10-11.
- Bijral, J.S. and T.R. Sharma (1998), "Production and Cytology of Intergeneric Hybridization between *Brassica napus* and *Diplotaxis catholica*", *Cruciferae Newslett. Eucarpia*, 20, pp. 15.
- Bijral, J. S., *et al.* (1993), "Morphocytogenetics of *Brassica napus* L. × *Sinapis alba* L. Sexual Hybrids", *Indian J. Genet. Pl. Breed.*, 53, pp. 442-444.
- Bijral, J.S., *et al.* (1995), "Interspecific Hybrids of *Brassica maurorum* with *Brassica* Crops, and their Cytology", *Cruciferae Newsl. Eucarpia*, 17, pp. 18-19.

- Bilsborrow, P.E., *et al.* (1998), "Contamination of Edible Double-low Oilseed Rape Cultivars via Pollen Transfer from High Erucic Cultivars", *J. Sci. Food Agric.*, 76, pp. 17-22.
- Bing, D.J., *et al.* (1991), "Potential of Gene Transfer among Oilseed *Brassica* and their Weedy Relatives", in D.I. McGregor (ed.), *Proc. 8th Int. Rapeseed Congr.*, Saskatoon, Canada, pp. 1022-1027.
- Bing, D.J., *et al.* (1996), "Assessment of Transgene Escape from *Brassica rapa* (*B. campestris*) into *B. nigra* or *Sinapis arvensis*", *Plant Breeding*, 115, pp. 1-4.
- Bohman, S., *et al.* (2002), "Arabidopsis thaliana-derived Resistance against *Leptosphaeria maculans* in a *Brassica napus* Genomic Background", *Theor. Appl. Genet.*, 105, pp. 498-504.
- Bonnemaison, L. (1965), "Insect Pests of Crucifers and their Control", *Ann. Rev. Entomol.*, 10, pp. 233-256.
- Booth, E.J., *et al.* (1996), "Assessment of the Ecological Consequences of Introducing Transgenic Rapeseed", *4th ESA Congress Book of Abstracts. (Persistence of Oil-modified Oilseed Rape, Sinapis arvensis and Brassica nigra)*, pp. 144-145.
- Bothmer, v. R., *et al.* (1995), "Brassica sect. Brassica (*Brassicaceae*) II. Inter- and Intraspecific Crosses with Cultivars of *B. oleracea*", *Gene Resource and Crop Evolution*, 42, pp. 165-179.
- Boulter, G.S. (1983), "The History and Marketing of Rapeseed Oil in Canada", in J.K. Kramer, *et al.*(eds.), *High and Low Erucic Acid Rapeseed Oils*, Academic Press, Toronto, pp. 62-83.
- Brewer, E.P., *et al.* (1999), "Somatic Hybridization between the Zinc Accumulator *Thlaspi caerulescens* and *Brassica napus*", *Theor. Appl. Genet.*, 99, pp. 761-771.
- Briggs, F.N. and P. F. Knowles (1967), *Introduction to Plant Breeding*, Reinhold Publishing. pp. 426.
- Brown, A.P., *et al.* (1996), "Gene Transfer between Canola (*Brassica napus*) and Related Weed Species", *Proc. 8th Symposium on Environmental Releases of Biotechnology Products: Risk Assessment Methods and Research Progress*, Ottawa, Canada.
- Brown, J. and A.P. Brown (1996), "Gene Transfer between Canola (*Brassica napus* L. and *B. campestris* L.) and Related Weed Species", *Annals of Appl. Biology*, 129, pp. 513-522.
- Brown, J., *et al.* (1997), "Intergeneric Hybridization between *Sinapis alba* and *Brassica napus*", *Euphytica*, 93, pp. 163-168.
- Brown, P.D. and M.J. Morra, (1997), "Control of Soil-born Plant Pests Using Glucosinolate-containing Plants", *Advan. Agron.*, 61, pp. 167-213.
- Budahn, H., *et al.* (2006), "Intergeneric Transfer of Nematode Resistance from *Raphanus* to *Brassica* using a Series of Rape-radish Chromosome Addition Lines", *Acta Hort*, pp. 706.
- Butterfass, T.H. (1989), "Nuclear Control of Plastid Division", in S.A. Boffey and D. Lloyd (eds.), *Division and Segregation of Organelles*, Soc. Expt. Biol., Seminar Series 25, pp. 21-38.
- Burkill, I.H. (1930), "The Chinese Mustards in the Malay Peninsula", *Gad' Bull.*, 5, pp. 99-117.
- Campbell, A.G. (1959), "A Germination Inhibitor and Root Growth Retarder in Chou Moellier (*Brassica oleracea* var.)", *Nature*, 183, pp. 1263-1264.
- Canadian Seed Growers Association (CSGA), "Canadian Regulations and Procedures for Breeder Seed Crop Production", www.seedgrowers.ca/pdfs/Breeder%20Seed%20Crop%20Regulations_ENGLISH_20090915.pdf, accessed 27 Oct 2011.
- Canola Council of Canada, "Canadian Canola Industry - Official Definition of Canola", www.canolacouncil.org/ind_definition.aspx, accessed 27 Oct 2011.
- CAN-TILL (The Canadian TILLING Initiative), "CAN-TILL", University of British Columbia, Vancouver, B.C., Canada, www.botany.ubc.ca/can-till/CAN-TILL.html, accessed 27 Oct 2011.
- Ceddia, G.C., *et al.* (2007), "Landscape Gene Flow, Coexistence and Threshold Effect: The Case of Genetically Modified Herbicide Tolerant Oilseed Rape (*Brassica napus*)", *Ecological Modelling*, 205, pp. 169-180.

- CETIOM (Centre technique interprofessionnel des oléagineux métropolitains) (2000), "Introduction of Genetically Modified Rapeseed Tolerant to Various Herbicides in the French Agriculture System: Evaluation of the Agro-environmental Impact and Development of Management Scenarios", *Summary of GMO 2000 report prepared in the framework of the Moratorium*, Ver. 4.2, pp. 29.
- CFIA (Canadian Food Inspection Agency) (2007), *Biology document BIO2007-01: The biology of Brassica juncea (Canola /Mustard)*, pp. 14.
- CFIA, (2009), "5.9 Legal Reference Samples", *Procedures for the Registration of Crop Varieties in Canada*, www.inspection.gc.ca/english/plaveg/variet/proced/regproe.shtml#aii59, accessed 27 Oct 2011.
- Chadoeuf, R., *et al.* (1998), "Survival of Buried Seeds of Interspecific Hybrids between Oilseed Rape, Hoary Mustard and Wild Radish", *Field Crop Res.*, 58, pp. 197-204.
- Champolivier, J., *et al.* (1999), "Management of Transgenic Crops within the Cropping System", in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 233-240.
- Chang, K.C. (1970), "The beginnings of agriculture in the Far East", *Antiquity*, Vol. 44: pp. 175-185.
- Chapman, G.P. and Y.W. Wang (2002), *The Plant Life of China: Diversity and Distribution*, Springer-Verlag, Berlin, pp. 257.
- Chen, B.Y., and W.K. Heneen (1991), "The Basic Number of *Brassica* Genomes: $x=3$?", *Cruciferae Newsl.*, 14-15, pp. 20-21.
- Chen, B.Y., *et al.* (1989), "Comparative and Genetic Studies of Isozymes in Resynthesized and Cultivated *Brassica napus* L., *B. campestris* L. and *B. alboglabra* Bailey", *Theor. Appl. Genet.*, 77, pp. 673-679.
- Chen, L., *et al.* (2005), "Production of Interspecific Somatic Hybrids between Tuber Mustard (*Brassica juncea*) and Red Cabbage (*Brassica oleracea*)", *Plant Cell Tissue Organ. Cult.*, 80, pp. 305-311.
- Chèvre, A.M., *et al.* (1994), "Comparison of Somatic and Sexual *B. napus*-*Sinapis alba* Hybrids and their Progeny by Cytogenetic and Molecular Characterization", *Genome.*, 37, pp. 367-374.
- Chèvre, A.M., *et al.* (1996), "Interspecific Gene Flow as a Component of Risk Assessment for Transgenic *Brassicas*", *Acta Hort.*, 407, pp. 169-179.
- Chèvre, A.M., *et al.* (1997a), "Gene Flow from Transgenic Crops", *Nature*, 389, pp. 924.
- Chèvre, A.M., *et al.* (1997b), "Selection of Stable *Brassica napus*-*B. juncea* Recombinant Lines Resistant to Black Leg (*Leptosphaeria maculans*). 1. Identification of Molecular Markers, Chromosomal and Genomic Origin of the Introgression", *Theor. Appl. Genet.*, 95, pp. 1104-1111.
- Chèvre, A.M., *et al.* (1998), "Characterisation of Backcross Generations Obtained under Field Conditions from Oilseed Rape-Wild Radish F1 Interspecific Hybrids: an Assessment of Transgene Dispersal", *Theor. Appl. Genet.*, 97, pp. 90-98.
- Chèvre, A.M., *et al.* (1999), "Gene Flow from Oilseed Rape to Weeds", in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 125-130.
- Chèvre, A.M., *et al.* (2000), "Assessment of Interspecific Hybridization between Transgenic Oilseed Rape and Wild Radish under Normal Agronomic Conditions", *Theor. Appl. Genet.*, 100, pp. 1233-1239.
- Chèvre, A.M., *et al.* (2001), "Impacts Agro-environnementaux de la Mise en Culture de Variétés de Colza Transgéniques Tolérantes à des Herbicides", *C. R. Acad. Agrc. Fr.*, 87 (5), pp. 11-20.
- Chèvre, A.M., *et al.* (2003), "Introduction of Black Leg Resistance from *Brassica rapa* into *B. napus*", *Proc. 11th Intern. Rapeseed Congr.*, Copenhagen, Denmark, 6-10 July, 1, pp. 32-35.
- Chèvre, A.M., *et al.* (2004), "A Review on Interspecific Gene Flow from Oilseed Rape to Wild Relatives", in H.C.M. den Nijs, *et al.* (eds.), *Introgression from Genetically Modified Plants into Wild Relatives*, CABI Pub, pp. 235-251.
- Chiang, M.S. (1974), "Cabbage Pollen Germination and Longevity", *Euphytica*, 23, pp. 579-584.

- Chiang, M.S., *et al.* (1977), "Transfer of Resistance to Race 2 of *Plasmodiophora brassicae* from *Brassica napus* to Cabbage (*B. oleracea* var. *capitata*). I. Interspecific Hybridization between *B. napus* and *B. oleracea* var. *capitata*", *Euphytica*, 26, pp. 319-336.
- Chiang, M.S., *et al.* (1980), "Transfer of Resistance to Race 2 of *Plasmodiophora brassicae* from *Brassica napus* to Cabbage (*B. oleracea* var. *capitata*): IV. A Resistant 18-Chromosome B1 Plant and its B2 Progeny", *Euphytica*, 29, pp. 47-55.
- Choudhary, B.R. and P. Joshi (1999), "Interspecific Hybridization in *Brassica*", *Proc. 10th Int. Rapeseed Congr.*, Australia, Contribution No 516, www.regional.org.au/au/gcirc/4/516.htm
- Christey, M.C. (2004), "*Brassica* Protoplast Culture and Somatic Hybridization", in E.C. Pua and C.J. Douglas (eds.), *Biotechnology in Agriculture and Forestry*, Springer, New York, Vol. 54, pp. 119-148.
- Chyi, Y.S., *et al.* (1992), "A Genetic Linkage Map of Restriction Fragment Length Polymorphism Loci for *Brassica rapa* (syn. *campestris*)", *Genome*, 35, pp. 746-757.
- Colbach, N., *et al.* (2005), "Spatial Aspects of Gene Flow between Rapeseed Varieties and Volunteers", *Agron. Sustain. Dev.*, 25, pp. 355-368.
- Coulthart, M.B. and K E. Denford (1982), "Isozyme Studies in *Brassica*. I Electrophoretic Techniques for Leaf Enzymes and Comparison of *B. napus*, *B. campestris* and *B. oleracea* Using Phosphoglucosyltransferase", *Can. J. Plant Sci.*, 62, pp. 621-630.
- Craig, A. and S. Millam (1995), "Modification of Oilseed Rape to Produce Oils for Industrial Use by Means of Applied Tissue Culture Methodology", *Euphytica*, 85, pp. 323-327.
- Crawford, J.W., *et al.* (1999), "Modelling Spread of Herbicide Resistance in Oilseed Rape", in A.J. Gray, *et al.* (eds.), *Environmental Impact of Genetically Modified Crops*, Research Report No. 10, DETR, London, pp. 97-106.
- Crawley, M.J. and S.L. Brown (1995), "Seed Limitation and the Dynamics of Feral Oilseed Rape on the M25 Motorway", *Proc. Roy. Soc. Lond.*, 259, pp. 49-54.
- Crawley, M.J., *et al.* (1993), "Ecology of Transgenic Oilseed Rape in Natural Habitat", *Nature*, 363, pp. 620-623.
- Crawley, M.J., *et al.* (2001), "Transgenic crops in natural habitats", *Nature*, 409, pp. 682-683.
- Cresswell, J.E. (1994), "A Method for Quantifying the Gene Flow that Results from a Single Bumblebee Visit Using Transgenic Oilseed Rape, *Brassica napus* L. cv. Westar", *Transgen. Res.*, 3, pp. 134-137.
- Daniels, R., *et al.* (2005), "The Potential for Dispersal of Herbicide Tolerance Genes from Genetically-modified, Herbicide-tolerant Oilseed Rape Crops to Wild Relatives", DEFRA Final Report, pp. 23.
- Darmency, H. and A. Fleury, (2000), "Mating System in *Hirschfeldia incana* and Hybridization to Oilseed Rape", *Weed Res.*, 40, pp. 231-238.
- Darmency, H., *et al.* (1995), "Effect of Transgenic Release on Weed Biodiversity: Oilseed Rape and Wild Radish", *Proc. Brighton Crop Protection Conf. Weeds*, BCPC, 2, pp. 433-438.
- Darmency, H., *et al.* (1998), "Spontaneous Hybridizations between Oilseed Rape and Wild Radish", *Mol. Ecol.*, 7, pp. 1467-1473.
- Dass, H., and N. Nybom, (1967), "The Relationships between *Brassica nigra*, *B. campestris*, *B. oleracea*, and their Amphidiploid Hybrids Studied by Means of Numerical Chemotaxonomy", *Can. J. of Genet. Cytol.*, 9, pp. 880-890.
- Daun, J.K. (1983), "The Introduction of Low Erucic Acid Rapeseed Varieties into Canadian Production", in J.K.G. Kramer, *et al.* (eds.), *High and Low Erucic Acid Rapeseed*, Academic Press., Toronto, pp. 161-195.
- Davey, V.M. (1959), "Cultivated *Brassicaceae*: Information Available to the Breeder", *Scottish Soc. Res. Plant Breed. Report*, pp. 23-62.
- De Candolle, A. (1886), *Origin of Cultivated Plants*, London.
- De Nettancourt, D. (1971), *Incompatibility in Angiosperms*, Springer-Verlag, Berlin, Heidelberg, New York, pp. 230.

- Deol, J.S., *et al.* (2003), “*Enarthrocarpus lyratus*-based Cytoplasmic Male Sterility and Fertility Restorer System in *Brassica rapa*”, *Plant Breed.*, 122, pp. 438-440.
- Devaux, C., *et al.* (2007), “Modelling and Estimating Pollen Movement in Oilseed Rape (*Brassica napus*) at the Landscape Scale Using Genetic Markers”, *Molecular Ecology*, 16, pp. 487-499.
- Dhillon, S.S., *et al.* (1985), “Root Tumours in Interspecific Crosses of *Brassica*”, *Cruciferae, Newsletter*, 10, pp. 27.
- Dickinson, H.G. (1995), “Dry Stigmas, Water and Self-incompatibility in *Brassica*”, *Sex. Plant Repr.*, 8, pp. 1-10.
- Dickson, M.H. and D.H. Wallace (1986), “Cabbage Breeding”, in M.J. Bassett (ed.), *Breeding Vegetable Crops*, AVI Publishing, pp. 295-432.
- Diederichsen, E. and M.D. Sacristan (1988), “Interspecific Hybridizations in the Genus *Brassica* Followed by In-ovule Embryo Culture”, *Cruciferae Newsl. Eucarpia*, 13, pp. 20-21.
- Dietz-Pfeilstetter, A. and P. Zwerger (2004), “Dispersal of Herbicide Resistance Genes during the Large Scale Cultivation of Different Transgenic, Herbicide Resistant Oilseed Rape Varieties”, *J. Plant Diseases Protect, Sonderheft XIX*, pp. 831-838.
- Dietz-Pfeilstetter, A. and P. Zwerger (2009), “In-field Frequencies and Characteristics of Oilseed Rape with Double Herbicide Resistance”, *Environ. Biosafety Res.*, 8, pp. 100-111.
- Dietz-Pfeilstetter, A., *et al.* (2006), “Assessment of Transgene Spread from Oilseed Rape by Population Dynamic and Molecular Analyses of Feral Oilseed Rape”, *J. Plant Diseases Protect, Sonderheft XX*, pp. 39-47.
- Dietz-Pfeilstetter, E., *et al.* (1998), “Untersuchungen zum Auskreuzungsverhalten von Basta-tolerant Winterraps auf Nicht Transgenem Raps (*Brassica napus*)”, *Mitt. a. d. Biol. Bundesanst.*, H.257, pp. 121 (in German).
- Dignam, M. (2001), *Bush, Parks and Road and Rail Weed Management Survey*, Mark Dignam and Associates, Australia, pp. 24.
- Dixelius, C. (1999), “Inheritance of the Resistance to *Leptosphaeria maculans* of *Brassica nigra* and *B.juncea* in Near-isogenic Lines of *B. napus*”, *Plant Breed.*, 118, pp. 151-156.
- Dixon, G.R. (2007), *Vegetable Brassicas and Related Crucifers*, Crop Production Science in Horticulture Series: 14, CABI, Oxfordshire, UK. pp. 327.
- Downey, R.K. (1964), “A Selection of *Brassica campestris* L. Containing No Erucic Acid and in its Seed Oil”, *Can. J. Plant Sci.*, 44, pp. 295.
- Downey, R.K. (1983), “The Origin and Description of the Brassica Oilseed Crops”, in J.K.G. Kramer, *et al.* (eds.), *High and Low Erucic Acid Rapeseed Oils*, Academic Press, Toronto, pp. 1-20.
- Downey, R.K. (1996), “Diversification of Canola/Rapeseed Fatty Acid Supply for the Year 2000”, *OCL* 3, pp. 9-13.
- Downey, R.K. (1999a), “Risk Assessment of Outcrossing of Transgenic *Brassica*, with Focus on *B. rapa* and *B. napus*”, *Proc. 10th Int. Rapeseed Congr.*, Australia, Contribution No. 61, www.regional.org.au/au/gcirc/4/61.htm
- Downey, R.K. (1999b), “Gene Flow and Rape – the Canadian Experience”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 109-116.
- Downey, R.K. and J.M. Armstrong (1962), “Inter-compatibility and Performance of Diploid and Derived Tetraploid Rape (*Brassica campestris* L.)”, *Can. J. Plant Sci.*, 42, pp. 672-680.
- Downey, R.K. and H. Beckie (2002), “Isolation Effectiveness in Canola Pedigree Seed Production”, Internal Res. Rep., Agric. and Agri-Food Can., Saskatoon Res. Cent., Saskatoon, SK, Canada.
- Downey, R.K. and J. Buth (2003), “Transgenic Rapeseed – Grower Adoption and Consumer Acceptance”, *Proc. 11th Rapeseed Congr.*, Copenhagen, pp. 1190-1194.
- Downey, R.K. and G.F.W. Rakow (1987), “Rapeseed and Mustard”, in W. Fehr (ed.), *Principles of Cultivar Development Vol. 2 Crop Species*, Macmillan, New York, pp. 437-486.
- Downey, R.K. and S.R. Rimmer (1993), “Agronomic Improvement in Oilseed Brassicas”, *Advances in Agron.*, 50, pp. 1-66.

- Downey, R.K. and G. Röbbelen (1989), "Brassica Species". in G. Röbbelen, *et al.* (eds.), *Oil Crops of the World*, McGraw-Hill, New York, pp. 339-362.
- Downey, R.K., *et al.* (1980), "Rapeseed and Mustard", in W.R. Fehr and H.H. Hadley (eds.), *Hybridization of Crop Plants*, Amer. Soc. Crop Sci, pp. 495-509.
- Dunfield, K.E. and J. J. Germida (2001), "Diversity of Bacterial Communities in the Rhizosphere and Root-interior of Field-grown Genetically Modified *Brassica napus*", *FEMS Microbiol.*, 38, pp. 1-9.
- Dunfield, K.E. and J. J. Germida (2003) "Seasonal Changes in the Rhizosphere Microbial Communities Associated with Field-grown Genetically Modified Canola (*Brassica napus*)", *Appl. Environ. Microbiol.*, 69, pp. 7310-7318.
- Dunfield, K.E. and J. J. Germida (2004), "Impact of Genetically Modified Crops on Soil- and Plant-associated Microbial Communities", *J. Environ. Qual.*, 33, pp. 806-815.
- Dushenkov, S., *et al.* (2002), "Bioengineering of a Phytoremediation Plant by Means of Somatic Hybridization", *Int. J. Phytoremediation*, 4, pp. 117-126.
- Eastham, K. and J. Sweet (2002), "Genetically Modified Organisms (GMOs): The Significance of Gene Flow through Pollen Transfer", *Env. Issue Rep.*, No.28, pp. 75, European Environment Agency, Copenhagen, Denmark, http://reports.eea.europa.eu/environmental_issue_report_2002_28/en/GMOs%20for%20www.pdf
- Eber, F., *et al.* (1994), "Spontaneous Hybridization between a MaleSterile Oilseed Rape and Two Weeds", *Theor. Appl. Genet.*, 88, pp. 362-368.
- Ekbom, B. (1995), "Insect Pests", in D.S. Kimber and D.I. McGregor (eds.), *Brassica Oilseeds, Production and Utilization*, CAB International, Oxon, UK, pp. 141-152.
- Elliott, R.H., *et al.* (2008), "Effects of Seed Size and Seed Weight on Seedling Establishment, Vigor and Tolerance of Argentine Canola (*Brassica napus*) to Flea Beetles, *Phyllotreta* spp", *Can. J. Plant Sci.*, 88, pp. 207-217.
- Ellis, R.H., *et al.* (1994), "The Long-term Storage of Seeds of Seventeen Crucifers at Very Low Moisture Contents", *Plant Genetics Resources Newsletter*, 98, pp. 32.
- Erickson, L.R., *et al.* (1983), "Restriction Patterns Reveal Origins of Chloroplast Genomes in *Brassica* Amphidiploids", *Theor. Appl. Genet.*, 65, pp. 201-206.
- Eskin, N.A.M., *et al.* (1989), "Stability of Low Linolenic Acid Canola Oil to Frying Temperature", *J. Am. Oil Chem. Soc.*, 66, pp. 1081-1084.
- European Commission (1998a), "Opinion of the Scientific Committee on Plants regarding the Glufosinate Tolerant Hybrid Rape Derived from Genetically Modified Parental Lines (MS8xRF3) Notified by Plant Genetic Systems (Notification C/B/96/01)", Submitted by the Scientific Committee on Plants 19 May 1998, pp. 7.
- European Commission (1998b), "Opinion of the Scientific Committee on Plants regarding Submission for Placing on the Market of Glufosinate Tolerant Oilseed Rape Transformation Event Liberator pHOE 6/AC Notified by the AgrEvo Company [Now Aventis CropScience] (Notification C/DE/98/6)", pp. 9.
- European Commission (1999), "Opinion of the Scientific Committee on Plants, Adopted on 18 May 1999, on the Invocation by France of Article 16 ('Safeguard Clause') of Council Directive 90/220/EEC with Respect to a Genetically Modified Oilseed Rape Notification C/UK/94/M1/1 (Plant Genetic Systems N.V.)-(SCP/GM/150-final)", pp. 3.
- European Commission (2000), "Opinion Regarding Submission for Placing on the Market of Glufosinate Tolerant Oilseed Rape Transformation Event Liberator PHOE 6/AC Notified by the Hoechst schering AGREVO COMPANY [NOW AVENTIS CROPSCIENCE] (Notification C/DE/98/6)", http://ec.europa.eu/food/fs/sc/scp/out88_gmo_en.html
- Fahleson, J., *et al.* (1994a), "Intertribal Somatic Hybrids between *Brassica napus* and *Thlaspi perfoliatum* with High Content of the *T. perfoliatum*-specific Nervonic Acid", *Theor. Appl. Genet.*, 87, pp. 795-804.
- Fahleson, J., *et al.* (1994b), "Intertribal Somatic Hybrids between *Brassica napus* and *Barbarea vulgaris* - Production of in Vitro Plantlets", *Plant Cell Rep.*, 13: pp. 411-416.

- Falk, K.C. and D.L. Woods (2003), "Seed Yield of Successive Synthetic Generations in Summer Turnip Rape", *Can. J. Plant Sci.*, 83, pp. 271-274.
- Falk, K.C. *et al.* (1994). "Performance of inter-cultivar summer turnip rape hybrids in Saskatchewan". *Can. J. Plant Sci.* 74, pp. 441-445.
- Fan, Z., Tai, W. and B.R. Stefansson (1985), "Male Sterility in *Brassica napus* L. Associated with an Extra Chromosome", *Can. J. Genet. Cytol.*, 27, pp. 467-471.
- FAOSTAT (2009), FAO Statistics online database, <http://faostat.fao.org/>, accessed 10 November 2011.
- Farnham, M.W. (2007), "Vegetable Crucifers –Status Report", www.ars-grin.gov/npgs/cgc_reports/crucifer1201.htm
- Fenwick, J.W., *et al.* (1983), "Glucosinolates and their Breakdown Products in Food and Food Plants", *Crit. Rev. Food Sci. Nutr.*, 18, pp. 123-201.
- Fernandez-Escobar, J., *et al.* (1988), "Genetics of Erucic Acid Content in Interspecific Hybrids of Ethiopian Mustard (*Brassica carinata* Braun) and Rapeseed (*B. napus* L.)", *Pl. Breed.*, 100, pp. 310-315.
- Ferrie, A.M. and W.A. Keller (2004), *Brassica* Improvement through Microsporiculture, in E.C. Pua and C.J. Douglas (eds.), *Biotechnology in Agriculture and Forestry*, Vol. 54: Brassica, pp. 148-168. Springer.
- Finkelstein, R.R., *et al.* (1985), "Role of ABA in Maturation of Rapeseed Embryos", *Plant Physiol.*, 78, pp. 630-636.
- Flannery, M.L., *et al.* (2006), "Plastid Genome Characterization in *Brassica* and Brassicaceae Using a New Set of Nine SSRs", *Theor. Appl. Genet.*, 113, pp. 1221-1231.
- Ford, C.S., *et al.* (2006), "Spontaneous Gene Flow from Rapeseed (*Brassica napus*) to Wild *Brassica oleracea*", *Proc. R. Soc. B.*, 273, pp. 3111-3115.
- Forsberg, J., *et al.* (1994), "Fertile Somatic Hybrids between *Brassica napus* and *Arabidopsis thaliana*", *Plant Sci.*, 95, pp. 213-223.
- Forsberg, J., *et al.* (1998), "UV dose-dependent DNA elimination in asymmetric hybrids between *Brassica napus* and *Arabidopsis thaliana*". *Plant Sci.*, 131, pp. 65-76.
- Förster, K. and W. Diepenbrock, (2002), "Use of Genetically Modified Plants – Consequences for Crop Production", *Schwellenwerte für Produkte aus Gentechnisch Veränderten Pflanzen*, Mitt. DFG, 7, pp. 50-58.
- Frandsen, K.J. (1943), "The Experimental Formation of *Brassica juncea* Czern. et Coss.", *Dansk. Bot. Ark.*, 11: pp. 1-17.
- Frandsen, K.J. (1947), "The Experimental Formation of *Brassica napus* L. var. *oleifera* DC. and *Brassica carinata* Braun", *Dansk Bot. Ark.*, 12, pp. 1-16.
- Frello, S., *et al.* (1995), "Inheritance of Rapeseed (*Brassica napus*) – Specific RAPD Markers and a Transgene in the Cross *B. juncea* × (*B. juncea* × *B. napus*)", *Theor. Appl. Genet.*, 91, pp. 236-241.
- Friesen, L.F., *et al.* (2003), "Evidence of Contamination of Pedigreed Canola (*Brassica napus*) Seed Lots in Western Canada with Genetically Engineered Herbicide Resistance Traits", *Agron. J.*, 95, pp. 1342-1347.
- Fu, T.D., *et al.* (1997), "Rapeseed varieties and improvement in China", *GCIRC Bulletin*, 14, pp. 90-95.
- Fujimoto, R. *et al.* (2006), "Suppression of Gene Expression of a Recessive *SP11/SCR* Allele by an Intrinscribed *SP11/SCR* Allele in *Brassica* Self-incompatibility", *Plant Mol. Biol.*, 61, pp. 577-587.
- Fukui, K.S., *et al.* (1998), "Quantitative Karyotyping of Three Diploid *Brassica* Species by Imaging Methods and Localization of 45S rDNA Loci on the Identified Chromosomes", *Theor. Appl. Genet.*, 96, pp. 325-330.
- George, R.A.T. (2009), *Vegetable Seed Production 3rd Edition*, CABI Publishing, Oxon, UK, pp. 320.
- Getinet, A., *et al.* (1994), "Development of Zero Erucic Acid Ethiopian Mustard through an Interspecific Cross with Zero Erucic Oriental Mustard", *Can. J. Plant Sci.*, 74, pp. 793-795.
- Gleba, Y.Y. and F. Hoffmann (1979), "*Arabidobrassica*: Plant Genome Engineering by Protoplast Fusion", *Naturwissenschaften*, 66, pp. 547-554.

- Gleba, Y.Y. and F. Hoffmann (1980), “*Arabidobrassica*: A Novel Plant Obtained by Protoplast Fusion”, *Planta*, 149, pp. 112-117.
- Gliddon, C.J. (1999), “Transgenic Plants: Field Testing and Commercialisation Including a Consideration of Novel Herbicide Resistant Oilseed Rape (*Brassica napus* L.)”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 49-56.
- Glimelius, K. (1999), “Somatic Hybridization”, in C. Gómez-Campo (Ed.), *Biology of Brassica Coenospecies*, Elsevier Science, Amsterdam, The Netherlands.
- Gómez-Campo, C. (1980), “Morphology and Morpho-taxonomy of the Tribe *Brassicaceae*”, in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, Japan Sci. Soc. Press, Tokyo, pp. 3-31
- Gómez-Campo, C. (1983), “Studies on Cruciferae: X. Concerning Some West Mediterranean Species of *Erucastrum*”, *An. Jard. Bot. Madrid*, 40, pp. 63-72.
- Gómez-Campo, C. (1999), “Taxonomy”, in C. Gómez-Campo (ed.), *Biology of Brassica Coenospecies*, Elsevier, Amsterdam, pp. 3-32.
- Gómez-Campo, C. and M.E. Tortosa (1974), “The Taxonomic and Evolutionary Significance of Some Juvenile Characters in Brassicaceae”, *Bot. J. Linn. Soc.*, 69, pp. 105-124.
- GoshDastidar, N. and N.S. Varma (1999), “A Study on Intercrossing between Transgenic *B. juncea* and Other Related Species”, *Proc. 10th Int. Rapeseed Congr.*, Australia, Contribution No. 244, www.regional.org.au/au/gc/4/244.htm.
- Government of Tasmania, Australia (2003), “Tasmanian Rural and Marine Industry Profiles, 2004”, [www.dpipwe.tas.gov.au/inter.nsf/Attachments/CART-64Z6Q9/\\$FILE/RAMIP-Aug%2004-Full_dd.pdf](http://www.dpipwe.tas.gov.au/inter.nsf/Attachments/CART-64Z6Q9/$FILE/RAMIP-Aug%2004-Full_dd.pdf), accessed 21 May 2012.
- Gowers, S. (1982), “The Transfer of Characters from *Brassica campestris* L. to *Brassica napus* L.: Production of Clubroot-resistant Oil-seed Rape (*Brassica napus* subsp. *oleifera*)”, *Euphytica*, 31: pp. 971-976.
- Gruber, M.Y., *et al.* (2006), ““HAIRY CANOLA” - Arabidopsis *GL3* Induces a Dense Covering of Trichomes on *Brassica napus* Seedlings”, *Plant Molecular Biol.*, 60, pp. 679-698.
- Gruber, S. and W. Claupein (2007), “Fecundity of Volunteer Oilseed Rape and Estimation of Potential Gene Dispersal by a Practice-related Model”, *Agric. Ecosys. Environ.*, 119, pp. 401-408.
- Gruber, S., *et al.* (2003), “Life Cycle and Gene Dispersal of Oilseed Rape Volunteers (*Brassica napus* L.)”, *Proc. BCPC Int. Cong.: Crop Sci. and Tech.*, Glasgow, Scotland, UK, pp. 1093-1098.
- Gruber, S., *et al.* (2004), “Seed Persistence of Oilseed Rape (*Brassica napus*): Variation in Transgenic and Conventionally Bred Cultivars”, *J. Agric. Sci.*, 142, pp. 29-40.
- Gruber, S., *et al.* (2009), “Classification of Canola (*Brassica napus*) Winter Cultivars by Secondary Dormancy”, *Can. J. Plant Sci.*, 89, pp. 613-619.
- Grundy, S.M. and M. Denke (1990), “Dietary Influences on Serum Lipids and Lipoproteins”, *J. Lipid Res.*, 31, pp. 1149-1172.
- Guéritaine, G., *et al.* (2003), “Emergence and Growth of Hybrids between *Brassica napus* and *Raphanus raphanistrum*”, *New Phytol.*, 158, pp. 561-567.
- Guéritaine, G., *et al.* (2002), “Fitness of Backcross Six of Hybrids between Transgenic Oilseed Rape (*Brassica napus*) and Wild Radish (*Raphanus raphanistrum*)”, *Molecular Ecology*, 11, pp. 1419-1426.
- Gulden, R.H., *et al.* (2003a), “Harvest Losses of Canola (*Brassica napus*) Cause Large Seedbank Inputs”, *Weed Sci.*, 51, pp. 83-86.
- Gulden, R.H., *et al.* (2003b), “Secondary Seed Dormancy Prolongs Persistence of Volunteer Canola in Western Canada”, *Weed Sci.*, 51, pp. 904-913.
- Gulden, R.H., *et al.* (2004a), “Relative Contribution of Genotype, Seed Size and Environment to Secondary Seed Dormancy Potential in Canadian Spring Oilseed Rape (*Brassica napus*)”, *Weed Res.*, 44, pp. 97-106.

- Gulden, R.H., *et al.* (2004b), “Secondary Dormancy, Temperature and Burial Depth Regulate Seedbank Dynamics in Canola”, *Weed Sci.*, 52, pp. 382-388.
- Gupta, S.K. (1997), “Production of Interspecific and Intergeneric Hybrids in *Brassica* and *Raphanus*”, *Cruciferae Newslett. Eucarpia*, 19, pp. 21-22.
- Gurr, M.I. (1992), “Dietary Lipids and Coronary Heart Disease: Old Evidence, New Perspective”, *Prog. Lipid Res.*, 31, pp. 195-243.
- Habman, M.H., *et al.* (2010), “Exploitation of a late flowering species *Brassica oleracea* L. for the improvement of earliness in *B. napus* L.: an untraditional approach”. *Euphytica*, 177, pp. 365-374.
- Hails, R.S., *et al.* (1997), “Burial and Seed Survival in *Brassica napus* subsp. *oleifera* and *Sinapis arvensis* Including a Comparison of Transgenic and Non-transgenic Lines of the Crop”, *Proc. Roy. Soc. Lond.*, 264, pp. 1-7.
- Håkansson, A. (1956), “Seed development of *Brassica oleracea* and *B. rapa* after certain reciprocal pollinations”, *Hereditas*, 42, pp. 373-395.
- Halfhill, M.D., *et al.* (2002), “*Bt*-transgenic Oilseed Rape Hybridization with its Weedy Relative, *Brassica rapa*”, *Environ. Safety Res.*, 1, pp. 19-28.
- Hall, L., *et al.* (2000), “Pollen Flow between Herbicide-resistant *Brassica napus* Volunteers”, *Weed Sci.*, 48, pp. 688-694.
- Hall, L.M., *et al.* (2005), “Volunteer Oilseed Rape – Will Herbicide Resistance Traits Assist Fertility?”, in J. Gressel (ed.), *Crop Fertility and Volunteerism*, CRC London Press, pp. 59-79.
- Hansen, L.B., *et al.* (2001), “Introgression between Oilseed Rape (*Brassica napus* L.) and its Weedy Relative *B. rapa* L. in a Natural Population”, *Genet. Res. Crop Evol.*, 48, pp. 621-627.
- Hansen, L.N. (1998), “Intertribal Somatic Hybridization between Rapid Cycling *Brassica oleracea* L. and *Camelina sativa* (L.) Crantz”, *Euphytica*, 104, pp. 173-179.
- Harberd, D.J. (1972), “A Contribution to the Cytotaxonomy of *Brassica* (Cruciferae) and its Allies”, *Bot. J. Linn. Soc.*, 65, pp. 1-23.
- Harberd, D.J. (1976), “Cytotaxonomic Studies of *Brassica* and Related Genera”, in J.G. Vaughan, *et al.* (eds.), *The Biology and Chemistry of the Cruciferae*, Academic Press, London, pp. 47-68.
- Harberd, D.J. and E.D. McArthur (1972), “Cytotaxonomy of *Rhynchosinapis* and *Hutera* (Cruciferae-Brassicaceae)”, *Heredity*, 28, pp. 254-257.
- Harberd, D.J. and E.D. McArthur (1980), “Meiotic Analysis of Some Species and Genus Hybrids in the Brassicaceae”, in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, Japan Sci. Soc. Press, Tokyo, pp. 65-67.
- Harker, K.N., *et al.* (2006), “Persistence of Glyphosate-resistant Canola in Western Canadian Cropping Systems”, *Agron. J.*, 98, pp. 107-119.
- Harper, F.R. and B. Berkenkamp (1975), “Revised Growth-stage Key for *Brassica campestris* and *B. napus*”, *Can. J. Plant Sci.*, 55, pp. 657-658.
- Hatakeyama K., *et al.* (1998), “Dominance Relationships between *S*-alleles in Self-incompatible *Brassica campestris* L.”, *Heredity*, 80, pp. 241-247.
- Hauser, T.P., *et al.* (1997), “Preferential Exclusion of Hybrids in Mixed Pollinations between Oilseed Rape (*Brassica napus*) and Weedy *B. campestris* (Brassicaceae)”, *Am. J. Bot.*, 81, pp. 436-443.
- Hauser, T.P., *et al.* (2001), “Environmental Effects on Male and Female Fitness of *Brassica* hybrids”, *European Science Foundation Meeting of a Working Group on: Interspecific Gene Flow from Oilseed Rape to Weedy Species*, June 2001, Rennes, France, pp. 24.
- Hayter, K.E. and J.E. Cresswell (2006), “The Influence of Pollinator Abundance on the Dynamics and Efficiency of Pollination in Agricultural *Brassica napus*: Implications for Landscape-scale Gene Dispersal”, *J. Appl. Ecol.*, 43, pp. 1196-1202.

- Heather, D.W. and J.B. Sieczka, (1991), "Effect of seed size and cultivar on emergence and stand establishment of broccoli in crusted soil". *J. Amer. Soc. Hort. Sci.* 116, pp. 946-949.
- Hedge, I.C. (1976), "A Systematic and Geographical Survey of the Old World Cruciferae", in J. C. Vaughan, *et al.* (eds.), *The Biology and Chemistry of the Cruciferae*, Academic Press, New York, pp. 1-45.
- Helm, J. (1963), "Morphologisch-taxonomische Gleiderung der Kultursippen von *Brassica oleracea*", *Kulturpflanze*, 11, pp. 92-210.
- Hemingway, J.S. (1995), "Mustards: *Brassica spp.* and *Sinapis alba* (Cruciferae)", in J. Smartt and N.W. Simmons (eds.), *Evolution of Crop Plants*, Longman, London, pp. 82-86.
- Herklots, G.A.C. (1972), *Vegetables in South-East Asia: Asiatic Cabbages*, George Allen & Unwin, London, pp. 190-224.
- Hertz, B. (1999), *Expert Testimony in Monsanto Canada and Monsanto Company vs. Percy Schmeiser and Schmeiser Enterprises*, Federal Court-Trial Division.
- Heyn, F.W. (1977), "Transfer of Restorer Genes from *Raphanus* to Cytoplasmic Male Sterile *Brassica napus*", *Cruciferae Newsl.*, 1, pp. 15-16.
- Heywood, V.H. (1964), "Brassica", in T.G. Tutin, *et al.* (eds.), *Flora Europaea*, 1, Cambridge University Press, pp. 335-339.
- Hilhorst, H.W.M. and P.E. Toorop (1997), "Review on Dormancy, Germinability and Germination in Crop and Weed Species", *Adv. Agron.*, 61, pp. 111-165.
- Hill, S.A., *et al.* (1991), "The Incidence and Importance of Beet Western Yellows Virus in Oilseed Rape", *IOBC/WPRS Bulletin*, 14, pp. 36-45.
- Hinata, K. and T. Nishio (1980), "Self-incompatibility in Crucifera", in S. Tunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, pp.223-234.
- Hinata, K., *et al.* (1994), "Manipulation of Sporophytic Self-incompatibilities on Plant Breeding", in E.G. Williams, *et al.* (eds.), *Genetic Control of Self-incompatibility and Reproductive Development in Flowering Plants*, Kluwer, Dordrecht, pp. 102-115.
- Holm, L., *et al.* (1997), *World Weeds, Natural Histories and Distribution*, John Wiley & Sons, N.Y., pp. 1129.
- Horricks, J.S. (1969), "Influence of Rape Residue on Cereal Production", *Can. J. Plant Sci.*, 49, pp. 632-634.
- Hosaka, K., *et al.* (1990), "Development and Chromosomal Localization of Genome Specific DNA Markers of *Brassica* and the Evolution of Amphidiploids and $n=9$ Diploid Species", *Genome*, 33, pp. 131-142.
- Hu, F.B., *et al.* (1997), "Dietary Fat Intake and the Risk of Coronary Heart Disease in Women", *New Engl. J. Med.*, 337, pp. 1491-1499.
- Hu, J. and C.F. Quiros (1991), "Molecular and Cytological Evidence of Deletions in Alien Chromosomes for Two Monosomic Addition Lines of *Brassica campestris-oleracea*", *Theor. Appl. Genet.*, 90, pp. 258-262.
- Hu, Q., *et al.* (1999), "Intergeneric Hybridization by Protoplast Fusion Aiming at Modification of Fatty Acid Composition in *Brassica napus* L.", *Proc. 10th Int. Rapeseed Congr.*, Canberra, Australia, Contribution No. 332, pp. 26-29.
- Hu, Q., *et al.* (2002), "Intergeneric Hybrid between *Brassica napus* and *Orychophragmus violaceus* Containing Traits of Agronomic Importance for Oilseed Rape Breeding", *Theor. Appl. Genet.*, 105, pp. 834-840.
- Hua, Y.W. and Z.Y. Li, (2006), "Genomic in Situ Hybridization Analysis of *Brassica napus* × *Orychophragmus violaceus* Hybrids and Production of *B. napus* Aneuploids", *Plant Breed.*, 125, pp. 144-149.
- Huang, B., *et al.* (2002), "Production and Cytogenetics of Intergeneric Hybrids between Ogura CMS *Brassica napus* and *Raphanus raphanistrum*", *Cruciferae Newsl. Eucarpia*, 24, pp. 25-27.
- Hüsken, A. and A. Dietz-Pfeilstetter (2007), "Pollen-mediated Intraspecific Gene Flow from Herbicide Resistant Oilseed Rape (*Brassica napus* L.)", *Transgenic Res.*, 16, pp. 557-569.

- Inaba, R. and T. Nishio (2002), “Phylogenetic Analysis of *Brassicaceae* Based on the Nucleotide Sequences of the S-locus Related Gene, SLR1”, *Theor. Appl. Genet.*, 105, pp. 1159-1165.
- Ingram, J. (2000), “The Separation Distances Required to Ensure Cross-pollination is below Specified Limits in Non-seed Crops of Sugar Beet, Maize and Oilseed Rape”, *Plant Var. Seeds*, 13, pp. 1981-1999.
- Inomata, N. (1988), “Intergeneric Hybridization between *Brassica napus* and *Sinapis arvensis* and their Crossability”, *Cruciferae Newsl.*, 13, pp. 22-23.
- Inomata, N. (2007), “*Brassica* Vegetable Crops”, in R. Singh (ed.), *Genetic Resources, Chromosome Engineering, and Crop Improvement*, CRC Press, New York, pp. 115-146.
- Jackson, S.A., et al. (2000), “Comparative Fluorescence *in Situ* Hybridization Mapping of a 431-kb *Arabidopsis thaliana* Bacterial Artificial Chromosome Contig Reveals the Role of Chromosomal Duplications in the Expansion of the *Brassica rapa* genome”, *Genetics*, 156, pp. 833-838.
- Jenkins, T.E., et al. (2001), “Investigating Gene Introgression from Rape to Wild Turnip”, *New Zealand Plant Protection*, 54, pp. 101-104.
- John Innes Centre, “RevGenUK”, Norwick, UK, <http://revgenuk.jic.ac.uk/>, accessed 27 Oct 2011.
- Johnston J.S., et al. (2005), “Evolution of Genome Size in Brassicaceae”, *Ann. Bot.*, 95, pp. 229–235.
- Jørgensen, R.B. (1999), “Gene Flow from Oilseed Rape (*Brassica napus*) to Related Species”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 117-124.
- Jørgensen, R.B. and B. Andersen (1994), “Spontaneous Hybridization between Oilseed Rape (*Brassica napus*) and Weedy *B. campestris* (Brassicaceae): a Risk of Growing Genetically Modified Oilseed Rape”, *J. Bot.*, 81, pp. 1620-1626.
- Jørgensen, R.B., et al. (1996), “Spontaneous Hybridization between Oilseed Rape (*Brassica napus*) and Weedy Relatives”, *Acta Hort.*, 407, pp. 193-200.
- Jørgensen, R.B., et al. (1998), “Introgression of Crop Genes from Oilseed Rape (*Brassica napus*) to Related Wild Species – an Avenue for the Escape of Engineered Genes”, *Acta Hort.*, 459, pp. 211-217.
- Jørgensen, T., et al. (2007), “Adventitious Presence of Other Varieties in Oilseed Rape (*Brassica napus*) from Seed Bank and Certified Seeds”, *Seed Sci. Res.*, 17, pp. 115-125.
- Kamler, F. (2000), “Transgene Rape and Honey Bees”, *Proc. 8th Int. Pollination Symp.*, Mosonmagyaróvár, Hungary, July.
- Karssen, C.M. (1980/81), “Environmental Conditions and Endogenous Mechanisms Involved in Secondary Dormancy of Seeds”, *Israel J. Bot.*, 29, pp. 45-64.
- Kerlan, M.C., et al. (1993), “Interspecific Hybrids between a Transgenic Rapeseed (*Brassica napus*) and Related Species - Cytogenetical Characterization and Detection of the Transgene”, *Genome*, 36, pp. 1099-1106.
- Kerlan, M.C., et al. (1992), “Risk Assessment of Outcrossing of Transgenic Rapeseed to Related Species: I. Interspecific Hybrid Production under Optimal Conditions with Emphasis on Pollination and Fertilization”, *Euphytica*, 62, pp. 145-153.
- Khanna, V.K. and J.B. Chowdhury (1974), “Studies on Pollen Tube Growth and Seed Set in Some Interspecific Crosses in Genus *Brassica*”, *J. Res.*, 4, pp. 231-235.
- Kirk, J.T.O. and R.N. Oram (1981) “Isolation of Erucic Acid-free Lines of *Brassica juncea*: Indian Mustard Now a Potential Oilseed Crop in Australia”, *J. Aust. Inst. Agric. Sci.*, 47, pp. 51-52.
- Kirti, P.B., et al. (1995), “Transfer of *Ogu* Cytoplasmic Male Sterility to *Brassica juncea* and Improvement of Male Sterile through Somatic Cell Fusion”, *Theor. Appl. Genet.*, 91, pp. 517-521.
- Kirti, P.B., et al. (1997), “Introgression of a Gene Restoring Fertility to CMS (*Trachystoma*) *Brassica juncea* and the genetics of Restoration”, *Plant Breed.*, 116, pp. 259-262.
- Kjaer, A. (1960), “Naturally Derived Isothiocyanates (Mustard Oils) and their Parent Glucosides”, in L. Zechmeister (ed.), *Progress in the Chemistry of Organic Natural Products*, Springer-Verlag, Vienna, pp. 122-176.

- Klein, E.K., *et al.* (2006), "Pollen Dispersal of Oilseed Rape: Estimation of the Dispersal Function and Effects of Field Dimension", *J. Appl. Ecol.*, 43, pp. 141-151.
- Klewer, A., *et al.* (2003), "Incorporation of Blackspot Resistance from Different Origins into Oilseed Rape", *Proc. 11th Int. Rapeseed Congr.*, Copenhagen, 6-10 July, 1, pp. 65-67.
- Knispel, A.L., *et al.* (2008), "Gene Flow and Multiple Herbicide Resistance in Escaped Canola Populations", *Weed Science*, 56, pp. 72-80.
- Koul, K.K., *et al.* (2000), "Seed Coat Microsculpturing in *Brassica* and Allied Genera (Subtribes Brassicinae, Raphaninae, Moricandiinae)", *Annals of Botany*, 86, pp. 385-397.
- Korzun, V. (2003), "Molecular Markers and their Applications in Cereals Breeding", in Proc. of "Marker Assisted Selection: A fast track to increase genetic gain in plant and animal breeding?", 17-18 Oct. 2003, Turin, Italy, Rome, FAO, pp.18-22, www.fao.org/biotech/docs/Korzun.pdf
- Kramer, J.K.G. *et al.* (1983), *High and Low Erucic Acid Rapeseed Oils*, Academic Press, Toronto, pp. 582.
- Kuckuck, H. (1979), *Gartenbauliche Pflanzenzüchtung*, 2nd ed, Parey, Berlin, Hamburg, pp. 64-72.
- Kumazawa, S. and S. Able (1955), "Studies on Varieties of Mustard", *J. Jpn. Soc. Hort. Sci.*, 24, pp. 69-84.
- Kusaba, M., *et al.* (2001), "Self-incompatibility in the Genus *Arabidopsis*: Characterization of the *S* Locus in the Outcrossing *A. lyrata* and its Autogamous Relative *A. thaliana*", *Plant Cell*, 13, pp. 627-643.
- Lagercrantz, U. and D. Lydiate (1996) "Comparative Genome Mapping in *Brassica*", *Genetics*, 144, pp. 1903-1910.
- Lal, G. (1993), "Seed Production Technology for Tropical and Subtropical Cauliflower", in *Breeding of Solanaceous and Cole Crops*, AVRDC, Chinese Taipei, pp. 268-279.
- Lamb, R.J. (1989), "Entomology of Oilseed *Brassica* Crops", *Ann. Rev. Entomol.*, 34, pp. 211-229.
- Landbo, L. and R.B. Jørgensen (1997), "Seed Germination in Weedy *Brassica campestris* and its Hybrids with *Brassica napus*: Implications for Risk Assessment of Transgenic Oilseed Rape", *Euphytica*, 97, pp. 209-218.
- Landbo, L., *et al.* (1996), "Natural Hybridisation between Oilseed Rape and a Wild Relative: Hybrids among Seeds from Weedy *B. campestris*", *Hereditas*, 125, pp. 89-91.
- Landry, B.S., *et al.* (1991), "A Genetic Map for *Brassica napus* Based on Restriction Fragment Length Polymorphisms Detected with Expressed DNA Sequences", *Genome*, 34, pp. 543-552.
- Lavigne, C., *et al.* (1998), "A Pollen Dispersal Experiment with Transgenic Oilseed Rape. Estimation of the Average Pollen Dispersal of an Individual Plant within a Field", *Theor. Appl. Genet.*, 96, pp. 886-896.
- Leadlay, E.A. and V.H. Heywood (1990), "The Biology and Systematics of the Genus *Coincya* Porta and Rigo ex Rouy (Cruciferae)", *Bot. J. Linn. Soc.*, 102, pp. 313-398.
- Leckie, D., *et al.* (1993), "Gene Movement from Oilseed Rape to Weedy Populations – a Component of Risk Assessment for Transgenic Cultivars", *Aspects of Appl. Bio.*, 35, pp. 61-66.
- Leeson, J.Y., *et al.* (2005), *Prairie Weed Surveys of Cereal, Oilseed and Pulse Crops from the 1970s to the 2000s*, Weed Survey Series Publ., 05-1, AAFC, Saskatoon, pp. 395.
- Leflon, M., *et al.* (2006), "Pairing and Recommendation at Meiosis of *Braeeica rapa* (AA) × *Brassica napus* (AACC) Hybrids", *Theor. Appl. Genet.*, 113, pp. 1467-1480.
- Lefol, E., *et al.* (1991), "Escape of Engineered Genes from Rapeseed to Wild Brassiceae", *Proc. Brighton Crop Protection Conf. Weeds*, 3, pp. 1049-1056.
- Lefol, E., *et al.* (1995), "Gene Dispersal from Transgenic Crops. I. Growth of Interspecific Hybrids between Oilseed Rape and the Wild Hoary Mustard", *J. Appl. Ecology*, 32, pp. 803-808.
- Lefol E., *et al.* (1996a), "Gene Dispersal from Transgenic Crops. II. Hybridization between Oilseed Rape and Wild Hoary Mustard", *Sex. Plant Reprod.*, 9, pp. 189-196.
- Lefol, E., *et al.* (1996b), "Predicting Hybridization between Transgenic Oilseed Rape and Wild Mustard", *Field Crop Res.*, 45, pp. 153-161.

- Lefol, E., *et al.* (1997), "Sexual Hybridization in Crosses of Cultivated Brassica Species with the Crucifers *Erucastrum gallicum* and *Raphanus raphanistrum*: Potential for Gene Introduction", *Euphytica*, 95, pp. 127-139.
- Légère, A., *et al.* (2001), "Presence and Persistence of Volunteer Canola in Canadian Cropping Systems", *Proc. Brighton Crop Protection Conf. Weeds*, BCPC, pp. 143-148.
- Lelivelt, C.L.C. and F.K. Krens (1992), "Transfer of Resistance to the Beet Cyst Nematode (*Heterodera schachtii* Schm) into the *Brassica* Gene Pool through Intergeneric Somatic Hybridization with *Raphanus sativus* L.", *Theor. Appl. Genet.*, 83, pp. 887-894.
- Lelivelt, C.L.C., *et al.* (1993), "Transfer of Resistance to the Beet Cyst Nematode (*Heterodera schachtii* Schm.) from *Sinapis alba* L. (White Mustard) to the *Brassica napus* L. Gene Pool by Means of Sexual and Somatic Hybridization", *Theor. Appl. Genet.*, 85, pp. 688-696.
- Lim, K.B., *et al.* (2005), "Characterization of rDNAs and Tandem Repeats in the Heterochromatin of *Brassica rapa*", *Mol Cells.*, 19(3), pp. 436-444.
- Linder, R. (1998), "Potential Persistence of Transgenes: Seed Performance of Transgenic Canola and Wild Relatives × Canola Hybrids", *Ecological Applications*, 8, pp. 1180-1195.
- Linder, R. and J. Schmitt (1995), "Potential Persistence of Escaped Transgenes: Performance of Transgenic Oil-modified *Brassica* Seeds and Seedlings", *Ecological. Appl.*, 5, pp. 1056-1068.
- Liu, H. (1985), *Genetics and Breeding of Rapeseed*, Shanghai Sci. and Tech. Press, Shanghai, China. pp. 592. (in Chinese)
- Liu, J., *et al.* (2005), "Intergeneric Somatic Hybridization and its Application to Crop Genetic Improvement", *Plant Cell Tissue Organ Cult.*, 82, pp. 19-44.
- Liu, J.H., *et al.* (1996), "Transfer of the *Brassica tournefortii* Cytoplasm to *B. napus* for the Production of Cytoplasmic Male Sterile *B. napus*", *Physiologia Plantarum*, 96, pp. 123-129.
- Lizgunova, T.V. (1959), "The History of Botanical Studies of the Cabbage, *Brassica oleracea* L.", *Bull. Appl. Bot., Genet. Pl. Breed.*, 32, pp. 37-70.
- Lokanadha, R.D. and N. Saria, (1994), "Hybridization of *Brassica tournefortii* and Cultivated Brassicas", *Cruciferae Newsl. Eucarpia*, 16, pp. 32-33.
- López-Granados, F. and P.J.W. Lutman (1998), "Effect of Environmental Conditions on the Dormancy and Germination of Volunteer Oilseed Rape Seed (*Brassica napus*)", *Weed Sci.*, 46, pp. 419-423.
- Love, H.K., *et al.* (1990), "Development of Low Glucosinolate Mustard", *Can. J. Plant Sci.*, 70, pp. 425-429.
- Lutman, P.J.W., *et al.* (2003), "The Long-term Persistence of Seeds of Oilseed Rape (*Brassica napus*) in Arable Fields", *J. Agric. Sci.*, 141, pp. 231-240.
- Lutman, P.J.W., *et al.* (2005), "Persistence of Seeds from Crops of Conventional and Herbicide Tolerant Oilseed Rape (*Brassica napus*)", *Proc. R. Soc. B.*, 272, pp. 1909-1915.
- Lysak, M.A. and C. Lexer (2006), "Towards the Era of Comparative Evolutionary Genomics in *Brassicaceae*", *Pl. Syst. Evol.*, 259, pp. 175-198.
- Lysak, M.A., *et al.* (2005), "Chromosome Triplication Found Across the Tribe *Brassicaceae*", *Genome Res.*, 15, pp. 516-525.
- Lysak, M.A., *et al.* (2006), "Mechanisms of Chromosome Number Reduction in *Arabidopsis thaliana* and Related *Brassicaceae* Species", *Proc. Natl. Acad. Sci. USA (PNAS)*, 103, pp. 5224-5229.
- Lysak, M.A., *et al.* (2007), "Ancestral Chromosomal Blocks are Triplicated in *Brassicaceae* Species with Varying Chromosome Number and Genome Size", *Plant Physiol.*, 145, pp. 402-410.
- Lysak, M.A., *et al.* (2009), "The Dynamic Ups and Downs of Genome Size Evolution in *Brassicaceae*.", *Mol. Biol. Evol.*, 26, pp. 85-98.

- MacDonald, R.L. and G.J. Kuntz (2000), "Monitoring Program to Assess the Occurrence and Fate of SeedLink Canola Volunteers Following the 1999 Growing Season in Western Canada", *Aventis CropScience Report*, No. ACC00-03, pp. 14.
- Manasse, R. and P. Kareiva (1991), "Quantifying the Spread of Recombinant Genes and Organisms", in L. Ginzburgh (ed.), *Assessing Ecological Risks of Biotechnology*, Butterworth-Heinemann, Boston, pp. 215-231.
- Martens, G. (2001), "From Cinderella to Cruella: Volunteer Canola", *2nd Annual Manitoba Agronomists Conference 2001, the University of Manitoba, Canada*, pp. 151-154.
- Marquard, R. and K.C. Walker (1995), "Environmental Impact of Rapeseed Production", in D.S. Kimber and D.I. McGregor (eds.), *Brassica Oilseeds, Production and Utilization*, CAB International, Oxon, UK, pp. 195-213.
- Masden, S.B. (1962), "Germination of Buried and Dry Stored Seed III. 1934-1960", *Proc. Int. Seed Testing Assoc.*, 27, pp. 920-928.
- Mason-Sedun, W., *et al.* (1986), "Differential Phytotoxicity among Species and Cultivars of the Genus *Brassica* to Wheat. I. Laboratory and Field Screening of Species", *Plant and Soil*, 93, pp. 3-16.
- Mathias, R. (1991), "Improved Embryo Rescue Technique for Intergeneric Hybridization between *Sinapis* Species and *B. napus*", *Cruciferae Newsl.*, 14/15, pp. 90-91.
- Mattson, F.H. and S.M. Grundy (1985), "Comparison of Effects of Dietary Saturated, Monounsaturated, and Polyunsaturated Fatty Acids on Plasma Lipids and Lipoproteins in Man", *J. Lipid Res.*, 26, pp. 194-201.
- Maxwell, M (1981), "Production of a Heinz Body Anaemia in the Domestic Fowl after Ingestion of Dimethyl Disulphide: a Haematological and Ultrastructural Study", *Rev. Veterinary Sci.*, 30, pp. 233-238.
- McCallum, C.M., *et al.* (2000), "Targeting Induced Local Lesions in Genomes (TILLING) for Plant Functional Genomics", *Plant Physiol.*, 123, pp. 439-42.
- McCartney, H.A. and M.E. Lacey (1991), "Wind Dispersal of Pollen from Crops of Oilseed Rape (*Brassica napus* L.)", *J. Aerosol. Sci.*, 22(4), pp. 467-477.
- McGrath, J.M., *et al.* (1990), "Identification of *Brassica oleracea* Monosomic Alien Chromosome Addition Lines with Molecular Markers Reveals Extensive Gene Duplication", *Mol. Gen. Genetics*, 223, pp. 198-204.
- McNaughton, I.H. (1995a), "Turnip and Relatives *Brassica campestris* (Cruciferae)", in J. Smartt and N.W. Simmonds (eds.), *Evolution of Crop Plants*, Longman, London, pp. 62-68.
- McNaughton, I.H. (1995b), "Swedes and Rapes *Brassica napus* (Cruciferae)", in J. Smartt and N.W. Simmonds (eds.), *Evolution of Crop Plants*, Longman, London, pp. 68-75
- Meng, J., *et al.* (1998), "The Production of Yellow-seeded *Brassica napus* (AACC) through Crossing Interspecific Hybrids *B. campestris* (AA) and *B. carinata* (BBCC) with *B. napus*", *Euphytica*, 103, pp. 329-333.
- Mesquida, J. and M. Renard (1982), "Study of the Pollen Dispersal by Wind and of the Importance of Wind Pollination in Rapeseed (*Brassica napus* var. *oleifera* Metzger)", *Apidologie*, 4, pp. 353-366, (English summary).
- Messéan, A., *et al.* (2007), "Occurrence of Genetically Modified Oilseed Rape in the Harvest of Subsequent Conventional Oilseed Rape over Time", *Europ. J. Agron.*, 27, pp. 115-122.
- Metz, P.L.J., *et al.* (1997), "The Impact on Biosafety of the Phosphinothricin-tolerance Transgene in Inter-specific *B. rapa* × *B. napus* Hybrids and their Successive Backcrosses", *Theor. Appl. Genet.*, 95, pp. 442-450.
- Metz, P.L.J., *et al.* (1995), "Hybridization of Radish (*Raphanus sativus* L.) and Oilseed Rape (*Brassica napus* L.) through a Flower-culture Method", *Euphytica*, 80, pp. 159-168.
- Mikkelsen, T.R., *et al.* (1996), "Inheritance of Oilseed Rape (*Brassica napus*) RAPD Markers in a Backcross Progeny with *Brassica campestris*", *Theor. Appl. Genet.*, 92, pp. 492-497.
- Mizushima U. (1972), "Evolution of Species in Brassiceae and their Breeding", *Kagaku to Seibutsu*, 10, pp. 78-85.
- Mizushima, U. (1980), "Genome Analysis in *Brassica* and Allied Genera", in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies, Biology and Breeding*, Jap. Scient. Soc. Press, Tokyo, pp. 89-108.

- Mizushima, U. and S. Tsunoda (1967), "A Plant Exploration in Brassica and Allied Genera", *Tohoku J. Agr. Res.*, 17, pp. 249-276.
- Mohammad, A. (1935), "Pollination Studies on Toria (*Brassica napus* L. var *dichotoma* Prain) and Sarson (*Brassica campestris* Prain)", *Indian J. Agric. Sci.*, 5, pp. 125-154.
- Momoh, E.J.J., *et al.* (2002), "Variation in the Development of Secondary Seed Dormancy in Oilseed Rape Genotypes under Conditions of Stress", *Weed Res.*, 42, pp. 446-455.
- Morinaga, T. (1928), "Preliminary Note on Interspecific Hybridization in *Brassica*", *Proc. Imp. Acad.*, 4, pp. 620-622.
- Morinaga, T. (1929), "Interspecific Hybridization in *Brassica*. II. The Cytology of F₁ Hybrids of *B. cernua* and Various Other Species with 10 Chromosomes", *Jap. J. Bot.*, 4, pp. 277-289.
- Morinaga, T. (1931), "Interspecific Hybridization in *Brassica*. IV. The Cytology of F₁ Hybrids of *B. carinata* and Some Other Species with 10 Chromosomes", *Cytologia*, 3, pp. 77-83.
- Morinaga, T. (1933), "Interspecific Hybridization in *Brassica*. V. The Cytology of F₁ Hybrids of *B. carinata* and *B. alboglabra*", *Japan J. Bot.*, 6, pp. 467-475.
- Morinaga, T. (1934a), "Interspecific Hybridization in *Brassica*: VI. The Cytology of F₁ Hybrids of *B. juncea* and *B. nigra*", *Cytologia*, 6, pp. 62-67.
- Morinaga, T. (1934b), "On the Chromosome Number of *Brassica juncea* and *B. napus*, on the Hybrid between the two, and on Offspring Line of the Hybrid", *Japan J. Genet.*, 9, pp. 161-163.
- Morris, W.K., *et al.* (1994), "Do Barren Zones and Pollen Traps Reduce Gene Escape from Transgenic Crops?", *Ecol. Appl.*, 4, pp. 157-165.
- Moyes, C.L., *et al.* (2002), "Barriers to Gene Flow from Oilseed Rape (*Brassica napus*) into Populations of *Sinapis arvensis*", *Mol. Ecol.*, 11, pp. 103-112.
- Mullis, K. and F. Faloona (1987), "Specific Synthesis of DNA *in Vitro* via a Polymerase-catalyzed Chain Reaction", *Meth. Enzyme.*, 44, pp. 224-166.
- Mun, J-H., *et al.* (2009), "Genome-wide Comparative Analysis of the *Brassica rapa* Gene Space Reveals Genome Shrinkage and Differential Loss of Duplicate Genes after Whole Genome Triplication", *Genome Biology*, 10, pp. 1-18.
- Nagpal, R., *et al.* (1996), "Transfer of *Brassica tournefortii* (TT) Genes to Allotetraploid Oilseed *Brassica* Species (*B. juncea* AABB, *B. napus* AACC, *B. carinata* BBCC): Homoeologous Pairing is More Pronounced in the Three-genome Hybrids (TACC, TBAA, TCAA, TCBB) as Compared to Allodiploids (TA, TB, TC)", *Theor. Appl. Genet.*, 92, pp. 566-571.
- Namai, H., *et al.* (1980), "Interspecific and Intergeneric Hybridization Breeding in Japan", in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies: Biology and Breeding*, Japan Sci. Soc. Press, Tokyo, pp 191-204.
- Narasimhulu, S.B., *et al.* (1994), "Intergeneric Protoplast Fusion between *Brassica carinata* and *Camelina sativa*", *Plant Cell Rep.*, 13, pp. 657-660.
- Nasrallah, J.B., *et al.* (1991), "The Self-incompatibility Genes of *Brassica*: Expression and Use in Genetic Ablation of Floral Tissues", *Annu. Rev. Plant Mol. Biol.*, 43, pp. 393-422.
- Navabi, Z. K., *et al.* (2010) "*Brassica* B-genome resistance to stem rot (*Sclerotinia sclerotiorum*) in a doubled haploid population of *Brassica napus* x *Brassica carinata*", *Can. J. Plant Pathol.* 32, pp. 237-246.
- Navrátilová, B. (2004), "Protoplast Cultures and Protoplast Fusion Focused on Brassicaceae – a Review", *Hort. Sci.*, 31, pp. 140-157.
- Navrátilová, B., *et al.* (1997), "Construction of Intergeneric Somatic Hybrids between *Brassica oleracea* and *Armoracia rusticana*", *Biol. Plant*, 39, pp. 531-541.
- Neutrofal, F. (1927), "Zytologische Studien uber die Kulturrassen von *Brassica oleracea*", *Ost. Bot. Z.*, 76, pp. 101-113.
- Nieuwhof, M. (1969), *Cole Crops*, World Crop Books, London, pp. 353.

- Nishi, S. (1968) “ ‘HAKURAN’ . An Artificial Synthesized Heading *Brassica napus*”, *JARQ.*, 3, pp. 18-22.
- Nishi, S. (1980), “Differentiation of *Brassica* Crops in Asia and the Breeding of “Hakuran”, a Newly Synthesized Leafy Vegetable”, in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, Japan Sci. Soc. Press, Tokyo, pp. 133-150.
- Nishiyama, I., *et al.* (1991), “Critical Discussion on Abortive Interspecific Crosses in *Brassica*”, *Plant Breeding*, 107, pp. 288-302.
- Nitovskaya, I.O. and A.M. Shakhovskiy (1998), “Obtaining of Asymmetrical Somatic Hybrids between *Brassica oleracea* L. and *Arabidopsis thaliana* L.”, *Tsitol. Genet.*, 32, pp. 72-81. (in Russian; English Abstr.)
- Nitovskaya, I.O., *et al.* (1998), “Production of “Brassicapsella” Somatic Hybrids Based on the Double Inactivation of Donor Protoplasts”, *Dopov. Nats. Akad. Nauk. Ukr.*, 0(9), pp. 181-186. (in Ukrainian; English Abstr.)
- Nitovskaya, I.O., *et al.* (2006a), “Production of *Brassica oleracea* (+*Arabidopsis thaliana*) and *Brassica napus* Cell Lines Resistant to Spectinomycin/Streptomycin as a Result of Plastome Genetic Transformation”, *Tsitol Genet.*, 40, pp. 3-10. (in Ukrainian; English Abstr.)
- Nitovskaya, I.O., *et al.* (2006b), “Construction of Cybrid Transplastomic *Brassica napus* Plants Containing *Lesquerella fendleri* Chloroplasts”, *Tsitol Genet.*, 40, pp. 11-21. (in Ukrainian; English Abstr.)
- Nou, I.S., *et al.* (1993), “Comparison of S-alleles and S-glycoproteins between Two Wild Populations of *Brassica campestris* in Turkey and Japan”, *Sex. Plant Repro.*, 6, pp. 79-86.
- Norris, C.E. and J. Sweet (2002), “Monitoring Large Scale Releases of Genetically Modified Crops (EPG 1/5/84)”, *Final Report of Monitoring Studies of Field Scale Releases of GM Oilseed Rape Crops in England from 1994-2000 (NIAB)*, pp. 120.
- Norris, C.E., *et al.* (1999), “Monitoring Weediness and Persistence of Genetically Modified Oilseed Rape (*Brassica napus*) in the UK”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 255-260.
- Ockendon, D. (2000), “The S-allele Collection of *Brassica oleracea*”, *Acta Hort.*, 539, pp. 25-30.
- OECD (Organisation for Economic Co-operation and Development) (1997), *Consensus Document on the Biology of Brassica napus L. (Oilseed rape)*, Series on Harmonisation of Regulatory Oversight of Biotechnology, No. 7, OECD, Paris, pp. 28.
- OECD (2011), *Revised Consensus Document on Compositional Considerations for New Varieties of Low Erucic Acid Rapeseed (Canola): Key Food and Feed Nutrients, Anti-nutrients and Toxicants*, Series on the Safety of Novel Foods and Feeds, No. 24, OECD, Paris, pp. 38.
- OECD, “OECD Seed Schemes website”, www.oecd.org/tad/seed, accessed 17 Nov 2011.
- Ogbonnaya, F.C., *et al.* (2003), “Progress in the Utilization of *Brassica nigra* in Breeding for Resistance to Black Leg (*Leptosphaeria maculans*)”, *Proc 11th Int. Rapeseed Congr.*, Copenhagen, Denmark, 1, pp. 39-41.
- Ohkawa, Y. (1986), “Comparison of *B. napus* to *B. campestris* and *B. oleracea* based on the Cytoplasmic Characters; Cytoplasmic Male Sterility and Chloroplast DNA”, *JARQ Japan Agricultural Research Quarterly*, 19, pp. 253-258.
- Ohio State Univ. (2009), “Vegetable seed production – “dry”seeds”. Deptment of Horticulture and Crop Science.<http://extension.osu.edu/~seedsci/vsp02.html>. Accessed 11/09/2009.
- Oikarinen, S. and P.H. Ryöppy (1992), “Somatic Hybridization of *Brassica campestris* and *Barbarea* species”, *Proc. XIIIth Eucarpia Congress: Reproductive Biology and Plant Breeding*, Angers, France, 6-11 July, pp. 261-262.
- Okamoto, S., *et al.* (2007), “Self-compatibility in *Brassica napus* is Caused by Independent Mutations in the S-locus Genes”, *Plant J.*, 50, pp. 391-400.
- Olsson, G. (1960), “Species Crosses within the Genus *Brassica*. II. Artificial *Brassica juncea* Coss.”, *Hereditas*, 46, pp. 171-222.

- Opeña, R.T., *et al.* (1988), “Breeding and Seed Production of Chinese Cabbage in the Tropics and Suntropics”, *Tech. Bull.*, No. 17, AVRDC, Chinese Taipei.
- Orson, J. (2002), “Gene Stacking in Herbicide Tolerant Oilseed Rape: Lessons from the North American Experience”, *English Nature Research Reports*, No. 443, pp. 17.
- Østergaard, L., *et al.* (2006), “Pod Shatter-resistant *Brassica* Fruit Produced by Ectopic Expression of the *FRUITFULL* Gene”, *Plant Biotechnology J.*, 4, pp. 45–51.
- Ovcharenko, O.O., *et al.* (2004), “Obtaining of Intertribal *Brassica juncea* + *Arabidopsis thaliana* Somatic Hybrids and Study of Transgenic Trait Behaviour”, *Tsitol Genet.*, 38, pp. 3-8.
- Ovcharenko, O.O., *et al.* (2005), “Creation and Analysis of *Brassica napus* + *Arabidopsis thaliana* Somatic Hybrids Possessing Maize Spm/dspm Heterologous Transposable System”, *Tsitol Genet.*, 39, pp. 50-56.
- Palmer, J.D. (1988), “Intraspecific Variation and Multicircularity in *Brassica* Mitochondrial DNAs”, *Genetics*, 118, pp. 341-351.
- Palmer, J.D., *et al.* (1983), “Chloroplast DNA Evolution and the Origin of Amphiploid *Brassica* Species”, *Theor. Appl. Genet.*, 65, pp. 181 – 189.
- Palmer, T.P. (1962), “Population Structure, Breeding System, Interspecific Hybridization and Allopolyploidy”, *Heredity*, 17, pp. 278-283.
- Parkin, I.A., *et al.* (1995), “Identification of the A and C Genomes of Amphiploid *Brassica napus* (Oilseed Rape)”, *Genome*, 38, pp. 1122-1131.
- Parkin, I.A., *et al.* (2003), “Patterns of Genome Duplication within *Brassica napus* Genome”, *Genome*, 46, pp. 291-303.
- Parkin, I.A., *et al.* (2005), “Segmental Structure of the *Brassica napus* Genome Based on Comparative Analysis with *Arabidopsis thaliana*”, *Genetics*, 171, pp. 765-781.
- Pascher, K.P., *et al.* (2006), “Feral Oilseed Rape – Investigations on its Potential for Hybridization”, *Final Report to the commission of the Federal Ministry of Health and Women*, Austria, pp. 85.
- Paul, E.M., *et al.* (1995), “Gene Dispersal from Genetically Modified Oilseed Rape in the Field”, *Euphytica*, 81, pp. 283-289.
- Paulmann, W. and G. Röbbelen (1988), “Effective Transfer of Cytoplasmic Male Fertility from Radish (*Raphanus sativus* L.) to Rape (*Brassica napus* L.)”, *Plant Breed.*, 100, pp. 299-309.
- PBO (Plant Biotechnology Office, Canada) (1998), “Decision Document 98-25: Determination of Environmental Safety of Rhône Poulenc’s Oxynil Herbicide-tolerant *Brassica napus* Canola Line Westar Oxy-235”, pp. 5.
- Pekrun, C. and P.J.W. Lutman (1998), “The Influence of Post-harvest Cultivation on the Persistence of Volunteer Oilseed Rape”, *Asp. Appl. Biol.*, 51, pp. 113–118.
- Pekrun, C., *et al.* (1997a), “Induction of Secondary Dormancy in Rape Seeds (*Brassica napus* L.) by Prolonged Imbibitions of Water Stress or Oxygen Deficiency in Darkness”, *European J. Agron*, 6, pp. 245-255.
- Pekrun C., *et al.* (1997b), “Genotypic Variation in the Development of Secondary Dormancy in Oilseed Rape and its Impact on the Persistence of Volunteer Rape”, *Proc. Brighton Crop Protection Conf. Weeds*, pp. 243-248.
- Pekrun C., *et al.* (1998a), “Cultural Control of Volunteer Oilseed Rape (*Brassica napus*)”, *J. Agric. Sci.*, 130, pp. 155-163.
- Pekrun, C., *et al.* (1998b), “Research on Volunteer Rape: A Review”, *Pflanzenbauwissenschaften*, 2, pp. 84-90.
- Pessel, F.D., *et al.* (2001), “Persistence of Oilseed Rape (*Brassica napus* L.) Outside of Cultivated Fields”, *Theor. Appl. Genet.*, 102, pp. 841-846.
- Peterka, H., *et al.* (2004), “Transfer of Resistance against the Beet Cyst Nematode from Radish (*Raphanus sativus*) to Rape (*Brassica napus*) by Monosomic Chromosome Addition”, *Theor. Appl. Genet.*, 109, pp. 30-41.

- Pinder, R., *et al.* (1999), "Evaluating the Risk of Transgene Spread from *Brassica napus* to Related Species", in P.J.W. Lutman (ed.), *Gene Flow and Agriculture: Relavance for Transgene Crops*, Proc. BCPC Sym, 72, pp. 275-280.
- Pivard, S., *et al.* (2008), "Where do the Feral Oilseed Rape Populations Come from? A Large Scale Study of their Possible Origin in a Farmland Area", *J. Appl. Ecology*, 45, pp. 476-485.
- Plieske, J., *et al.* (1998), "Inheritance of Resistance derived from the B-genome of *Brassica* against *Phoma lingam* and the Development of Molecular Markers", *Theor. Appl. Genet.*, 97, pp. 929-936.
- Plümper, B. (1995), "Somatische und Sexuelle Hybridisierung für den Transfer von Krankheitsresistenzen auf *Brassica napus* L.", PhD Thesis, Free University of Berlin, Berlin.
- Potapov, D.A. and G.M. Osipova (2003), "Development of Yellow Seeded *Brassica napus* in Siberia", *Proc. 11th Int. Rapeseed Congr.*, Copenhagen, Denmark, pp 250-252.
- Prain, D. (1898), "The Mustards Cultivated in Bengal", *Agr. Ledger*, 5, pp. 1-80.
- Prakash, O. (1961), *Food and Drinks in Ancient India*, Mitra. R., Delhi, pp. 265-266.
- Prakash, S., *et al.* (2001), "Expression of Male Sterility in Alloplasmic *Brassica juncea* with *ErUCAstrum canariense* Cytoplasm and Development of Fertility Restoration System", *Plant Breed.*, 120, pp. 178-182.
- Prakash, S., *et al.* (2009), "*Brassica* and its Close Allies: Cytogenetics and Evolution", *Plant Breeding Rev.*, 31, pp. 21-187.
- Prakash, S. and V. L.Chopra (1991), "Cytogenetics of Crop Brassicas and their Allies", in T. Tsuchiya and P.K. Gupta (eds.), *Chromosome Engineering in Plants: Genetics, Breeding, Evolution*, Elsevier, Amsterdam, pp. 161-180.
- Prakash, S. and K. Hinata (1980), "Taxonomy, Cytogenetics and Origin of Crop Brassicas, a Review", *Opera. Bot.*, 55, pp. 3-57.
- Prakash, S., *et al.* (1995), "Cytoplasmic Male Sterility (CMS) Systems Other Than *ogu* and Polima in *Brassicacae*: Current Status", *Proc. 9th Int. Rapeseed Congr*, Cambridge, U.K., 1, pp. 44-48.
- Prakash, S., *et al.* (1998), "A *Moricandia arvensis* Based Cytoplasmic Male Sterility and Fertility Restoration System in *Brassica juncea*", *Theor. Appl. Genet.*, 97, pp. 488-492.
- Primard, C., *et al.* (1988), "Interspecific Somatic Hybridization between *Brassica napus* and *Brassica hirta* (*Sinapis alba* L.)", *Theor. Appl. Genet.*, 75, pp. 546-552.
- Price, J.S., *et al.* (1996), "Seed Losses in Commercial Harvesting of Oilseed Rape", *J. Agric. Eng. Res.*, 65, pp. 183-191.
- Przybylski, R., *et al.* (1993), "Stability of Low Linolenic Acid Canola Oil to Accelerated Storage at 60 Degrees C", *Lebensm Wiss & Technol.*, 26, pp. 205-209.
- Purugganan, M.D., *et al.* (2000), "Variation and Selection at the CAULIFLOWER Floral Homeotic Gene Accompanying the Evolution of Domesticated *Brassica oleracea*", *Genetics*, 155, pp. 855-862.
- Qi, C.K., *et al.* (1995), "A Successful Transfer of Yellow-seeded Trait from *Brassica carinata* to *B. napus*", *Proc. 9th Int. Rapeseed Congr.*, Cambridge, U.K., 4-7 July, 4, pp. 1137-1140.
- Qi, X., *et al.* (2007), "Molecular Phylogeny of Chinese Vegetable Mustard (*Brassica juncea*) Based on the Internal Transcribed Spacers (ITS) of Nuclear Ribosomal DNA", *Genet. Res. Crop Evol.*, 54, pp. 1709-1716.
- Quazi, M.H. (1988), "Interspecific Hybrids between *Brassica napus* L. and *B. oleracea* L. Developed by Embryo Culture", *Theor. Appl. Genet.*, 75, pp. 309-318.
- Quiros, C.F., *et al.* (1988), "Exploring the Role of x=7 Species in *Brassica* Evolution: Hybridization with *B. nigra* and *B. oleracea*", *J. Hered.*, 79, pp. 351-358.
- Rajhathy, T. (1976), "Haploid Flax Revisited", *Z. Pflanzenzüchtg*, 67, pp. 1-10.
- Rakow, G. (2004), "Species Origin and Economic Importance of *Brassica*", *Biotechnology in Agric. and Forestry*, 54, pp. 6-7.

- Rakow, G. and D. Rode (2009b), "AC Vulcan Oriental Mustard", *Can. J. Plant Sci.*, 89, pp. 325-329.
- Rakow, G. and D. L. Woods (1987), "Outcrossing in Rape and Mustard under Saskatchewan Prairie Conditions", *Can. J. Plant Sci.*, 67, pp. 147-151.
- Rakow, G., *et al.* (2009a), "Centennial Brown Brown Condiment Mustard", *Can. J. Plant Sci.*, 89, pp. 337-340.
- Rahman, M.H. (2001), "Production of Yellow Seeded *Brassica napus* through Interspecific Crosses", *Plant Breed.*, 120, pp. 463-472.
- Ramanujam, S. and D. Srinivasachar (1943), "Cytogenetic Investigations in the Genus *Brassica* and the Artificial Synthesis of *B. juncea*", *Indian J. Genet. Pl. Breed.*, 3, pp. 73-88.
- Ramsay, G., *et al.* (1999), "Honeybees as Vectors of GM Oilseed Rape Pollen", in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 209-214.
- Ramsay, G., *et al.* (2003), "Quantifying Landscape-scale Gene Flow in Oilseed Rape", Department for Environment, Food and Rural Affairs, London, UK, http://webarchive.nationalarchives.gov.uk/20081023141438/http://www.defra.gov.uk/environment/gm/research/pdf/epg_rg0216.pdf
- Rana, D., *et al.* (2004), "Conservation of the Microstructure of Genome Segments on *Brassica napus* and its Diploid Relatives", *The Plant J.*, 40, pp. 725-733.
- Ranito-Lehtimäki, A. (1995), "Aerobiology of Pollen and Pollen Antigens", in C.S. Cox and C.M. Wathes (eds.), *Bioaerosols Handbook*, CRC Lewis Press: Boca Raton, Florida, pp. 387-406.
- Rasche, E. and M. Gadsby (1997), "Glufosinate-ammonium Tolerant Crops – International Commercial Developments and Experience", *Proc. Brighton Crop Protection Conf. Weeds*, pp. 941-946.
- Rashid, A., *et al.* (1994), "Development of Yellow Seeded *Brassica napus* through Interspecific Crosses", *Plant Breeding*, 112, pp. 127-134.
- Raybould, A.F. and A.J. Gray (1993), "Genetically modified crops and hybridization with wild relatives: A UK perspective". *J. Appl. Ecology*, 30, pp.199-219.
- Reboud, X. (2003), "Effect of Gap on Gene Flow between Otherwise Adjacent Transgenic *Brassica napus* Crops", *Theor. Appl. Genet.*, 106, pp. 1048-1058.
- Ren, J.P., *et al.* (2001a), "Improved Resistance to Bacterial Softrot by Protoplast Fusion between *Brassica rapa* and *B. oleracea*", *Theor. Appl. Genet.*, 100, pp. 810-819.
- Ren, J. P., *et al.* (2001b), "CC-14-1 and CC-18-2 Progenies of Chinese Cabbage Derived from Somatic Hybridization for Resistance to Bacterial Soft Rot", *Hort. Sci.*, 36, pp. 990-991.
- Renfrew, J.M. (1973), *Palaeoethnobotany: The Prehistoric Food Plants of the Near East and Europe*, Columbia University Press, New York.
- Reuter, H., *et al.* (2008), "Hazard Mitigation or Mitigation Hazard? Would Genetically Modified Dwarfed Oilseed Rape (*Brassica napus*) Increase Feral Survival?", *Environ. Sci. Pollut. Res.*, 15, pp. 529-535.
- Rice, E.L. (1984), *Allelopathy. 2nd Edn.*, Academic Press, New York, pp. 422.
- Rich, T.C.G. (1991), *Crucifers of Great Britain and Ireland*, B.S.B.I. Handbook, No.6., Botanical Society of the British Isles, London, pp. 336.
- Rieger, M.A., *et al.* (1999), "Risks of Gene Flow from Transgenic Herbicide-resistant Canola (*Brassica napus*) to Weedy Relatives in Southern Australian Cropping System", *Australian J. Agric. Res.*, 50, pp. 115-128.
- Rieger, M.A. *et al.* (2001), "Hybridization between *Brassica napus* L. and *Raphanus raphanistrum* L. under Agronomic Field Conditions", *Theor. Applied Genetics.*, 103, pp. 555-560.
- Rieger, M.A., *et al.* (2002), "Pollen-mediated Movement of Herbicide Resistance between Commercial Canola Fields", *Science*, 296, pp. 2386-2388.
- Rimmer, S.R., *et al.* (2007), "Compendium of *Brassica* Diseases", *APS Press St. Paul MN., USA*, pp. 117.

- Ringdahl, E.A., *et al.* (1987), "Intergeneric Hybridization of *Diplotaxis* spp. with *Brassica napus*: a Source of New CMS Systems?", *Can. J. Plant Sci.*, 67, pp. 239-243.
- Ripley, V.L. and P.G. Arnison (1990), "Hybridization of *Sinapis alba* L. and *Brassica napus* L. via Embryo Rescue", *Pl. Breed.*, 104, pp. 26-33.
- Röbbelen, G. (1960), "Beiträge zur Analyse des Brassica-Genomes", *Chromosoma*, 11, pp. 205-228.
- Röbbelen, G. and W. Thies (1980), "Variation in Rapeseed Glucosinolates and Breeding for Improved meal Quality", in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies: Biology and Breeding*, Japan Sci. Press, Tokyo, pp. 285-300.
- Rousselle, P., *et al.* (1985), "Restauration de la Fertilité pour l'Androsterilité Genocytosplasmique Chez le Colza (*Brassica napus* L.)", *Utilization des Raphano-Brassica. Agronomie*, 5, pp. 431-437.
- Roy, N.N. (1977), "Interspecific Cross Compatibility and F₁ Sterile Oilseed Brassicas", *Proc. 3rd Int. SABRAO Cong.*, 2, pp. 31-33.
- Roy, N.N. (1980), "Species Crossability and Early Generation Plant Fertility in Interspecific Crosses of *Brassica*", *SABRAO J.*, 12, pp. 43-54.
- Roy, N.N. (1984), "Interspecific Transfer of *Brassica juncea*-type High Blackleg Resistance to *Brassica napus*", *Euphytica*, 33, pp. 295-303.
- Ryschka, U., *et al.* (1999), "Somatic Cell Hybridization for Transfer of Disease Resistance in *Brassica*", in A. Altman, *et al.* (eds.), *Plant Biotechnology and in vitro Biology in the 21st Century*, Kluwer Academic Publishers, Amsterdam, pp. 205-208.
- Ryschka, U., *et al.* (2003), "High Frequency Recovery of Intergeneric Fusion Products of *Brassica oleracea* (+) *Lepidium meyenii* and their Molecular Characterization by RAPD and AFLP", *Acta. Hort.*, 625, pp. 145-149.
- Saal, B., *et al.* (2004), "Identification of a *Brassica juncea*-derived Recessive Gene Conferring Resistance to *Leptosphaeria maculans* in Oilseed Rape", *Plant Breed.*, 123, pp. 505-511.
- Sacristan, M.D. and M. Gerdemann (1986), "Different Behavior of *Brassica juncea* and *B. carinata* as Sources of *Phoma lingam* Resistance in Experiments of Interspecific Transfer to *B. napus*", *Pl. Breed.*, 97, pp. 304-314.
- Sadik, S. (1962), "Morphology of the Curd of Cauliflower", *Am. J. Bot.*, 49, pp. 290-297.
- Sakai, T., *et al.* (1996), "Introduction of a Gene from Fertility Restored Radish (*Raphanus sativus*) into *Brassica napus* by Fusion of X-irradiated Protoplasts from a Radish Restorer Line and Iodacetamide-treated Protoplasts from a Cytoplasmic Male-sterile Cybrid of *B. napus*", *Theor. Appl. Genet.*, 93, pp. 73-379.
- Saji, H., *et al.* (2005), "Monitoring the Escape of Transgenic Oilseed Rape around Japanese Ports and Roadsides", *Environ. Biosafety Res.*, 4, pp. 217-222.
- Sakhno, L.O., *et al.* (2007), "Phosphinothricin-resistant *Brassica napus* + *Orychophragmus violaceus* Somatic Hybrids", *Cytol. Genet.*, 41, pp. 1-5.
- Salisbury, P. (2002), *Genetically Modified Canola in Australia: Agronomic and Environmental Considerations*, R.K. Downey (ed.), Australian Oilseeds Federation, pp. 69.
- Sarwar, M. and J.A. Kirkegaard (1998), "Biofumigation Potential of Brassicas II. Effect of Environment and Ontogeny on Glucosinolate Production and Implications for Screening", *Plant and Soil*, 201 (1), pp. 91-101.
- Sauer, J.D. (1993), *Historical Geography of Crop Plants*, CRC Press, Boca Raton, Fl. USA. pp. 309.
- Sauermann, W. (1987), "Entwicklung des Glucosinolatgehalts von 00-Raps in den Nängigkeit vom Fremddurchwuchs", *Raps*, 5, pp. 12-13.
- Scarth, R., *et al.* (1988), "Stellar Low Linolenic High Linoleic Acid Summer Rape", *Can. J. Plant Sci.*, 68, pp. 509-511.
- Scarth, R., *et al.* (1995), "Apollo Low Linolenic Summer Rape", *Can. J. Plant Sci.*, 75, pp. 203-204.
- Schafer, E.H. (1977), "T'ang", in K.C. Chang (ed.), *Food in Chinese Culture*, Yale Univ. Press, New Haven, pp. 429.

- Scheffler, J.A. and P.J. Dale (1994), "Opportunities for Gene Transfer from Transgenic Oilseed Rape (*Brassica napus*) to Related Species", *Transgen. Res.*, 3, pp. 263-278.
- Scheffler, J.A., *et al.* (1993), "Frequency and Distance of Pollen Dispersal from Transgenic Oilseed Rape (*Brassica napus*)", *Transgen. Res.*, 2, pp. 356-364.
- Scheffler J.A., *et al.* (1995), "Evaluating the Effectiveness of Isolation Distances for Field Plots of Oilseed Rape (*Brassica napus*) Using a Herbicide-resistance Transgene as a Selectable Marker", *Plant Breeding*, 114, pp. 317-21.
- Schiemann, E. (1932), *Entstehung der Kulturpflanzen*, Habd. Vererbwis. Lfg., 15.
- Schlink, S. (1995), "Überdauerungsvermögen und Dormanz von Rapssamen (*Brassica napus* L.) im Boden", *9th European Weed Research Society Symposium*, Budapest, Doorwerth, Netherlands, European Weed Research Society, pp. 65-73.
- Schlink, S. (1998), "10 Year Survival of Rape Seed (*Brassica napus* L.) in Soil", *Zeitschrift für Pflanzenkrankheiten und Pflanzenschutz, Sonderheft*, XVI, pp. 169-172.
- Schröder -Pontoppidan, M., *et al.* (1999), "Very Long Chain and Hydroxylated Fatty Acids in Offspring of Somatic Hybrids between *Brassica napus* and *Lesquerella fendleri*", *Theor. Appl. Genet.*, 99, pp. 108-114.
- Schulz, O.E. (1919), "IV. 105 Cruciferae-Brassicaceae. Part 1. Subtribes I. Brassicinae and II. Raphaninae", in A. Engler (ed.), *Das Pflanzenreich*, Heft 68-70, Wilhelm Engelmann, Leipzig, pp. 1-290.
- Schulz, O.E. (1936), "Cruciferae", in A. Engler and H. Harms (eds.), *Die Natürlichen Pflanzenfamilien*, 2nd ed. 17B, Leipzig, pp. 227-658.
- Scott, S.E. and M.J. Wilkinson, (1998), "Transgene Risk is Low", *Nature*, 393, pp. 320.
- Sharma, T.R. and B.M. Singh, (1992), "Transfer of Resistance to *Alternaria brassicae* in *Brassica juncea* through Interspecific Hybridization among *Brassica*.", *J. Genet. Breed.*, 46, pp. 373-378.
- Sharpe, A.G., *et al.* (1995), "Frequent Nonreciprocal Translocations in the Amphidiploids Genome of Oilseed Rape (*Brassica napus*)", *Genome*, 38, pp. 1112-1121.
- Shiga, T. (1970), "Rapeseed Breeding by Interspecific Crossing between *Brassica napus* and *Brassica campestris* in Japan", *J. A. R. Q.*, 5, pp. 5-10.
- Shpota, V.I. and V.E. Podkolzina (1986), "Development of Erucic Acid-free Forms of Indian Mustard", *Seleksiia I Semenovodstvo*, 4, pp. 19-21.
- Siciliano, S.D. and J.J. Germida (1999), "Taxonomic Diversity of Bacteria Associated with the Roots of Field-grown Transgenic *Brassica napus* cv. Quest, Compared to the Non-transgenic *B. napus* cv. Excel and *B. rapa* Parkland", *FEMS Microbiol.*, 29, pp. 263-272.
- Siemens, J. (2002), "Interspecific Hybridisation between Wild Relatives and *Brassica napus* to Introduce New Resistance Traits into the Oilseed Rape Gene Pool", *Czech J. Genet. Pl. Breed.*, 38, pp. 155-157.
- Siemens, J. and M.D. Sacristán (1995), "Production and Characterization of Somatic Hybrids between *Arabidopsis thaliana* and *Brassica nigra*", *Plant Sci.*, 111, pp. 95-106.
- Sigareva, M.A. and E.D. Earle (1999), "Regeneration of Plants from Protoplasts of *Capsella bursa-pastoris* and Somatic Hybridization with Rapid Cycling *Brassica oleracea*", *Plant Cell Rep.*, 18, pp. 412-417.
- Sigareva, M., *et al.* (1999), "Introgression of Resistance to *Alternaria brassicicola* from *Sinapis alba* to *Brassica oleracea* via Somatic Hybridization and Back-crosses", *Cruciferae Newsl.*, 21, pp. 135-136.
- Simard, M.J., *et al.* (2002), "The Frequency and Persistence of Canola (*Brassica napus*) Volunteers in Québec Cropping Systems", *Weed Technol.*, 16, pp. 433-439.
- Simpson, E.C., *et al.* (1999), "Gene Flow in Genetically Modified Herbicide Tolerant Oilseed Rape (*Brassica napus*) in the UK", in P. J. W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 75-81.
- Sinskaia, E.N. (1928), "The Oleiferous Plants and Root Crops of the Family Cruciferae Bull", *Appl. Bot. Genet. Pl. Breed.*, 19, pp. 1-648.

- Skarzhinskaya, M., *et al.* (1996), “Production of Intertribal Somatic Hybrids between *Brassica napus* L. and *Lesquerella fendleri* (Gray) Wats”, *Theor. Appl. Genet.*, 93, pp. 1242-1250.
- Skarzhinskaya, M., *et al.* (1998), “Genome Organization of *Brassica napus* L. and *Lesquerella fendleri* and Analysis of their Somatic Hybrids Using Genomic *in situ* Hybridization”, *Genome*, 41, pp. 691-701.
- Slocum, M.K. (1989), “Analyzing the Genomic Structure of *Brassica* species Using RFLP Analysis”, in T. Helentjaris and B. Burr (eds.), *Development and Application of Molecular Markers to Problems in Plant Genetics*, Cold Spring Harbor Lab. Press, New York, pp. 73-80.
- Slocum, M.K., *et al.* (1990), “Linkage Arrangement of Restriction Fragment Length Polymorphisms in *Brassica oleracea*”, *Theor. Appl. Genet.*, 80, pp. 57-64.
- Smith, L.B. and G.J. King (2000), “The Distribution of *BoCAL-a* Alleles in *Brassica oleracea* is Consistent with a Genetic Model for Curd Development and Domestication of the Cauliflower”, *Molecular Breeding*, 6, pp. 603-613.
- Snogerup, S. (1980), “The Wild Forms of the *Brassica oleracea* Group (2n = 18) and their Possible Relations to the Cultivated Ones”, in S. T. Hinata and C. Gómez-Campo (eds.), *Brassica Crops and Wild Allies*, Japan Sci. Soc. Press, Tokyo, pp. 121-132.
- Snogerup, S., *et al.* (1990), “*Brassica* Sect. *Brassica* (Brassicaceae). I. Taxonomy and Variation”, *Willdenowia*, 19, pp. 271-365.
- Snow, A.A. and R.B. Jørgensen (1999), “Fitness Costs Associated with Transgenic Glufosinate Tolerance Introgressed from *Brassica napus* subsp. *oleifera* (Oilseed Rape) into Weedy *Brassica rapa*”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 137-142.
- Snow, A.A., *et al.* (1999), “Costs of Transgenic Herbicide Resistance Introgressed from *Brassica napus* into Weedy *B. rapa*”, *Mol Ecol*, 8, pp. 605-615.
- Snow, A.A., *et al.* (2001), “Fitness of Hybrids between Weedy and Cultivated Radish: Implications for Weed Evolution”, *Ecological Applications*, 11, pp. 934-943.
- Snowdon, R., *et al.* (2002), “Identifying the Chromosomes of the A and C Genome Diploid *Brassica* Species *B. rapa* and *B. oleracea* in their Amphidiploid *B. napus*”, *Theor. Appl. Genet.*, 104, pp. 533-538.
- Snowdon, R., *et al.* (2007a), “*Brassica* oilseeds”, in R. J. Singh (ed.), *Genetic Resources, Chromosome Engineering and Crop Improvement*, Vol. 4, CRC Press, Boca Raton, pp. 195-229.
- Snowdon, R., *et al.* (2007b), “Oilseed Rape”, in C. Kole (ed.), *Genome Mapping and Molecular Breeding in Plants*, Vol. 2 Oilseeds, Springer-Verlag, Berlin, pp. 55-114.
- Snowdon, R.J. (2007), “Cytogenetics and Genome Analysis in *Brassica* crops”, *Chromosome Res.*, 15, pp. 85-95.
- Snowdon, R.J. and W. Friedt (2004), “Molecular Markers in *Brassica* Oilseed Breeding: Current Status and Future Possibilities”, *Plant Breed.*, 123, pp. 1-8.
- Snowdon, R.J., *et al.* (2000), “Development and Characterisation of *Brassica napus*-*Sinapis arvensis* Addition Lines Exhibiting Resistance to *Leptosphaeria maculans*”, *Theor. Appl. Genet.*, 101, pp. 1008-1014.
- Snowdon, R.J., *et al.* (2003), “Fishing for Physical Genome Information-*Brassica* Cytogenetics Past, Present and Future”, *Proc 11th Int. Rapeseed Congr.*, Copenhagen, Denmark, 1, pp. 116-119.
- Sobrino-Vesperinas, E. (1988), “Obtainment of Some New Intergeneric Hybrids between Wild Brassicaceae”, *Candollea*, 43, pp. 499-504.
- Song, G.K., *et al.* (1995), “Rapid Genome Change in Synthetic Polyploids of *Brassica* and its Implications for Polyploid Evolution”, *PNAS*, 92, pp. 7719-7723.
- Song, K. and T.C. Osborn (1992), “Polyphyletic Origins of *Brassica napus*: New Evidence Based on Organelle and Nuclear RFLP Analyses”, *Genome*, 35, pp. 992-1001.
- Song, K.M., *et al.* (1988a), “*Brassica* Taxonomy Based on Nuclear Restriction Fragment Length Polymorphisms (RFLPs). I. Genome Evolution of Diploid and Amphidiploid Species”, *Theor. Appl. Genet.*, 75, pp. 784-794.

- Song, K.M., *et al.* (1988b), “Brassica Taxonomy Based on Nuclear Restriction Fragment Length Polymorphisms (RFLPs). 2. Preliminary Analysis of Subspecies within *B. rapa* (syn. *campestris*) and *B. oleracea*”, *Theor. Appl. Genet.*, 76, pp. 593-600.
- Song, K.M., *et al.* (1991), “A Linkage Map of *Brassica rapa* (syn. *campestris*) Based on Restriction Fragment Length Polymorphisms”, *Theor. Appl. Genet.*, 82, pp. 296-304.
- Sparrow, S.D., *et al.* (1990), “Canola Seed Survival over Winter in the Field in Alaska”, *Can. J. Plant Sci.*, 70, pp. 799–807.
- Spect, C.E. and A. Diederichsen (2001), “*Brassica*”, in P. Haelt (ed.), *Mansfield’s Encyclopedia of Agric. and Hort. Crops*, Vol. 3, pp. 1453-2456.
- Squire, G.R., *et al.* (1999), “Gene Flow at the Landscape Level”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 57-64.
- Sridevi, O. and N. Saria (1996), “Reciprocal Hybridization between *Sinapis alba* and *Brassica* Species”, *Cruciferae Newslett. Eucarpia*, 18, pp. 16.
- Srivastava, A., *et al.* (2001), “AFPL Based Genetic Diversity Assessment among Agronomically Important Natural and Some Newly Synthesized Lines of *Brassica juncea*”, *Theor. Appl. Genet.*, 104, pp. 1092-1098.
- Staniland, B.K., *et al.* (2000), “Effectiveness of Border Areas in Confining the Spread of Transgenic *Brassica napus* Pollen”, *Can. J. Pl. Sci.*, 80, pp. 521-526.
- Stanton, R., *et al.* (2002), “Sheep are Potential Vectors for the Spread of Canola (*Brassica napus*) Seed”, *Aust. J. Exp. Agric.*, 43, pp. 535-538.
- Stefansson, B.R. (1983), “The Development of Improved Rapeseed Cultivars”, in J.K.G. Kramer, *et al.* (eds.), *High and Low Erucic Acid Rapeseed Oils*, Academic Press, Toronto, pp. 144-159.
- Stefansson, B.R., *et al.* (1961) “Note on the Isolation of Rape Plants with Seed Oil Free from Erucic Acid”, *Can. J. Plant Sci.*, 47, pp. 281-219.
- Stewart, C.N. Jr., *et al.* (2002), “Gene Flow and its Consequences: *Brassica napus* (Canola, Oilseed Rape) to Wild Relatives”, *Proc. Sci. Methods Workshop: Ecological and Agronomic Consequences of Gene Flow from Transgenic Crops to Wild Relatives*, Ohio State Univ., USA, March, pp.106-112.
- Stiewe, G. and G. Röbbelen (1994), “Establishing Cytoplasmic Male Steility in *Brassica napus* by Mitochondrial Recombination with *B. tournefortii*”, *Plant Breed.*, 113, pp. 294-304.
- Stiewe, G., *et al.* (1995), “Natural and Experimental Evolution of CMS for Rapeseed Breeding”, *Adv. Plant Breeding*, 18, pp. 59-76.
- Stringam, G.R. and R.K. Downey (1982), “Effectiveness of Isolation Distance in Seed Production of Rapeseed (*Brassica napus*)”, *Agron. Abstr.*, pp. 136-137.
- Struss, D., *et al.* (1991), “Development of B-genome Chromosome Addition Lines of *B. napus* Using Different Interspecific *Brassica* Hybrids”, *Plant Breed.*, 106, pp. 209-214.
- Struss, D., *et al.* (1992), “Mapping of Molecular Markers on *Brassica* B-genome Chromosomes Added to *Brassica napus*”, *Plant Breed.*, 108, pp. 320-323.
- Struss, D., *et al.* (1996), “Construction of *Brassica* B Genome Synteny Groups Based on Chromosomes Extracted from Three Different Sources by Phenotype, Isozyme and Molecular Markers”, *Theor. Appl. Genet.*, 93, pp. 1026-1032.
- Sun, W.C., *et al.* (1991), “*Brassica* and *Brassica*-related Oilseed Crops in Gansu, China”, *Proc. 8th Int. Rapeseed Congr.*, Saskatoon, Canada, 4, pp. 1130-1135.
- Sundberg, E. and K. Glimelius (1991), “Effects of Parental Ploidy Level and Genetic Divergence on Chromosome Elimination and Chloroplast Segregation in Somatic Hybrids within Brassicaceae”, *Theor. Appl. Genet.*, 83, pp. 81-88.
- Sweet, J.B. and R. Shepperson (1998), “The Impact of Releases of Genetically Modified Tolerant Oilseed Rape in UK”, *Acta Horticulturae*, 459, pp. 225-234.

- Sweet, J.B., *et al.* (1997), “The Impact of Releases of Genetically Modified Herbicide Tolerant Oilseed Rape in the UK”, *Proc. Brighton Crop Protection Conf. Weeds*, pp. 291-302.
- Sweet, J.B., *et al.* (1999a), “Assisting the Impact and Consequences of the Release and Commercialization of Genetically Modified Crops”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 241-246.
- Sweet, J.B., *et al.* (1999b), “Hybridisation and Persistence in Herbicide Tolerant Oilseed Rape (*Brassica napus*)”, *Proc. 10th Int. Rapeseed Congr.*, Canberra, Australia, pp. 6.
- Szabo, T. (1985), “Variability of Flower, Nectar, Pollen and Seed Production in Some Canadian Canola Varieties”, *Amer. Bee Jour.*, 124, pp. 351-354.
- Takahashi, O. (1987), “Utilization and Seed Production of Hybrid Vegetable Varieties in Japan”, in W.P. Feistritzer and A.F. Kelly (eds.), *Hybrid Production of Selected Cereal, Oilseed and Vegetable Crops*, FAO, Rome, pp. 314-328.
- Takahata, Y. and K. Hinata (1980), “A Variation Study of Subtribe *Brassicinae* by Principle Component Analysis”, in S. Tunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, Tokyo, Japan Sci. Soc. Press, pp. 33-49.
- Takahata, Y. and K. Hinata (1983), “Studies on Cytodemes in Subtribe *Brassicinae* (Cruciferae)”, *Tohoku J. Agri. Res.*, 33, pp. 111 – 124.
- Takahata, Y. and K. Hinata (1986), “Consideration of the Species Relationships in Subtribe *Brassicinae* (Cruciferae) in View of Cluster Analysis of Morphological Characters”, *Plant Species Biol.*, 1, pp. 79 – 88.
- Takayama, S. and A. Isogai (2003), “Molecular Mechanism of Self-recognition in *Brassica* Self-incompatibility”, *J. Exp. Bot.*, 54, pp. 149-156.
- Takayama, S. and A. Isogai (2005), “Self-incompatibility in Plants”, *Ann. Rev. Plant Biol.*, 56, pp. 467-489.
- Takayama, S., *et al.* (2000), “The Pollen Determinant of Self-incompatibility in *Brassica campestris*”, *Nature*, 413, pp. 534-538.
- Takeshita, M., *et al.* (1980), “Application of Ovule Culture to the Production of Intergeneric or Interspecific Hybrids in *Brassica* and *Raphanus*.”, *Jap. J. Genet.*, 55, pp. 373-387.
- Thalmann, C., *et al.* (2001), “Search for Spontaneous Hybridization between Oilseed Rape (*Brassica napus* L.) and Wild Radish (*Raphanus raphanistrum* L.) in Agricultural Zones and Evaluation of the Genetic Diversity of the Wild Species”, *Bot. Helvetica*, 111, pp. 107-119.
- The *Arabidopsis* Genome Initiative (2000), “Analysis of the Genome Sequence of the Flowering Plant *Arabidopsis thaliana*”, *Nature*, 408, pp. 796–815.
- The *B. rapa* Genome Sequencing Project Consortium (2011), “The Genome of the Mesopolyploid Crop Species *Brassica rapa*”, *Nature Genetics*, 43, pp. 1035 - 1039.
- This, P., *et al.* (1990), “Dissection of the *Brassica nigra* Genome by Chromosome Addition Lines”, *Plant Breeding*, 105, pp. 211-220.
- Thomas, P. (1994), *Canola Growers Manual*, Canola Council of Canada.
- Thompson, C.E., *et al.* (1999), “Regional Patterns of Gene Flow and its Consequences for GM Oilseed Rape”, in P.J.W. Lutmann (ed.), *Gene Flow and Agriculture: Relevance for Transgenic Crops*, BCPC Symp. Proc., 72, pp. 95-100.
- Thompson, K.F. (1976), “Cabbages, Kales etc. *Brassica oleracea* (Cruciferae)”, in N.W. Simmonds (ed.), *Evolution of Crop Plants*, Longman, London, pp. 49-52.
- Thompson, K.F. and J.P. Taylor (1966), “Non-linear Dominance Relationships between *S* Alleles”, *Heredity*, 21, pp. 345-362.
- Timmons, A.M., *et al.* (1995), “Assessing the Risks of Wind Pollination from Fields of *Brassica napus* subsp. *oleifera*”, *Euphytica*, 85, pp. 417-423.
- Timmons, A.M., *et al.* (1996), “Risks from Transgenic Crops”, *Nature*, 380, pp. 487.

- Tonguc, M. and P.D. Griffiths (2004), "Transfer of Powdery Mildew Resistance from *Brassica carinata* to *Brassica oleracea* through Embryo Rescue", *Plant Breed.*, 123, pp. 587-589.
- Truco, M.J. and C.F. Quiros (1994), "Structure and Organization of the B Genome Based on a Linkage Map in *Brassica nigra*", *Theor. Appl. Genet.*, 89, pp. 590-598.
- Truco, M.J., *et al.* (1996), "Inter- and Intra-genomic Homology of the *Brassica* Genomes: Implications for their Origin and Evolution", *Theor. Appl. Genet.*, 93, pp. 1225-1233.
- Tsen, M. and S.H. Lee (1942), "A Preliminary Study of Chinese Cultivated Brassicas", *Hortus Sinicus.*, 2, pp. 1-32.
- Tsunoda, S. (1980), "Eco-physiology of Wild and Cultivated Forms in *Brassica* and Allied Genera", in S. Tsunoda, *et al.* (eds.), *Brassica Crops and Wild Allies*, Japan Sci. Soc. Press, Tokyo, pp. 109-120.
- Tsunoda, S. and S. Nishi (1968), "Origin, Differentiation and Breeding of Cultivated *Brassica*", *Proc. XII Int. Congr. Genet.*, 2, pp. 77-88.
- U, N. (1935), "Genome Analysis in *Brassica* with Special Reference to the Experimental Formation of *B. napus* and Peculiar Mode of Fertilization", *Jap. J. Bot.*, 7, pp. 389-452.
- Uchimiya, H. and S.G. Wildman (1978), "Evolution of Fraction I Protein in Relation to Origin of Amphidiploid *Brassica* Species and Other Members of the Cruciferae", *J. Hered.*, 69, pp. 299-303.
- United States Census Bureau, Foreign Trade Statistics, www.census.gov/foreign-trade/, accessed 23 May 2012.
- United Nation Statistics Division, UN comtrade, <http://comtrade.un.org/db/>, accessed 23 May 2012.
- USDA-ARS (2011), Germplasm Resources Information Network - (GRIN) [Online Database], www.ars-grin.gov/
- USDA-Foreign Agricultural Service (2011), Production, Supply and Distribution Online [Online Database], www.fas.usda.gov/psdonline/psdHome.aspx, accessed 10 November 2011.
- Vasilenko, M., *et al.* (2003), "Obtaining and Analysis of Intergeneric Somatic Hybrids between *Brassica napus* and Albino Line of *Orychophragmus violaceus*", *Tristol. Genet.*, 37, pp. 3-10.
- Vavilov, N.I. (1949), "The Origin, Variation, Immunity and Breeding of Cultivated Plants", *Chron. Bot.*, 13, pp. 1-364.
- Vašák, J.V. (2002), "The Role and Effects of Glucosinolates of *Brassica* Species – a Review", *Rostlinná Výroba*, 48, pp. 175-180.
- Vaughan, J.G. (1977), "A Multidisciplinary Study of the Taxonomy and Origin of Brassica Crops", *Bio. Sci.*, 27, pp. 35-40.
- Vaughan, J.H. and J.M. Whitehouse (1971), "Seed Structure and Taxonomy of the Cruciferae", *Bot. J. Linnean Soc.*, 64, pp. 383-409.
- VDI Richtlinien. (2006), "Monitoring the Effects of Genetically Modified Organisms, Pollen Monitoring, Biological Pollen Sampling Using Bee Colonies", VDI 4330 Blatt 4/Part 4.
- VDI Richtlinien. (2007), "Monitoring the Effects of Genetically Modified Organisms, Pollen Monitoring, Technical Pollen Sampling Using Pollen Mass Filter", VDI 4330 Blatt 3/Part 3.
- Vera, C.L., *et al.* (1987), "Detrimental Effects of Volunteer *Brassica* on Production of Certain Cereals and Oilseed Crops", *Can. J. Plant Sci.*, 67, pp. 983-995.
- Verkerk, R., *et al.* (2009), "Glucosinolates in *Brassica* Vegetables: The Influence of the Food Supply Chain on Intake, Bioavailability and Human Health", *Mol. Nutr. Food Res.*, 53, S219-S265.
- Vijayakumar, C.H.M., *et al.* (1994), "An Assay of Basal Branching F2 Segregants from Intra- and Inter-specific Crosses in *Brassica*", *Cruciferae Newslett. Eucarpia*, 16, pp. 25-26.
- Von Ernst, D., *et al.* (1998), "Sicherheitsforschung zu Freisetzungsversuch in Roggenstein (Bayern)", *Bundesgesundheitsblatt*, 12, pp. 525-530 (in German).
- Voss, A., *et al.* (2000), "Intergeneric Transfer of Nematode Resistance from *Raphanus sativus* into the *Brassica napus* Genome", *Acta Hort.*, 539, pp. 129-134.

- Waddington, J. and K.E. Bowren (1978), "Effect of Crop Residue on Production of Barley, Bromegrass and Alfalfa in the Greenhouse and in the Field", *Can. J. Plant Sci.*, 58, pp. 249-225.
- Wang, H., *et al.* (2007a), "Studies on Rapeseed Production and Cultivation Science and Technology in China", *Proc. 12th Int. Rapeseed Congr.*, Wuhan, China, III, pp. 2-7.
- Wang, R., *et al.* (2007b), "Pod Shatter Resistance Evaluation in Cultivars and Breeding Lines of *Brassica napus*, *B. juncea* and *Sinapis alba*", *Plant Breeding*, 126, pp. 588-595.
- Wang, Y.P., *et al.* (2003), "Development of Rapeseed with High Erucic Acid Content by Asymmetric Somatic Hybridization between *Brassica napus* and *Crambe abyssinica*", *Theor. Appl. Genet.*, 106, pp. 1147-1155.
- Wang, Y.P., *et al.* (2004a), "Production and Characterization of Asymmetric Somatic Hybrids between *Brassica napus* and *Crambe abyssinica*", *Votr. Pflanzenzüchtung*, 64, pp. 85-86.
- Wang, X-W. (2010), "The Genome of *Brassica rapa*", Plant and Anim. Genomes XVIII Conference, San Diego, CA. Abstract W104, www.intl-pag.org/18/abstracts/W14_PAGXVIII_104.html.
- Warwick, S.I. (2007), "Gene Flow between GM Crops and Related Species in Canada", in C. Swanton and R. Gulden (eds.), *The First Decade of Herbicide Resistant Crops in Canada*, Vol 4 of Topics in Canadian Weed Science, Canadian Weed Science Soc., Saint-Anne-de Bellevue, QC, Canada, pp. 101-113.
- Warwick, S.I. and I.A. Al-shehbaz (2006), "Brassicaceae: Chromosome Number Index and Database on CD-Rom", *Pl. Syst. Evol.*, 259, pp. 237-248.
- Warwick, S.I. and L.D. Black (1991), "Molecular Systematics of *Brassica* and Allied Genera (Subtribe Brassicinae, Brassiceae) – Chloroplast Genome and Cytodeme Congruence", *Theor. Appl. Genet.*, 82, pp. 839-850.
- Warwick, S. I. and L.D. Black (1993), "Molecular Relationships in the Subtribe Brassicinae (Cruciferae, Tribe, Brassiceae)", *Can. J. Bot.*, 71, pp. 906 - 918.
- Warwick, S. I. and L.D. Black (1997), "Molecular Phylogenies from Theory to Application in Brassica and Allies (Tribe Brassiceae, Cruciferae)", *Opera Bot.*, 132, pp. 159-168.
- Warwick, S.I. and J.C. Hall (2009), "Phylogeny of *Brassica* and Wild Relatives", in S.K. Gupta (ed.), *Biology and Breeding of Crucifers*, CRC Press, Boca Raton, FL, pp. 19-36.
- Warwick, S.I. and C.A. Sauder (2005), "Phylogeny of Tribe Brassiceae (Brassicaceae) Based on Chloroplast Restriction Site Polymorphisms and Nuclear Ribosomal Internal Transcribed Spacer and Chloroplast *trnL* Intron Sequences", *Cand. J. Bot.*, 83, pp. 467-483.
- Warwick, S.I., *et al.* (1992), "Molecular Systematics of *B~ASS~anCdA* Allied Genera (Subtribe Brassicinae, Brassiceae) - Chloroplast DNA Variation in the Genus *Diplotaxis*", *Theor. Appl. Genet.*, 83, pp. 839-850.
- Warwick S.I., *et al.* (1999), "Transgenic Crops: New Weed Problems for Canada?", *Phytoprotection*, 80, pp. 71-84.
- Warwick, S.I., *et al.* (2003), "Hybridization between Transgenic *Brassica napus* L. and its Wild Relatives: *Brassica rapa* L., *Raphanus raphanistrum* L., *Sinapis arvensis* L., and *Erucastrum gallicum* (Willd.) O.E. Schulz", *Theoretical and Applied Genetics*, 107, pp. 528-539.
- Warwick, S.I., *et al.* (2006), "Brassicaceae: Species Checklist and Database on CD-Rom", *Pl. Syst. Evol.*, 259, pp. 249-258.
- Warwick, S.I., *et al.* (2009a), "Gene Flow, Invasiveness, and Ecological Impact of Genetically Modified Crops", *Ann. N. Y. Acad. Sci.*, Vol. 1168, The Year in Evolutionary Biology 2009, pp. 72-99.
- Warwick, S.I., *et al.* (2009b), "Guide to Wild Germplasm: *Brassica* and Allied Crops (Tribe Brassiceae, Brassicaceae)", Third Edition, www.brassica.info/info/publications/guide-wild-germplasm.php, accessed 23 March 2010.
- Watanabe, M. and K. Hinata (1999), "Self-incompatibility", in C. Gómez-Campo(ed.), *Biology of Brassica Coenospecies*", Elsevier Science, pp. 149-183.
- Weekes, R., *et al.* (2005), "Crop-to-crop Gene Flow Using Farm Scale Sites of Oilseed Rape (*Brassica napus*) in the UK", *Transgenic Res.*, 14, pp. 749-759.

- Wei, W. and H. Darmency (2008), "Gene flow hampered by low seed size of hybrids between oilseed rape and five wild relatives", *Seed Sci Research* 18, pp. 115-123.
- Westcott, L. and D. Nelson (2001), "Canola Pollination: an Update", *Bee World*, 82, pp. 115-129.
- Wikipedia contributors, (2008), *Brassica oleracea*, Wikipeddia, The Free Encyclopedia, http://en.wikipedia.org/w/index.php?title=Brassica_oleracea&oldid=196822776 , accessed 12 March 2008.
- Wikipedia contributors, (2011), Computational Phylogenetics, Wikipeddia, The Free Encyclopedia, http://en.wikipedia.org/wiki/Computational_phylogenetics, accessed 27 Oct 2011.
- Wikipedia contributors, (2011), Phylogenetics, Wikipeddia, The Free Encyclopedia, <http://en.wikipedia.org/wiki/Phylogenetics>, accessed 27 Oct 2011.
- Wikipedia contributors, (2011), Triangle of U, Wikipeddia, The Free Encyclopedia, http://en.wikipedia.org/wiki/Triangle_of_U, accessed 17 Nov 2011.
- Wilkinson, M.J., *et al.* (1995), "Problems of Risk Assessment with Genetically Modified Oilseed Rape", *Proc. Brighton Crop Protect. Conf. Weeds*, 3, pp. 1035-1044.
- Wilkinson, M.J., *et al.* (2000), "A Direct Regional Scale Estimate of Transgene Movement from Genetically Modified Oilseed Rape to its Wild Progenitors", *Mol. Ecol.*, 9, pp. 983-991.
- Williams, I., *et al.* (1986), "The Pollination Requirements of Oil-seed Rape", *J. Agric. Sci.*, 106, pp. 27-30.
- Williams, I., *et al.* (1987), "The Effect of Insect Pollination on Plant Development and Seed Production in Winter Oil-seed Rape (*Brassica napus* L.)", *J. Agric. Sci.*, 109, pp. 135-139.
- Williamson, M.H. (1996), *Biological Invasions*, Chapman and Hall, London, pp. 244.
- Winter, H. *et al.* (2003), "Transfer of New Black Leg Resistance into Oilseed Rape", *Proc 11th Int. Rapeseed Congr.*, Copenhagen, Denmark, 6-10 July, 1, pp. 19-21.
- Wu, X., *et al.* (2009), "Genetic Diversity in Oil and Vegetable Mustard (*Brassica juncea*) Landraces by SRAP Markers", *Genet. Resour. Crop Evol.*, 56, pp. 1011-1022.
- Yamagishi, H. and S. Nakagawa (2004), "Somatic Hybrids between *Arabidopsis thaliana* and Cabbage (*Brassica oleracea*)", *J. Japan. Soc. Hort. Sci.*, 73, pp. 319-23.
- Yamagishi, H., *et al.* (2002), "Production of Asymmetric Hybrids between *Arabidopsis thaliana* and *Brassica napus* Utilizing an Efficient Protoplast Culture System", *Theor. Appl. Genet.*, 104, pp. 959-964.
- Yarnell, S.H. (1956), "Cytogenetics of the Vegetable Crops. II. Crucifers", *Botanical Review*, 2, pp. 81-166.
- Yang, Y.W., *et al.* (1999), "Molecular Phylogenetic Studies of *Brassica*, *Rorippa*, *Arabidopsis* and Allied Genera Based on the Internal Transcribed Spacer Region of 18S-25S rDNA", *Mol. Phylogenet. Evol.*, 13, pp. 455-462.
- Yang, Y.W., *et al.* (2002), "A Study of the Phylogeny of *Brassica rapa*, *B. nigra*, *Raphanus sativa* and their Related Genera Using Noncoding Regions of Chloroplast DNA", *Mol. Phylogenet. Evol.*, 23, pp. 268-275.
- Yoshimura, Y., *et al.* (2006), "Transgenic Oilseed Rape along Transportation Routes and Port of Vancouver in Western Canada", *Environ. Biosafety Res.*, 5, pp. 2-9.
- Zhang, X. and S.R. Wessler (2004), "Genome-wide Comparative Analysis of the Transposable Elements in the Related Species *Arabidopsis thaliana* and *Brassica oleracea*", *PNAS*, 101, pp. 5589-5594.
- Zhu, J., *et al.* (1993), "Studies on Resistance to *Phoma lingam* in *Brassica napus*-*Brassica nigra* Addition Lines", *Plant Breed.*, 111, pp. 192-197.

APPENDIX – COMMON PATHOGENS AND PESTS

Table A. 1. Insect, mite and other Brassicaceous crop pests and their regional distribution

ORDER, GENUS AND SPECIES	COMMON NAME	REGIONS AFFECTED
Coleoptera		
<i>Acalymma vittatum</i> (F.)	Striped cucumber beetle	N. America
<i>Agriotes lineatus</i> (L.)	Lined click beetle	N. America, Europe, Russia
<i>Baris laticollis</i> Marsh.	Not reported	Europe
<i>Ceutorhynchus assimillis</i> Payk.	Cabbage seed weevil	N. America, Europe
<i>Ceutorhynchus hepaticus</i> Gyll.	Not reported	Europe
<i>Ceutorhynchus napi</i> Gyll.	Rape stem weevil	Europe
<i>Ceutorhynchus obstricus</i> (Marsh.)	Cabbage seedpod weevil	Europe
<i>Ceutorhynchus pallidactylus</i> (Marsh.)	Cabbage stem weevil	Europe
<i>Ceutorhynchus pleurostigma</i> Marsh.	Turnip gall weevil	Europe, N. Africa, Russia
<i>Ceutorhynchus rapae</i> (Gyll.)	Cabbage curculio	N. America, Europe, Russia
<i>Chaetocnema indica</i> Weise	Not reported	India
<i>Entomoscelis americana</i> Brown	Red turnip beetle	Canada
<i>Listroderes costirostris</i> Schönh.	Vegetable weevil	USA, S. America, Europe, Africa, Asia, Australia, New Zealand
<i>Meligethes aeneus</i> F.	Pollen, rape or blossom beetle	N. America, Europe, N. Africa, Russia, China
<i>Meligethes viridescens</i> (F.)	Pollen or blossom beetle	N. America, Europe
<i>Phyllotreta aerea</i> Allard	Leaf beetle	N. America, Europe, N. Africa, Russia, India
<i>Phyllotreta atra</i> F.	Cabbage flea beetle	Russia
<i>Phyllotreta chotanica</i> Duvivier	Striped flea beetle	India, S.E. Asia
<i>Phyllotreta consobrina</i> (Curtis.)	Turnip flea beetle	No distribution info. found
<i>Phyllotreta cruciferae</i> (Goeze)	Crucifer flea beetle	N. America, Europe, N. Africa, Russia, India
<i>Phyllotreta flexuosa</i> (Ill.)	Not reported	Thailand, Malaysia
<i>Phyllotreta nemorum</i> (L.)	Striped flea beetle	Europe
<i>Phyllotreta striolata</i> (F.)	Cabbage flea beetle	N. America, Europe, Russia, India, Asia
<i>Phyllotreta undulata</i> Kutschera	Lesser striped flea beetle	N. America, Europe, Australia
<i>Psylliodes chrysocephala</i> L.	Cabbage stem flea beetle	Canada, Europe, N. Africa, Russia,
<i>Psylliodes punctulata</i> Melsh.	Hop flea beetles	N. America
Diptera		
<i>Atherigona orientalis</i> Schiner	Pepper fruit fly	USA, Central & S. America, Africa, India, Asia, Australia
<i>Chromatomyia horticola</i> Gour.	Pea leaf miner	Europe, Africa, India, Asia
<i>Contarinia nasturtii</i> (Kief.)	Swede midge	N. America, Europe
<i>Dasineura brassicae</i> (Winn.)	Brassica pod midge	Europe
<i>Delia floralis</i> (Fall.)	Turnip maggot	N. America, Europe, Russia, China, Japan,
<i>Delia radicum</i> (L.)	Cabbage root fly	N. America, Europe, N. Africa, Russia, China
<i>Liriomyza brassicae</i> Riley	Serpentine leaf miner	Worldwide, except Russia
<i>Liriomyza bryoniae</i> Kltb.	Tomato leaf miner	Europe, Russia, India, China, Japan
<i>Phytomyza horticola</i> Gour.	Cruciferous leaf miner	Europe, India, Asia
<i>Phytomyza rufipes</i> Meig.	Cabbage leaf miner	USA, Europe

Homoptera		
<i>Brevicoryne brassicae</i> (L.)	Cabbage aphid	Worldwide
<i>Smynthuodes betae</i> Westw.	Gall-forming aphid, bean root aphid	USA, Europe, Middle East, Australia
Hemiptera		
<i>Aleyrodes proletella</i> (L.)	Cabbage whitefly	Europe
<i>Bagrada hilaris</i> (Burm.)	Painted bug	India, Sri Lanka, Africa, Arabia
<i>Bemisia tabaci</i> (Genn.)	Tobacco whitefly	Worldwide
<i>Eurydema olerace</i> (L.)	Cabbage bug	Turkey, Russia
<i>Eurydema pulchrum</i> (Westw.)	Small cabbage bug	India, Asia
<i>Eurydema rugosum</i> Mots.	Cabbage bug	Russia, China, Japan
<i>Eurydema</i> species	Orange stink or shield bugs	Europe, N. Africa, Russia, India, Asia, Australia
<i>Eurydema ventralis</i> Kolenati	Cabbage bug	Europe, Africa, Russia
<i>Lipaphis erysimi</i> Kltb.	Mustard aphid	Worldwide
<i>Lygus borealis</i> (Kelton)	Not reported	Canada
<i>Lygus elisus</i> Van D.	Pale legume bug	N. America
<i>Lygus hesperus</i> Knight	Western tarnished plant bug	N. America
<i>Lygus lineolaris</i> (P. de B.)	Tarnished plant bug	N. America
<i>Lygus ruqulipennis</i> Popp.	Bishop bug	Canada, Europe, Russia
<i>Murgantia histrionica</i> (Hahn)	Harlequin bug	USA
<i>Myzus persicae</i> Sulz.	<i>Spinach aphid or Green peach aphid</i>	Worldwide
<i>Nysius niger</i> Baker	False chinch bug	India, N. America, Caribbean
<i>Nezara viridula</i> (L.)	Green stink bug	Worldwide
<i>Pemphigus populitransversus</i> Riley	Poplar petiolegall aphid	USA
<i>Pseudococcus calceolariae</i> (Mask.)	Scarlet mealybug	USA, Central & S. America, Europe, Africa, China, Australia, New Zealand
Hymenoptera		
<i>Athalia lugens</i> (Klug)	Mustard sawfly	India
<i>Athalia rosae</i> (L.)	Turnip or cabbage leaf sawfly	Europe, Russia, China, Japan
Lepidoptera		
<i>Acrionicta rumicis</i> (L.)	Knotgrass moth	Europe, Russia, India, China
<i>Agrotis exclamationis</i> L.	Heart and dart moth	Europe, Russia
<i>Agrotis ipsilon</i> (Hufn.)	Black cutworm	Worldwide
<i>Agrotis orthogonia</i> Morr.	Pale western cutworm	Canada
<i>Agrotis segetum</i> D. & S.	Turnip moth	Europe, Africa, India, China, Japan
<i>Argyrogramma signata</i> (F.)	Green semi-looper	India, S. E. Asia
<i>Ascia monuste</i> (L.)	Gulf white cabbage worm	S. America
<i>Autographa californica</i> Speyer	Alfalfa looper	N. America, Malaysia
<i>Autographa gamma</i> (L.)	Silver-Y moth	Europe, N. Africa, India, Asia
<i>Autographa nigrisigna</i> (Wlk.)	Beet worm	Russia, India, China, Japan
<i>Cacoecimorpha pronubana</i> Hbn.	Carnation tortrix	USA, Europe, N. Africa, Japan
<i>Chrysodeixis agnata</i> Stgr.	Three-spotted plusia	China, Japan
<i>Clepsia spectrana</i> (Treit.)	Oblique-banded caterpillar	Europe, Canada
<i>Crocidolomia pavenana</i> (F.)	Large cabbage-heart caterpillar	India, Africa, Asia, Australia
<i>Cydia nigricana</i> F.	Pea moth	Caribbean, Europe, Russia
<i>Diacrisia oblique</i> Wlk.	Jute hairy caterpillar	India, Asia
<i>Elasmopalpus lignosellus</i> (Zell.)	Lesser corn stalk borer	USA, Central America, Thailand
<i>Estigmene acraea</i> Drury	Salt marsh caterpillar	USA, Central America
<i>Euxoa ochrogaster</i> (Gn.)	Red-barred cutworm	N. America
<i>†Evergestis forficalis</i> L.	Crucifer caterpillar	Europe, India, Japan
<i>Evergestis rimosalis</i> (Gn.)	Cross striped cabbageworm	N. America
<i>Hadula trifolii</i> (Hufn.)	Clover cutworm	N. America, Europe, Africa, Russia, India, China

<i>Helicoverpa armigera</i> (Hbn.)	Cotton bollworm	Europe, Africa, India, Russia, S. E. Asia, Australia, New Zealand
<i>Hellula phidylealis</i> (Wlk.)	Cabbage budworm	Central America
<i>Hellula undalis</i> (F.)	Cabbage webworm	Europe, Africa, India, Asia, Australia, New Zealand
<i>Lacanobia oleracea</i> (L.)	Bright-line brown-eye moth	Europe
<i>Lacanobia suasa</i> D. & S.	Not reported	Europe, Russia
<i>Loxostege sticticalis</i> L.	Beet webworm	N. America, Asia, Europe, Russia
<i>Mamestra brassicae</i> (L.)	Cabbage moth	Europe, Russia, India, Asia
<i>Mamestra configurata</i> Wlk.	Bertha armyworm	N. & Central America
<i>Noctua pronuba</i> (L.)	Common yellow underwing moth	Europe
<i>Ochropleura flammata</i> D. & S.	Indian cutworm	India
<i>Peridroma saucia</i> (Hbn.)	Pearly underwing moth	The Americas, Europe, India, China, Japan
<i>Pieris brassicae</i> (L.)	Cabbage caterpillar	S. America, Europe, Russia, India, China, Japan, Africa
<i>Pieris canidia</i> (Sparman)	Small cabbage butterfly	China, S.E. Asia
<i>Pieris napi</i> (L.)	Green-veined white butterfly	Europe, N. Africa, Russia, India, China, Japan,
<i>Pieris rapae</i> L.	Imported cabbageworm or cabbage white butterfly	N. & Central, America, Europe, N. Africa, Russia, India, Asia, Australia, New Zealand
<i>Plutella xylostella</i> L.	Diamondback moth	Worldwide
<i>Pontia daplidice</i> (L.)	Not reported	Russia
<i>Spodoptera exigua</i> (Hbn.)	Beet armyworm	N. & Central America, Europe Africa, Russia, India, Asia, Australia
<i>Spodoptera frugiperda</i> J. E. Smith	Fall armyworm	The Americas
<i>Spodoptera littoralis</i> (Bdv.)	Cotton leafworm	Africa, Middle East
<i>Trichoplusia ni</i> (Hbn.)	Cabbage looper	Worldwide, except Australia & New Zealand
<i>Vanessa cardui</i> L.	Painted lady butterfly	N. America, Europe, Africa, Russia, Australia
<i>Xestia c-nigrum</i> (L.)	Spotted cutworm	N. & Central America, Europe, Russia, India, Asia
Acari		
<i>Halotydeus destructor</i> (Tucker)	Redlegged earth mite	Australia, New Zealand, S. Africa
<i>Tyrophagus putrescentiae</i> (Schr.)	Cereal mite	USA, Central & S. America, Europe, Africa, India, China
Stylommatomophora		
<i>Arion lusitanicus</i> Mabilie	Spanish slug	Europe
<i>Deroceas reticulatum</i> Müll	Grey field slug	N. America, Europe, Russia, Australia, New Zealand
<i>Lissachatina fulica</i> (Bowdich)	Giant African land snail	S. America, Africa, India, S. E. Asia
Thysanoptera		
<i>Thrips tabaci</i> Lind.	Onion thrips	Worldwide
Tylenchida		
<i>Meloidogyne ethiopica</i> Whitehead	Not reported	S. America, Europe, Africa
<i>Meloidogyne graminicola</i> Golden & Birchfield	Rice root knot nematode	USA, S. America, S. Africa, India, S. E. Asia
<i>Pratylenchus neglectus</i> (Rensch) Filipjev & Stekhoven	California meadow nematode	Europe, Russia, Pakistan

Source: Information drawn from CAB International Crop Protection Compendium, Bonnemaison (1965), Lamb (1989), Thomas (1994) and R. Elliott and J. Soroka, personal communication

Table A.2. Diseases of Rapeseed = Canola [*B. napus* L. and *Brassica rapa* L. (= *B. campestris* L.)]

COMMON NAME(S)	SCIENTIFIC NAME (AND SYNONYMS)
BACTERIAL DISEASES	
Bacterial black rot	<i>Xanthomonas campestris</i> pv. <i>campestris</i> (Pammel 1895) Dowson 1939 = <i>Xanthomonas campestris</i> pv. <i>raphani</i> = <i>Xanthomonas campestris</i> pv. <i>aberrans</i>
Bacterial leaf spot	<i>Xanthomonas campestris</i> pv. <i>armoraciae</i> (McCulloch 1929) Dye 1978
Bacterial pod rot*	<i>Pseudomonas syringae</i> pv. <i>maculicola</i> (McCulloch 1911) Young, Dye and Wilkie 1978) (Canada, UK)
Bacterial soft rot	<i>Erwinia carotovora</i> (Jones 1901) Bergey <i>et al.</i> 1923 <i>Pseudomonas marginalis</i> pv. <i>marginalis</i> (Brown 1918) Stevens 1925
Scab	<i>Streptomyces</i> spp. <i>Streptomyces scabiei</i> corrig. (ex Thaxter 1891) Lambert and Loria 1989 = <i>Streptomyces scabies</i> (Thaxter 1891) Waksman and Henrici 1948
Crown gall	<i>Agrobacterium tumefaciens</i> (Smith and Townsend 1907) Conn 1942
FUNGAL DISEASES	
Alternaria black spot = Dark pod spot (UK)	<i>Alternaria brassicae</i> (Berk.) Sacc. <i>A. brassicicola</i> (Schwein.) Wiltshire <i>A. japonica</i> H. Yoshii = <i>A. raphani</i> Groves and Skolko
Anthracnose	<i>Colletotrichum gloeosporioides</i> (Penz.) Penz. and Sacc. in Penz. <i>Glomerella cingulata</i> (Stoneman) Spauld. and H. Schrenk [teleomorph] <i>C. higginsianum</i> Sacc. in Higgins
Black leg = stem canker (UK)	<i>Leptosphaeria maculans</i> (Desmaz.) Ces. and De Not <i>Phoma lingam</i> (Tode:Fr.) Desmaz. [anamorph]
Black mold rot	<i>Rhizopus stolonifer</i> (Ehrens.:Fr.) Vuill.
Black root	<i>Aphanomyces raphani</i> Kendrick
Brown girdling root rot*	<i>Rhizoctonia solani</i> Kühn (Canada) <i>Thanatephorus cucumeris</i> (A.B. Frank) Donk [teleomorph]
Cercospora leaf spot	<i>Cercospora brassicicola</i> Henn.
Clubroot	<i>Plasmodiophora brassicae</i> Woronin
Downy mildew	<i>Peronospora parasitica</i> (Pers.:Fr.) Fr.
Fusarium wilt	<i>Fusarium oxysporum</i> Schlechtend.:Fr. f. sp. <i>conglutinans</i> (Wollenweb.) W.C. Snyder and H.N. Hans.
Gray mold	<i>Botrytis cinerea</i> Pers.:Fr. <i>Botryotinia fuckeliana</i> (de Bary) Whetzel [teleomorph]
Head rot	<i>Rhizoctonia solani</i> Kühn <i>Thanatephorus cucumeris</i> (A.B. Frank) Donk [teleomorph]
Leaf spot*	<i>Alternaria alternata</i> (Fr.:Fr.) Keissl. (Canada) <i>Ascochyta</i> spp. (former USSR)
Light leaf spot	<i>Pyrenopeziza brassicae</i> Sutton and Rawlinson in Rawlinson <i>et al.</i> (1978). <i>Cylindrosporium concentricum</i> Grev. [anamorph]
Pod rot*	<i>Alternaria alternata</i> (Fr.:Fr.) Keissl. (Canada) <i>Cladosporium</i> sp.
Powdery mildew	<i>Erysiphe polygoni</i> DC. <i>E. cruciferarum</i> Opiz ex Junell.
Ring spot	<i>Mycosphaerella brassicicola</i> (Duby) Lindau in Engl. and Prantl

	<i>Asteromella brassica</i> (Chev.) Boerema and Van Kesteren [anamorph]
Root rot	<i>Alternaria alternata</i> (Fr.:Fr.) Keissl. <i>Fusarium</i> spp. <i>Macrophomina phaseolina</i> (Tassi) Goidanich <i>Phymatotrichopsis omnivora</i> (Duggar) Hennebert <i>Phytophthora megasperma</i> Drechs. <i>Pythium debaryanum</i> Auct. non R. Hesse <i>P. irregulare</i> Buisman <i>Rhizoctonia solani</i> Kühn <i>Thanatephorus cucumeris</i> (A.B. Frank) Donk [teleomorph] <i>Sclerotium rolfsii</i> Sacc. <i>Athelia rolfsii</i> (Curzi) Tu and Kimbrough [teleomorph]
Sclerotinia stem rot	<i>Sclerotinia sclerotiorum</i> (Lib.) de Bary
Seed rot, damping-off	<i>Alternaria</i> spp. <i>Fusarium</i> spp. <i>Gliocladium roseum</i> (Link) Bainier <i>Nectria ochroleuca</i> (Schwein.) Berk [teleomorph] <i>Pythium</i> spp. <i>Rhizoctonia solani</i> Kühn <i>Thanatephorus cucumeris</i> (A.B. Frank) Donk [teleomorph] <i>Rhizopus stolonifer</i> (Ehrenb.:Fr) Vuill. <i>Sclerotium rolfsii</i> Sacc.
Root gall smut*	<i>Urocystis brassicae</i> Mundkur (China, India)
Southern blight (leaf, root and seed rot)	<i>Sclerotium rolfsii</i> Sacc.
Verticillium wilt*	<i>Verticillium longisporum</i> (comb. Nov. Karapappa <i>et al.</i>) (Europe)
White blight*	<i>Rhizoctonia solani</i> Kühn (India) <i>Thanatephorus cucumeris</i> (A.B. Frank) Donk [teleomorph]
White leaf spot = grey stem (Canada)	<i>Pseudocercospora capsellae</i> (Ellis and Everh.) Deighton = <i>Cercospora brassicae</i> (Faitrey and Roum.) Höhn. <i>Mycosphaerella capsellae</i> (Inman and Sivansen) [teleomorph]
White rust = staghead	<i>Albugo candida</i> (Pers.) Kunze = <i>A. cruciferarum</i> (DC.) S.F. Gray (<i>Peronospora</i> sp. commonly present in staghead phase)
Yellows	<i>Fusarium oxysporum</i> Schlechtend.:Fr.
NEMATODES, PARASITIC	
Cyst nematode	<i>Heterodera cruciferae</i> Franklin <i>H. schachtii</i> Schmidt
Lesion nematode	<i>Pratylenchus</i> spp. <i>P. pratensis</i> (de Man) Filipjev
Root-knot nematode	<i>Meloidogyne</i> spp.
VIRAL DISEASES	
Crinkle*	genus <i>Carmovirus</i> , <i>Turnip crinkle virus</i> (TCV) (Yugoslavia)
Mosaic	genus <i>Caulimovirus</i> , <i>Cauliflower mosaic virus</i> (CaMV) genus <i>Cucumovirus</i> , <i>Cucumber mosaic virus</i> * (CMV) (Hungary) genus <i>Comovirus</i> , <i>Radish mosaic virus</i> (RaMV) genus <i>Potyvirus</i> , <i>Turnip mosaic virus</i> (TuMV)
Yellows	genus <i>Luteovirus</i> , <i>Beet western yellows virus</i> (BWYV) genus <i>Cytorhabdovirus</i> , <i>Broccoli necrotic yellows virus</i> * (BNYV)

PHYTOPLASMAL DISEASES	
Aster yellows and phyllody	Aster yellows phytoplasma
MISCELLANEOUS DISEASES and DISORDERS	
Autogenic necrosis	Genetic disorder
Black speck	Physiological
Sulfur deficiency	Sulfur deficiency
Tipburn	Calcium deficiency

Source: Reproduced from the American Phytopathological Society listing of known *Brassica* pathogens and disorders at www.apsnet.org/publications/commonnames/Pages/Rapeseed.aspx. Last update 3/8/2001.

* Not known to occur naturally in the USA.